

**ANTISENSE OLIGONUCLEOTIDE WHICH INHIBITS EXPRESSION OF THE
OB-RGRP PROTEIN AND METHOD FOR DETECTING COMPOUNDS WHICH
MODIFY THE INTERACTION BETWEEN PROTEINS OF THE OB-RGRP
FAMILY AND THE LEPTIN RECEPTOR**

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The present application relates to antisense oligonucleotides which inhibit expression of the OB-RGRP protein and to uses thereof for preventing and/or treating leptin-related pathological conditions.

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It also relates to a method for detecting leptin receptor ligands using the energy transfer between, firstly, fusion proteins composed of leptin receptors and of energy-donor or -acceptor proteins and, secondly, fusion proteins composed of OB-RGRP or of MYO47 and of energy-donor or -acceptor proteins.

It also relates to fusion proteins for implementing this method.

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Leptin is a 16 kDa protein secreted mainly by the adipose tissue, which binds to a receptor (OB-R) belonging to the cytokine receptor family. Five membrane-bound isoforms of this receptor have been identified, and derive from alternative splicing of the same gene. These isoforms which have the same extracellular and transmembrane domain are characterized by

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intracellular domains of varying sizes (Tartaglia et al (1995) Cell 83, 1263-1271). A soluble form of the receptor has also been identified and comes from an alternative splicing or a proteolytic cleavage of the extracellular domain of the membrane-bound forms. The short form of the receptor (OB-Rs), which appears to be involved in transporting leptin across the blood-brain barrier, is the most expressed isoform. The long form (OB-RI) is only expressed in a few

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tissues, such as the hypothalamus, and appears to be responsible for most of the biological effects of leptin (Sweeney, G. (2002) Cell Signal 14, 655-663). Leptin and its receptor have been the subject of particular attention due to their involvement in the regulation of energy balance and of the metabolism, and in the neuroendocrine response to food intake. Recently, it has been shown that leptin is also involved in important addition functions, such as regulation of the bone mass, angiogenesis, thrombus formation, sexual maturation, hematopoiesis, the regulation of immunity and inflammation, fetal development and cancer. The administration of leptin to leptin-deficient

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organisms such as mice (ob/ob) and certain humans causes a decrease in the lipid mass in various tissues, such as the liver and the adipose tissue (Halaas et al. (1995) Science 269, 543-546, Pelleymounter et al. (1995) Science 269, 540-543, Campfield et al. (1995) Science 269, 546-549, Farooqi et al. (1999) N Engl J Med 341, 879-884). This treatment with leptin also improves the sensitivity to insulin and decreases the fatty mass in mice and humans exhibiting lipodistrophy (Shimomura et al. (1999) Nature 401, 73-76, Oral et al. (2002) New England Journal of Medicine 346, 570-578, Petersen et al. (2002) J Clin Invest 109, 1345-1350. Obese individuals are generally resistant to leptin. The reasons for this resistance are still poorly understood, but several mechanisms have been suggested: a deficiency in leptin transport across the blood-brain barrier, a deficiency in activation of OB-R or in the signaling by these receptors, and the overexpression of negative regulators such as SOCS3 and PTP-1B (Bjorbaek et al. (2000) J Biol Chem 275, 40649-40657, Cheng et al. (2002) Developmental Cell 2, 497-503, Cook and Unger

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(2002) Developmental Cell 2, 385-387. Understanding the mechanisms of resistance to leptin requires a more detailed characterization of the mechanisms involved in OB-R activation.

OB-R is constitutively associated with janus kinase 2 (JAK 2). The binding of JAK2 to the receptor is critical for the signaling by OB-R and has been proposed as being involved in stabilizing the OB-R receptor dimers. Activation by agonists is thought to cause a change in conformation in the juxtamembrane region of the cytoplasm tail of the OB-R. JAK2, which is constitutively linked to the box1 motif in this region, is activated by autophosphorylation and then phosphorylates the OB-R receptor but not the OB-Rs receptor. The phosphorylation of OB-R allows anchoring of STAT proteins, which bind to the receptor and are activated by phosphorylation of tyrosine. The activated STAT proteins dimerize and translocate into the nucleus in order to stimulate the transcription of genes via STAT response elements Tartaglia (1997) J Biol Chem 272, 6093-6096.

Recently, a second promoter for the leptin receptor has been discovered. Interestingly, a second transcript is co-expressed with the OB-R messengers from this promoter. This transcript has been observed in several species, such as mice, rats, humans, yeast and *C. elegans*, Bailleul et al. (1997) Nucleic Acids Res 25, 2752-2758. *In situ* hybridization experiments confirm the coexpression of OB-R and of the associated gene in the brain of mice, including the hypothalamic regions involved in regulating body weight (Mercer et al., J Neuroendocrinol 2000 July; 12(7):649-55). The corresponding protein is composed of 131 amino acids and is called OB-R-gene related protein (OB-RGRP). This protein was the subject of patent application WO 98/05792.

The fact that OB-RGRP is expressed in yeast and nematodes, which are organisms lacking leptin receptors, indicates a more general role for OB-RGRP, supported by the deletion of this protein in yeast which causes a deficiency in transport of proteins from the golgi to the vacuoles (Belgareh-Touze et al. (2002) Molecular Biology Of The Cell 13, 1694-1708).

In 2002, a cDNA called MY047 was cloned from a human brain cDNA library (16). The function of the corresponding protein is still unknown. MY047 exhibits 68% homology with OB-RGRP, suggesting that these two proteins belong to the same family. Analysis of the sequences available for the human genome sequencing project shows that no other homolog exists.

The applicants have endeavored to determine the role of OB-RGRP and its relationships with leptin receptors.

They have thus shown the specificity of the interactions between OB-RGRP and the OBRs receptor.

They have also shown that it is possible to specifically modify the expression of leptin receptors at the cell surface using antisense oligonucleotides directed against the leptin receptor gene associated protein (OB-RGRP).

A subject of the present application is therefore optionally modified oligonucleotides comprising from 8 to 50 nucleotides which hybridize

specifically with the sequence SEQ ID No. 1 and which inhibit OB-RGRP expression.

Advantageously, these oligonucleotides promote the expression of leptin receptors at the cell surface.

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Preferentially, these oligonucleotides are antisense oligonucleotides.

Preferentially, these oligonucleotides comprise a sequence exhibiting at least 60%, 70%, 80% or 90% identity with the sequence SEQ ID No. 2.

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According to an advantageous embodiment, in these oligonucleotides, nucleotides are thioesterified.

According to another advantageous embodiment, in these oligonucleotides, nucleotides are 2'-O-methylated.

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According to another advantageous embodiment, these oligonucleotides have a triethylene glycol residue at their 3' ends.

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Although the most commonly used form of antisense compounds is in the form of antisense oligonucleotides, the present invention includes oligonucleotide derivatives and compounds which mimic their structure, such as those described hereinafter, without this list being limiting. The antisense compounds in agreement with this invention preferably comprise from 8 to 50 nucleobases (i.e. they are oligomers made up of 8 to 50 nucleotide units). The antisense compounds particularly targeted are antisense oligonucleotides, more specially those which are made up of approximately 12 to 30 nucleobases. The antisense compounds comprise ribozymes, oligozymes or other short catalytic RNAs or catalytic oligonucleotides which hybridize with the target nucleic acid and modulate its expression. A nucleoside is a combination of a nitrogenous base and a sugar. The base of a nucleoside is generally a heterocyclic nitrogenous base. The two most common types of heterocyclic base are purine and pyrimidine bases. The nucleotides are nucleosides which carry a phosphate group covalently bonded to the sugar of the nucleoside. For the nucleosides comprising a pentanofuranose, the phosphate may be bonded to the hydroxyl at position 2', 3' or 5' of the sugar. The formation of nucleotides comes from the covalent attachment of the phosphate group to two adjacent nucleosides, which makes it possible, step by step, to obtain a linear oligomer. The two ends of such a linear polymer can, in turn, join together to form a circular structure, but the open structure is generally preferred. In the nucleotide structure, the phosphate groups are considered to form the internucleoside skeleton of the oligonucleotide. The normal bond in the RNA or DNA skeleton is a 3'-5' phosphodiester linkage. Specific examples of antisense compounds which can be used in this invention include oligonucleotides containing a modified backbone or unnatural internucleoside bonds. Thus, oligonucleotides with a modified backbone comprise those which conserve a phosphate atom in their skeleton and those which are lacking therein. For the needs of the present invention, modified oligonucleotides which do not have a phosphorus atom in their internucleoside bond can, nevertheless, be considered to be oligonucleotides. The backbone of these modified oligonucleotides may comprise, for example,

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the following groups: phosphorothioates, chiral phosphorothioates, phosphorodithioates, phosphotriesters, aminoalkylphosphotriesters, methyl and other alkyl phosphonates, including 3'-alkylene phosphonates, 5'-alkylene phosphonates and chiral phosphonates, phosphinates, phosphoramidates, including 3'-aminophosphoramidate and aminoalkylphosphoramidates, thionophosphoramidates, thionoalkylphosphonates, thionoalkylphosphotriesters, selenophosphates and borophosphates which form normal 3'-5' bonds, and analogs thereof which form 2'-5' bonds, and also those which exhibit a reverse polarity, i.e. comprising at least one internucleoside bond of the 3'-3', 5'-5' or 2'-2' type. The form of oligonucleotides having a reverse polarity which is preferentially used is that which has the first internucleoside bond in 3' is of the 3'-3' type. This corresponds to a single inverted nucleotide residue which may, moreover, be abasic, i.e. in which the heterocyclic nitrogenous base is missing or replaced with a hydroxyl group. The various forms (saline or free acid) are included in the field of this invention.

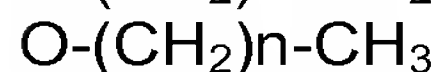
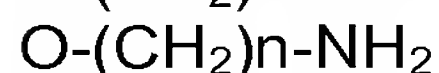
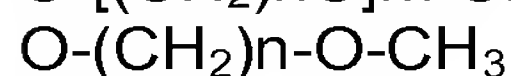
The backbone of the modified oligonucleotides lacking a phosphorus atom is preferentially made up of short alkyl or cycloalkyl chains, including derivatives thereof comprising one or more hetero atoms, acting as an internucleoside bond. This type of backbone may be based on a morpholino bond (partly consisting of the sugar of the nucleoside), on siloxane, on formacetyl and thioformacetyl, on methylene formacetyl and methylene thioformacetyl, on riboacetyl, on alkenes, on sulfamates, on sulfonate and sulfonamide, on methyleneimine and methylene hydrazine, on amide, and on any other group comprising various nitrogen, sulfur and oxygen atoms or methyl groups.

For other oligonucleotide analogs, the sugar and the internucleoside bond (i.e. the backbone) are replaced at the same time in the nucleotide structure with new groups. The heterocyclic nitrogenous base is conserved in order to ensure hybridization with the target nucleic acid. Such oligomeric compounds, PNAs (for Peptide Nucleic Acids), have shown an excellent capacity for hybridization. In these compounds, the skeleton of the oligonucleotide is replaced with an amide-based backbone, in particular with aminoethyl glycine, grafted directly or indirectly onto the nitrogenous bases. In addition, thorough teaching regarding these PNAs may be found in Nielsen et al., Science, 1991, 254, 1497.

The invention incorporates more particularly oligonucleotides with a phosphorothioate, amide and morpholine backbone, and the oligonucleotides with a hetero atom skeleton, more precisely:

-CH₂-NH-O-CH₂-
 -CH₂-N(CH₃)-O-CH₂- (called methylene(methylimino) or MMI skeleton)
 -CH₂-O-N(CH₃)-CH₂-
 -CH₂-N(CH₃)-N(CH₃)-CH₂-
 -O-N(CH₃)-CH₂-CH₂- (in which the phosphodiester bridge is: O-P-O-CH₂).

The modification of the oligonucleotides may also be carried on the sugars: the preferred substitutions are at position 2' (F; O-, N- or S-alkane, O-, N- or S-alkene or O-, N- or S-alkyne derivatives of length C1 to C11, which may or may not be substituted) in particular, the preferred derivatives are:



in which n and m range from 1 to 10.

Other modifications of the 2' position include the following groups: aliphatic chains, which may or may not be substituted, of length C1 to C10, aryl chains, aryl-alkyl chains and alkyl-aryl chains; -SH, -SCH₃, -OCN, -Cl, -Br, -CN, CF₃, -OCF₃, -SO₂CH₃, -ONO₂, -NO₂, -N₃, -NH₂; substituted silyls; "reporter" groups; intercalating groups; RNA cleavage groups; group to improve the pharmacodynamic capacities of an oligonucleotide. The preferred modifications include the groups:

2'-methoxyethoxy (2'-O-CH₂CH₂OCH₃, also called 2'-O-(2-methoxyethyl) or 2'-MOE) (Martin et al., *Helv. Chim. Acta*, 1995, 78f 486-504) 2'-dimethylaminoethoxy (O(CH₂)₂ON(CH₃)₂, also called 2'-DMAOE;

2'-dimethylaminoethoxyethoxy (2'-O-CH₂OCH₂-N(CH₂)₂, also called 2'-dimethylaminoethoxyethyl or 2'-DMAEOE).

Another advantageous modification leads to the formation of LNAs (Locked Nucleic Acids) in which the hydroxyl at position 2' is attached to the carbon at position 3' or 4' of the sugar, then forming a sugar with a bicyclic structure. The preferred bridging occurs via a methyl or ethyl linkage between the 2' oxygen and the 4' carbon.

Other preferred substitutions at position 2' include:

-O-CH₃ (2'-methoxy)

-O-(CH₂)₃-NH₂ (2'-aminopropoxy)

-CH₂-CH=CH₂ (2'-allyl)

-O-CH₂-CH=CH₂ (2'-O-allyl)

-F (2'-fluoro).

These modifications at 2' may be in the ribo (lower) or arabino (upper) position. The 2'-fluoro substituent is the preferred one in the arabino position.

Similar modifications may be made on other positions, in particular at position 3' of the sugar of the nucleotide at the 3'-terminal end or in the oligonucleotides with a 2'-5' backbone, and at position 5' of the sugar at the 5'-terminal end. The sugars of the oligonucleotides can also be replaced with analogs (for example a cyclobutyl can be substituted for a pentofuranyl).

The oligonucleotides can also comprise modifications or substitutions on the nucleobases (nitrogenous heterocyclic bases called "bases" by those skilled in the art). The natural (unmodified) bases are purines (adenine A and guanine G) and pyrimidines (cytosine C, thymine T and uracil U). Included among the modified bases are natural or synthetic molecules such as 5-methylcytosine, 5-hydroxymethylcytosine, xanthine, hypoxanthine, 2-aminoadenine; 6-methyl, 2-methyl and other alkyl derivatives of purine bases (A and G); 2-thio derivative (C, T and U); 5-halo derivative (U, C); 5-propynyl cytosine derivative (U and C); 6-azo derivative (U, T and C); 5-uracil; 4-thiouracil; 8-halo, 8-amino, 8-thiol, 8-thioalkyl, 8-hydroxyl and other adenines and guanine substituted at position 8; 5-halo (in particular 5-bromo), 5-trifluoromethyl and other uracils and cytosines substituted in

position 5; 7-methylguanine and 7-methyladenine; 2-fluoroadenine; 2-aminoadenine; 8-azaguanine and 8-azaadenine; 7-deazaguanine and 7-deazaadenine; 3-deazaguanine and 3-deazaadenine. In the other modified bases, tricyclic pyrimidines are found such as phenoxazine cytidine (1H-pyrimido[5,4-b][1,4]benzoxazin-2-(3H)-one), phenothiazine cytidine (1H-pyrimido[5,4-b][1,4]benzothiazin-2(3H)-one), substituted phenoxazine cytidine (such as 9-(2-aminoethoxy)-H-pyrimido[5,4-b][1,4]benzoxazine-2(3H)-one), or carbazole cytidine (2H-pyrimido[4,5-b]indol-'2-one).

The modified bases comprise the compounds in which the purine or pyrimidine heterocycle is replaced with another heterocycle, for example 7-deazaadenine, 7-deazaguanosine, 2-aminopyridine or 2-pyridone (The Concise Encyclopedia Of Polymer Science And Engineering, pages 858-859, Kroschwitz, J.I., ed. John Wiley & Sons, 1990; Englisch et al., Angewandte Chemie, International Edition, 1991, 30, 613; Sanghvi, Y.S., Chapter 15, Antisense Research and Applications, pages 289-302, Crooke, S.T. and Lebleu, B., ed., CRC Press, 1993). Some of these modified bases may be of great value for increasing the affinity of the oligomeric compounds of the invention, such as pyrimidines substituted at position 5, azapyrimidines, or N- and O-substituted purines (such as 2-aminopropyladenine, 5-propynyl uracil, 5-propynyl cytosine). The substituted 5-methylcytosines have a positive effect on the stability of oligomer-nucleic acid duplexes (Sanghvi, Y.S., Crooke, S.T. and Lebleu, B., eds., Antisense Research and Applications, CRC Press, Boca Raton, 1993, pp. 276-278) and are the preferred substitution, in particular in combination with 2'-methoxyethyl modifications of the sugars.

Preferentially, these oligonucleotides are in the single-stranded form.

According to a particularly advantageous embodiment, these oligonucleotides comprise a sequence exhibiting at least 60% identity with the sequence SEQ ID No. 2, in which the nucleotides at positions 2, 4, 6, 7, 9, 11, 13, 15, 17, 19 and 20, in the 5' to 3' direction, are thioesterified.

According to a particularly advantageous embodiment, these oligonucleotides comprise a sequence exhibiting at least 60% identity with the sequence SEQ ID No. 2, in which the nucleotides at positions 1, 2, 3, 4, 5, 16, 17, 18, 19 and/or 20, in the 5' to 3' direction, are 2'-O-methylated.

Preferentially, the oligonucleotides according to the present invention are DNAs.

A subject of the present invention is also oligonucleotides of the iRNA (Interfering Ribonucleic Acid) type comprising from 15 to 25 nucleotides, which hybridize specifically to the sequence SEQ ID No. 21 and which inhibit the expression of OB-RGRP.

Preferentially, such iRNAs comprise 17 or 19 nucleotides taken continuously from the sequence SEQ ID No. 21, or from the sequence complementary thereto. Nucleotides A(A/G) and (C/T)T can be added respectively in 5' and in 3' of this sequence of 17 or 19 nucleotides. Other types of residues or

chemical groups can, however, be added to these two ends, provided that they do not decrease the activity of the antisenses.

The nucleotide modifications described for the antisenses are also possible for those making up the composition of the siRNAs.

5 The present invention also includes any modifications of the antisenses or of the iRNAs which are directed toward increasing the resistance of these compounds to cellular nucleases, or their penetration into cells and/or their effectiveness in targeting the OB-RGRP sequence.

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When they are DNAs, the oligonucleotides according to the present invention can be produced conveniently and routinely by the well-known technique of solid-phase synthesis. The equipment for such synthesis is sold by various specialized companies, such as Applied Biosystems (Foster City, CA). The synthesis of the
15 antisenses in the present invention makes use of chemical synthesis on a suitable support according to methods known to those skilled in the art, in particular described by E. Uhlmann, A. Peyman, A. Rytte, A. Schmidt and E. Buddecke (1999, Methods in Enzymology 313: 268-284) and by E. Uhlmann (Recent advances in the medicinal chemistry of antisense oligonucleotides, Current
20 Opinion of Drug Discovery and Development 3: 203-213, 2000). Any other method of synthesis known to those skilled in the art may also be used.

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When they are iRNAs, the oligonucleotides according to the present invention can be synthesized by chemical synthesis, when they are synthetic iRNAs, or expressed in situ using vectors expressing such oligonucleotides.

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siRNAs (small iRNAs) can be obtained from various suppliers, such as Proligo (Proligo France SAS 1 rue Robert et Sonia Delaunay 75011 Paris) Dharmacon (Dharmacon, Inc. 1376 Miners Drive #101 Lafayette, CO 80026) and Ambion (Ambion (Europe) Ltd. Ermine Business Park Spitfire Close Huntingdon, Cambridgeshire PE29 6XY United Kingdom), or can be synthesized using kits
30 marketed by various companies, such as Dharmacon and Ambion.

Preferentially, the iRNAs according to the present invention are in double-stranded form.

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After synthesis, the iRNAs are first of all taken up in RNase-free water. The pairing of the two single-stranded molecules can be carried out as follows: 20 $\mu\text{mol.L}^{-1}$ of each strand are mixed in the pairing buffer (100 mmol.L^{-1} of potassium acetate, 30 mmol.L^{-1} of HEPES-KOH, pH 7.4, 2 mmol.L^{-1} of magnesium acetate) and then heated at 90°C for 1 min, followed by incubation for
40 1 h at 37°C.

Transfection of the siRNAs can be carried out using the same protocol as for transfection of the antisenses.

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An alternative for the iRNA is the use of vectors which allow synthesis of antisense RNAs specific for the gene to be silenced and which will pair in the transfected cells to give an siRNA. A first vector system allows expression of an antisense sequence by two promoters in opposite direction, on each side of this sequence, producing two complementary RNAs which will pair in the transfected cells and give an siRNA. Another vector system uses the synthesis of an RNA

having the sequence of the antisense followed by the sense sequence, a few nucleotides apart, which will create a stem-loop RNA structure which will be cleaved in the transfected cells to give an siRNA. These vectors are transfected conventionally as described above for the various DNAs. Stable lines which exhibit a knockout of the target gene can be obtained by antibiotic selection conventionally used to obtain lines.

In general, those skilled in the art may refer, for the iRNAs to the following publications: Elbashir S.M. et al. (2001, *Nature* **411**: 494-498), Elbashir S.M. Lendeckel W. and Tuschl T. (2001, *Genes & Dev.* **15**: 188-200) and Masters J.R., et al. (2001. *Proc. Natl. Acad. Sci. USA* **98**: 8012-8017).

Vectors which allow the expression of iRNAs can be obtained as described by Brummelkamp T.R., Bernards R., Agami R. (2002. *Science* **296**: 550-553) and Yu J.Y., DeRuiter S.L., and Turner D. (2002., *Proc. Natl. Acad. Sci. USA* **99**: 6047-6052).

Such vectors, and also cells containing such vectors, are subjects of the present application.

A subject of the present application is also medicinal products containing such oligonucleotides, vectors and cells, and pharmaceutical compositions containing a pharmacologically active amount of such oligonucleotides, vectors and cells and pharmaceutically acceptable excipients.

Another subject of the present invention is the use of such oligonucleotides, vectors and cells, for producing a medicinal product for preventing and/or treating leptin-related pathological conditions.

A subject of the invention is also a method of curative or preventive treatment of leptin-related diseases, consisting in administering such oligonucleotides, vectors and cells to a patient suffering from said disease.

Another subject of the invention is a method for determining the modification, by a compound, of the interaction between the OB-RGRP or the MYO47 protein, or a protein exhibiting at least 65% identity with this protein or with the MYO47 protein, and the leptin receptor.

It also relates to fusion proteins for implementing this method, and also to nucleic acids encoding these proteins.

A subject of the invention is also a method of curative or preventive treatment of leptin-related diseases, consisting in administering a ligand selected using the method defined above to a patient suffering from said disease.

A first subject of the present invention is therefore a fusion protein which is composed of a sequence exhibiting at least 65% identity with the sequence SEQ ID No. 4, or the sequence SEQ ID No. 16, or of a substantial part of the sequence SEQ ID No. 4 or of the sequence SEQ ID No. 16, and of an energy-donor or

energy-acceptor protein, or of a substantial and active part of an energy-donor or energy-acceptor protein.

The fusion proteins according to the present invention are composed in substance of a component corresponding to part or all of a sequence exhibiting at least 65%, preferentially at least 75%, and even more preferentially at least 85% or 95%, identity with the sequence SEQ ID No. 4 or the sequence SEQ ID No. 16, or of a substantial part of the sequence SEQ ID No. 4 or of the sequence SEQ ID No. 16, and of a component corresponding to an energy-donor or -acceptor protein. They may, however, comprise other amino acid sequences, derived from other proteins, such as signal sequences.

Advantageously, the energy-donor protein is Renilla luciferase. It may, however, be any other energy-donor protein such that the emission spectrum of the donor overlaps the excitation spectrum of the acceptor sufficiently to allow efficient energy transfer between the two partners. It may thus be GFP, if the energy transfer is FRET, or else aequorin if the energy transfer is CRET. Aequorin can be obtained and used as described in patent application EP 0 187 519, or in the article by Inouye et al. (PNAS USA 82 : 3154-3158 (1985)).

As regards the energy-acceptor fluorescent protein, it is preferentially DsRed, GFP or a mutant of this protein, such as YFP, EYFP, wild-type GFP, GFPS65T, Topaz or GFP₁₀.

It may however be any other energy-acceptor fluorescent protein such that the excitation spectrum of the acceptor and the emission spectrum of the donor overlap sufficiently to allow efficient energy transfer between the two partners.

These proteins are known to those skilled in the art, who can find their sequences in the literature, in particular in the review by Blinks et al. (Pharmacol. Rev. 28 : 1-93 (1976)). In particular, GFP is described by Tsien (Annu. Rev. Biochem. 67 : 509-544 (1998)) and the cloning thereof is described by Prasher et al. (Gene 111 : 229-233 (1992)). As regards the cloning of DsRed, it is described by Matz et al. (Nat. Biotechnol. 17 : 969-973 (1999)). For Rluc, those skilled in the art can refer to Blinks et al. (Pharmacol. Rev. 28 : 1-93 (1976)) or else to Lorenz et al. (PNAS 88: 4438-4442 (1991)).

Particularly advantageously, the donor and acceptor fusion proteins have one of the sequences SEQ ID No. 6, SEQ ID No. 8, SEQ ID No. 12, SEQ ID No. 14, SEQ ID No. 18 or SEQ ID No. 20, or a variant of this sequence exhibiting at least 65% identity.

Other subjects of the present invention are nucleic acids encoding these proteins. Such nucleic acids may be complementary or genomic DNAs, or RNAs. These nucleic acids or polynucleotides can be in single-chain form or in the form of duplex.

They are particularly advantageously complementary DNAs.

Preferentially, a subject of the invention is a nucleic acid having at least 65%, preferentially at least 75%, and even more preferentially at least 85% or 95%, nucleotide identity with a nucleic acid of sequence SEQ ID No. 5, SEQ ID No. 7, SEQ ID No. 11, SEQ ID No. 13, SEQ ID No. 17 or SEQ ID No. 19.

According to yet another aspect, the invention relates to a nucleic acid which hybridizes, under high stringency hybridization conditions, with a nucleic acid as defined above, and more particularly a nucleic acid of nucleotide sequence SEQ ID No. 5, SEQ ID No. 7, SEQ ID No. 11, SEQ ID No. 13, SEQ ID No. 17 or SEQ ID No. 19, or a nucleic acid of complementary sequence.

For the purpose of the present invention, the "percentage identity" between two nucleotide or amino acid sequences can be determined by comparing two optimally aligned sequences through a window of comparison.

The part of the nucleotide sequence or polypeptide in the window of comparison, may thus comprise additions or deletions (for example gaps) compared to the reference sequence (which does not comprise these additions or these deletions) so as to obtain optimal alignment of the two sequences.

The percentage is calculated by determining the number of positions at which an identical nucleic acid base or amino acid residue is observed for the two (nucleic acid or peptide) sequences compared, in dividing the number of positions at which there is identity between the two bases or amino acid residues by the total number of positions in the window of comparison, and then multiplying the result by 100 in order to obtain the percentage sequence identity.

The optimal alignment of the sequences for comparison can be produced on a computer using known algorithms contained in the WISCONSIN GENETICS SOFTWARE PACKAGE, GENETICS COMPUTER GROUP (GCG), 575 Science Doctor, Madison, WISCONSIN.

By way of illustration, the percentage sequence identity may be produced using the BLAST software (versions BLAST 1.4.9 of March 1996, BLAST 2.0.4 of February 1998 and BLAST 2.0.6 of September 1998), using exclusively the default parameters (S. F. Altschul et al, J. Mol. Biol. 1990 215 : 403-410, S. F Altschul et al, Nucleic Acids Res. 1997 25 : 3389-3402). Blast searches for sequences similar/homologous to a reference "request" sequence using the algorithm of Altschul et al. The request sequence and the databases used may be peptide-based or nucleic acid-based, any combination being possible.

For the purpose of the present invention, the expression "high stringency hybridization conditions" will be intended to mean the following conditions:

1 – Membrane competition and PRE HYBRIDIZATION:

- Mix : 40 µl of salmon sperm DNA (10 mg/ml)+ 40 µl of human placenta DNA (10 mg/ml)
- Denature for 5 min at 96°C, then immerse the mixture in ice.
- Remove the 2X SSC and pour 4 ml of formamide mix into the hybridization tube containing the membranes.
- Add the mixture of the two denatured DNAs.
- Incubate at 42°C for 5 to 6 hours, with rotation.

2- Labeled probe competition:

- Add 10 to 50 μ l of Cot I DNA to the labeled and purified probe, depending on the amount of repetition.

- Denature for 7 to 10 mn at 95°C.

5 - Incubate at 65°C for 2 to 5 hours.

3- Hybridization :

- Remove the prehybridization mix.

- Mix 40 μ l of salmon sperm DNA + 40 μ l of human placental DNA; denature for 5 min at 96°C, then immerse in ice.

10 - Add 4 ml of formamide mix, the mixture of the two DNAs and the denatured Cot I DNA/labeled probe to the hybridization tube.

- Incubate for 15 to 20 hours at 42°C, with rotation.

4- Washes :

- One wash at ambient temperature in 2X SSC, to rinse.

15 - 2 times 5 minutes at ambient temperature in 2X SSC and 0.1% SDS at 65°C.

- 2 times 15 minutes at 65°C in 1X SSC and 0.1% SDS at 65°C.

Wrap the membranes in Saran wrap and expose.

20 The hybridization conditions described above are suitable for hybridization, under high stringency conditions, of a nucleic acid molecule of varying length of 20 nucleotides to several hundred nucleotides.

It goes without saying that the hybridization conditions described above can be adjusted as a function of the length of the nucleic acid the hybridization of which is desired, or of the type of labeling chosen, according to the techniques well known to those skilled in the art.

25 The suitable hybridization conditions may, for example, be adjusted according to the teaching contained in the work by HAMES and HIGGINS (1985, "Nucleic acid hybridization : a practical approach", Hames and Higgins Ed., IRL Press, Oxford) or else in the work by F. AUSUBEL et al. (1989, Current Protocols in Molecular Biology, Green Publishing Associates and Wiley Interscience, N.Y.).

35 The proteins which are the subjects of the present invention can be obtained by any means known to those skilled in the art. They are, however, advantageously obtained by expression of the nucleic acids as described above, encoding these proteins, optionally inserted into expression vectors, into cells advantageously chosen, optionally followed by an extraction and a purification which may be total or partial.

40 The invention also relates to a recombinant vector comprising a nucleic acid according to the invention.

Advantageously, such a recombinant vector will comprise a nucleic acid chosen from the following nucleic acids:

45 a) a nucleic acid encoding a protein having at least 65% amino acid identity with a sequence SEQ ID No. 6, SEQ ID No. 8, SEQ ID No. 18 or SEQ ID No. 20, or a peptide fragment or a variant thereof;

b) a nucleic acid comprising a polynucleotide having a sequence SEQ ID No. 5, SEQ ID No. 7, SEQ ID NO. 17 or SEQ ID No. 19, or a fragment or a variant thereof;

c) a nucleic acid having at least 65% nucleotide identity with a nucleic acid having a sequence SEQ ID No. 5, SEQ ID No. 7, SEQ ID No. 17 or SEQ ID No. 19, or a fragment or a variant thereof;

5 d) a nucleic acid which hybridizes, under high stringency hybridization conditions, with a nucleic acid of sequence SEQ ID No. 5, SEQ ID No. 7, SEQ ID No. 17 or SEQ ID No. 19, or a fragment or a variant thereof.

10 For the purposes of the present invention, the term "vector" will be intended to mean a circular or linear DNA or RNA molecule which is indifferently in single-stranded or double-stranded form.

According to one embodiment, the expression vector comprises, besides a nucleic acid in accordance with the invention, regulatory sequences which make it possible to direct the transcription and/the translation thereof.

15 According to an advantageous embodiment, a recombinant vector according to the invention will in particular comprise the following elements:

(1) elements for regulating the expression of the nucleic acid to be inserted, such as promoters and enhancers;

20 (2) the coding sequence included in the nucleic acid in accordance with the invention to be inserted into such a vector, said coding sequence being placed in phase with the regulatory signals described in (1); and

(3) suitable transcription initiation and stop sequences.

25 In addition, the recombinant vectors according to the invention may include one or more origins of replication in the cellular hosts in which their amplification or their expression is desired, markers or selection markers.

By way of examples, the promoters for eukaryotic cells will comprise the thymidine kinase promoter of the HSV virus or else the mouse metallothionein-L promoter.

30 In general, in choosing a suitable promoter, those skilled in the art may advantageously refer to the work by SAMBROOK et al. (1989, "Molecular Cloning : A Laboratory Manual," 2nd ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.) or else to the techniques described by FULLER et al. (1996, *Immunology in Current Protocols in Molecular Biology*, Ausubel et al).

35 The preferred vectors according to the invention are plasmids, such as, for example, the vectors pCDNA3 (Invitrogen), pQE70, pQE60, pQE9 (Qiagen), psiX174, pBluescript SA, pNH8A, pNH16A, pNH18A, pNH46A, pWLNEO, pSV2CAT, pOG44, pXTI and pSG(Stratagene).

40 They may also be vectors of the *baculovirus* type, such as the vector pVL1392/1393 (PharMingen) used to transfect cells of the Sf9 line (ATCC No. CRL 1711) derived from *Spodoptera frugiperda*.

They may also be adenoviral vectors, such as human adenovirus type 2 or 5.

45 A recombinant vector according to the invention may also be a retroviral vector or else an adeno-associated vector (AAV). Such adeno-associated vectors are, for example, described by FLOTTE et al. (1992, *Am. J. Respir. Cell Mol. Biol.*, **7** : 349-356

Objects of the present invention are also cells comprising a protein, a nucleic acid or a vector as described above, or fragments of these cells, lysates of these cells or else membranes of these cells.

Such cells may be cells isolated from an organism and cultured in a suitable growth medium. They are, however, preferentially cell lines. Thus, such lines are particularly advantageously the cells lines HEK 293, COS (ATCC No. CRL 1650),
 5 COS-M6 and HeLa (ATCC No. CCL2), or else Cv 1 (ATCC No. CCL70), Sf-9 (ATCC No. CRL 1711), CHO (ATCC No. CCL-61) or 3T3 (ATCC No. CRL-6361).

The membranes of these cells can be prepared by any method known to those skilled in the art.

10 Preferentially, they will be prepared by mechanical grinding of the cells and then centrifugation of the suspensions obtained, as illustrated in the examples which follow.

15 The present invention also relates to compositions comprising cells as described above and saponin.

The present invention also relates to a method for determining the modification, by a compound, of the interaction between the OB-RGRP, the MY047 protein or a protein exhibiting at least 65% identity with the
 20 sequence SEQ ID No. 4 or the sequence SEQ ID No. 16, and the leptin receptor, comprising the steps consisting in:

- bringing said compound into contact with a protein exhibiting at least 65% identity with the sequence SEQ ID No. 4 or the sequence SEQ ID No. 16, and the leptin receptor, or cells, or fragments or lysates or membranes of
 25 cells, comprising such proteins, and optionally a suitable enzyme substrate, and

- measuring the interaction between a protein exhibiting at least 65% identity with the sequence SEQ ID No. 4 or the sequence SEQ ID No. 16, and the leptin receptor.

30 Preferentially, said compound is brought into contact with an energy-donor fusion protein and an energy-acceptor fusion protein, or cells, or fragments or lysates or membranes of cells, comprising such a protein, and optionally a suitable enzyme substrate.

35 Preferentially, said method is brought into contact with cells treated with an agent which permeabilizes these cells, such as saponin.

The energy-donor fusion proteins and the energy-acceptor fusion proteins are chosen such that the energy resulting from the activation of the donor may be
 40 transferred efficiently to the acceptor.

In an advantageous embodiment of said method, the energy-donor fusion protein is a protein from fusion with luciferase or a substantial part of luciferase, in which case the substrate is advantageously coelenterazine.

45 In a preferential embodiment of said method, the energy-acceptor fusion protein is a protein from fusion with YFP or a substantial part of YFP.

In an advantageous embodiment of said method, the energy transfer measured in the presence of the test compound is compared to that measured in the absence of the test compound.

5 In another advantageous embodiment of said method, the energy transfer measured in the presence of the test compound and of leptin (or a ligand of the receptor) is compared to that measured in the presence of the compound in the absence of leptin (or a ligand of the receptor).

10 Preferentially, the method is carried out on cell membranes, as described above.

Preferentially, the donor and acceptor proteins according to the present invention are chosen such that the energy transfer takes place by first or second generation BRET (for Bioluminescence Resonance Energy Transfer) or LRET (for
15 Luminescence Resonance Energy Transfer). However, such an energy transfer may be effected by FRET (for Fluorescence Resonance Energy Transfer) or else by CRET (for Chemiluminescence Resonance Energy Transfer).

Whatever the type of energy transfer, the energy-donor fusion protein/energy-acceptor fusion protein pairs are chosen so as to allow such transfer.

20 BRET2 (2nd generation) consists of energy transfer between *Renilla* luciferase and a mutant GFP, GFP₁₀, using a suitable substrate, DeepblueCTM coelenterazine (Biosignal Packard).

CRET consists of energy transfer between aequorin, which is a luciferase, and GFP.

25 FRET consists of energy transfer between two proteins of the GFP family having different spectra.

To implement these transfers, those skilled in the art may refer to D. Ramsay et al. (Biochem J 365: 429-40 (2002)) and to K. Yoshioka et al. (FEBS Lett 523: 147-151 (2002)) for BRET2, to Baubet et al. (PNAS USA 97 : 7260-7265 (2000)) for
30 CRET, and to Matyus (J Photochem Photobiol B 12: 323-337 (1992)) and Pollok and Heim (Trends Cell Biol 9:57-60 (1999)) for FRET.

Another subject of the present invention is a method for screening or detecting compounds intended for the prevention and/or treatment of leptin-
35 related pathological conditions, comprising the steps consisting in:

- bringing said compound into contact with a protein exhibiting at least 65% identity with the sequence SEQ ID No. 4 or the sequence SEQ ID No. 16, and the leptin receptor, or cells, or fragments or lysates or membranes of cells, comprising such proteins, and optionally a suitable enzyme substrate,
40 and

- measuring the interaction between a protein exhibiting at least 65% identity with the sequence SEQ ID No. 4 or the sequence SEQ ID No. 16, and the leptin receptor.

45 Preferentially, the protein exhibiting at least 65% identity with the sequence SEQ ID No. 4 or the sequence SEQ ID No. 16 is the OB-RGRP or MY047.

The method according to the present invention is compatible with the 96-well or 384-well plates generally used. It does not require the use of radioactive molecules, but is sensitive, reproducible and rapid, and the result is easy to read. This characteristic is particularly advantageous for carrying out large scale screening.

The present invention also relates to the use of compounds selected using a method consisting in:

- bringing said compound into contact with a protein exhibiting at least 65% identity with the sequence SEQ ID No. 4 or the sequence SEQ ID No. 16, and the leptin receptor, or cells, or fragments or lysates or membranes of cells, comprising such proteins, and optionally a suitable enzyme substrate, and
- measuring the interaction between a protein exhibiting at least 65% identity with the sequence SEQ ID No. 4 or the sequence SEQ ID No. 16, and the leptin receptor.

A subject of the present invention is, finally, a method of curative or preventive treatment of leptin-related diseases or diseases related to its receptor, comprising the steps of:

- selecting said compound using a method consisting in:
 - +bringing said compound into contact with a protein exhibiting at least 65% identity with the sequence SEQ ID No. 4 or the sequence SEQ ID No. 16, and the leptin receptor, or cells, or fragments, or lysates or membranes of cells, comprising such proteins, and optionally a suitable enzyme substrate, and
 - +measuring the interaction between a protein exhibiting at least 65% identity with the sequence SEQ ID No. 4 or the sequence SEQ ID No. 16, and the leptin receptor
- administering said compound to a patient suffering from said disease.

Leptin-related pathological conditions may be diseases related to a decrease in bone density, such as, for example, osteoporosis, or, conversely, those related to considerable calcification.

They may also be diseases which have an effect on weight, such as obesity, diabetes or anorexia.

They may also be diseases which have an effect on sexual maturation, hematopoiesis, angiogenesis, thrombus formation, the regulation of immunity and inflammation, fetal development and cancer.

The compounds of the invention, oligonucleotides, iRNAs, or other compounds, may be formulated in pharmaceutical compositions for the purpose of topical, oral, parenteral, intranasal, intravenous, intramuscular, subcutaneous, intraocular administration, etc. Preferentially, the pharmaceutical compositions contain pharmaceutically acceptable vehicles for an injectable formulation. They may in particular be isotonic, sterile, saline (monosodium or disodium phosphate, sodium, potassium, calcium or magnesium chloride, etc., or mixtures of such salts) solutions, or dry, in particular lyophilized, compositions which, by addition, as

appropriate, of sterilized water or of physiological saline, make it possible to constitute injectable solutes.

The formulation of therapeutic compositions and their administration fall within the competence of those skilled in the art.

5 The formulation of the compounds may include various products known to those skilled in the art. Preferentially, the compounds may, for example, have salts, such as sodium, potassium, ammonium, magnesium, calcium, polyamines, or hydrochloric, hydrobromic, sulfuric, phosphoric or nitric acid, added to them. Other salts can also be used, such as those originating from acetic, oxalic, tartaric, 10 succinic, maleic, fumaric, gluconic, citric, malic, ascorbic, benzoic, tannic, palmitic, alginic, polyglutamic, naphthalenesulfonic, methanesulfonic, p-toluenesulfonic, naphthalenedisulfonic or polygalacturonic acid. Finally, chlorine, bromine and iodine salts can also preferentially be used.

15 The composition and the formulation for topical administration can include transdermal patches, ointments, lotions, creams, gels, drops, suppositories, sprays, liquids and powders.

20 The composition and the formulation for oral administration can include powders, granules, microparticles, nanoparticles, suspensions, solutions, which may or may not be aqueous, capsules, gelatin capsules, sachets, tablets or mini tablets. Thickeners, flavors, diluents, emulsifiers, dispersing agents or binders may be added.

25 The composition and the formulation for parenteral, intrathecal or intraventricular administration can include sterile aqueous solutions which can also contain buffers, diluents and other additives, such as, but not limited to, penetration-increasing agents, transporting products and excipients.

30 The composition can be formulated and used as a foam, an emulsion, a microemulsion, cationic, pH-sensitive or negatively charged liposomes, and transferomes.

35 In general, the various formulations can contain a mixture of one or more agents, such as, but not limited to, agents which increase the penetration of the compound (surfactants, bile salts, chelating agents, non-chelating surfactants), excipients (binders, fillers, lubricants, disintegrating agents, wetting agents), or transporters (water, saline solutions, alcohols, polyethylene glycol, gelatin, lactose, amylose, magnesium stearate, talc, silicic acid, viscous paraffin, 40 hydroxymethylcellulose, polyvinylpyrrolidone). Other components can be added, such as dyes, flavors, preserving agents, antioxidants, opacifiers, thickeners and stabilizers.

45 The dosage depends on the severity and on the sensitivity of the state of the disease to be treated, with a treatment period possibly ranging from a few days to a few months, or until the treatment is effective or a reduction in the disease is observed. The optimum dosage can be calculated from measurements of accumulation of the therapeutic agent in the patient's body. Those skilled in the art can easily determine the optimum dosages, the methods of dosage and the rates

of repetition of these dosages. The optimum dosages can vary as a function of the relative effectiveness of each oligonucleotide or iRNA, and can, in general, be estimated by measuring the EC50s of the doses used in vitro and in vivo in animal models. In general, the dosage is between 0.01 μ g and 100 g per kilo of bodyweight and can be administered one or more times, daily, weekly, monthly or annually, or even once every 2 to 20 years.

Competent individuals can easily determine the rate of repetition of the dosages based on the amount of time the compound is present in the body fluids or the tissues. Subsequent to a successful treatment, it may be desirable for the patient to continue a maintenance therapy in order to prevent reappearance of the disease; to do this, the oligonucleotide or the iRNA is administered at maintenance doses ranging from 0.01 μ g to 100 g per kilo of bodyweight, one or more times a day, up to once every 20 years.

The administration of antisense in vivo has been carried out successfully by various authors, using protocols of simple injection of antisense intravenously (He et al. (1998) *Zhonghua Shi Yan He Lin Chuang Bing Du Xue Za Zhi* 12:1-4) or intracerebrally (Yoburn et al. (2003) *Synapse* 47: 109-116, Tischkau et al. (2003) *J. Biol. Chem.* 278: 718-723). In the last two years, more complex systems for targeting antisense in the organism have been developed and used successfully (Morishita et al. (2002) *J. Endocrinol.* 175: 475-485, Bartsch et al. (2002) *Pharm. Res.* 19: 676-680), making it possible, in mice and rats, to treat various cancers (Rait et al. (2002) *Mol. Med.* 8: 475-486, Ochietti et al. (2002) *J. Drug. Target* 10: 113-121, Eder et al. (2002) *Cancer Gene Ther.* 9:117-125). The transfection of antisenses involves the same methods as for the transfection of iRNAs, making it possible to envision the same applications in vivo for the iRNAs. With this in mind, it is possible to imagine targeting the antisense or the iRNAs to the central nervous system, in order to treat disorders of central origin (obesity), but also those produced by a peripheral action of leptin receptors. More particularly, it is possible to envision there being an action of the antisenses or of the iRNAs on the transport of leptin across the blood-brain barrier, involving OB-R. Moreover, endothelial cells have already been successfully targeted using an in vivo antisense strategy (Bartsch et al. (2002) *Pharm. Res.* 19: 676-680).

Figures :

Figure 1

Sequences of the various antisense ODNs used.

Figure 2

Alignment of the OB-RGRP protein sequences of various species and of the human MY047 protein sequence. The potential transmembrane domains were determined by various methods (HMMTOP, TMHMM, TopPred2, TMpred) and are written in bold.

Figure 3

Topology of OB-RGRP studied by BRET, using the double fusion protein YFP-OB-RGRP-Luc. Figure 3a: diagrammatic representation of the topology of OB-

RGRP for the models 3 and 4TM. Figure 3b: results of the BRET experiments using the proteins indicated. The data are expressed in mBU.

Figure 4

Study of the oligomerization of OB-RGRP with SDS-PAGE experiments and immunoprecipitations. Figure 4a: the cells expressing the fusion proteins indicated were treated or not treated with 2 mmol.L⁻¹ dithiobis(succinimidyl propionate) (DSP) in PBS (1X, pH7.4) in order to crosslink the protein complexes. The proteins were separated by SDS-PAGE and the proteins from fusions with YFP were detected using a specific anti-YFP antibody. Figure 4b: the cells expressing the construct 6Myc-OB-RGRP were solubilized with 1% of digitonin or 5% of SDS and the solubilized material was immunoprecipitated with an anti-myc antibody. The precipitates were subjected to separation by SDS-PAGE and the proteins tagged with myc were detected with an anti-myc antibody.

Figure 5

Identification of the molecular determinants involved in the oligomerization of OB-RGRP. The proteins from fusions with the OB-RGRP truncations were treated as described in Fig. 4b. TM, transmembrane domain.

Figure 6

Study of the oligomerization of OB-RGRP in live HEK cells, by BRET technology. Figure 6a: the fusion proteins indicated were coexpressed at an equimolar ratio, and BRET measurement experiments were carried out. Figure 6b: constant amounts of the plasmid OB-RGRP-Luc were coexpressed with increasing amounts of the plasmid OB-RGRP-YFP and BRET measurements were carried out. MT2R-Luc, protein from fusion of the melatonin receptor MT2 with luciferase.

Figure 7

Interaction of OB-R_s and of OB-RGRP studied by BRET. The fusion proteins indicated were coexpressed at an equimolar ratio and BRET measurements were carried out. IR-YFP, protein from fusion of the insulin receptor with YFP.

Figure 8

Dose-dependent activation of the reporter genes for STAT3 (Figure 8a) and STAT5 (Figure 8b) in HeLa cells, by OB-R_i in the presence of overexpression of the OB-RGRP protein constructs as indicated.

Figure 9

Effect of the overexpression of OB-RGRP on the expression of OB-R at the surface of the cells. HEK 293 cells transfected or not transfected with the OB-RGRP expression vector, and COS cells transfected with the OB-R_i or OB-R_s expression vectors and +/- the OB-RGRP vectors, were used to determine the amount of receptors expressed at the surface and the total expressed in the cells, by ¹²⁵I-leptin-binding experiments.

Figure 10

Effect of the various antisense oligodeoxynucleotides (ODNs) on the level of OB-RGRP messengers observed by semiquantitative RT-PCR. Figure 10a:

determination of the linear zone of amplification of the OB-RGRP and GAPDH transcripts, as a function of the number of PCR cycles. Figure 10b: quantification of the results shown in panel a. Figure 10c: determination of the relative levels of expression of the OB-RGRP mRNAs at 26 PCR cycles, in the cells incubated with the various antisense ODNs.

Figure 11

Effect of the OB-RGRP-specific antisense ODNs on the activation of a STAT3 reporter gene. The HeLa cells were cotransfected firstly with the OB-R_I expression vector and the constructs of the reporter genes for STAT3 or 5, and then with the antisense ODNs indicated. After 48 hours of stimulation or no stimulation with 10 nmol.L⁻¹ of leptin.

Figure 12

Effect of the OB-RGRP-specific antisense ODNs on the surface expression of OB-R. The HeLa cells were transfected or not transfected with the OB-R_I or OB-R_S expression plasmids, before a second transfection with the antisense ODNs indicated, or no second transfection. 48 h post-transfection, the total amount of OB-R and the fraction exposed at the surface were determined in binding experiments with ¹²⁵I-leptin.

The present invention is illustrated, without, however, being limited, by the following examples.

Materials and methods used in the examples

Plasmid construction

The proteins from fusion of OB-R with YFP and luciferase were constructed by ligation of YFP and of luciferase to the C-terminal portion of the OB-R receptors, by standard molecular biology techniques. The coding region of YFP was obtained from the vector Cytogem®-Topaze (pGFPtpz-N1) (Packard, Meriden, CT) and was inserted into the EcoRV site of the vector pcDNA3/CMV (Invitrogen, Groningen, The Netherlands) containing a modified polylinker. The coding region of *Renilla* luciferase was obtained from the vector pRL-CMV (Promega, Madison, WI) and inserted into the EcoRV site of the modified vector pcDNA3. The coding regions of OB-R_I and of OB-R_S (a gift from Dr. Gainsford, Royal Melbourne Hospital, Victoria, Australia) were inserted into the two vectors described above, respectively into the EcoR1/BamH1 and Nhe1 sites. The stop codons were deleted by site-directed mutagenesis and the frame of the fusion proteins was adjusted at the same time.

The vector pcDNA3-OB-RGRP was obtained by insertion of the coding region of OB-RGRP, obtained from the vector pCDNA3-Di1, into the EcoR1 and Xba1 sites of the vector pcDNA3/CMV (Invitrogen, Groningen, The Netherlands). The stop codon of OB-RGRP was deleted by site-directed mutagenesis. The vector pcDNA3-OB-RGRP-Luc was obtained by digestion of the vector pRL-CMV N3 (Promega, Madison, WI) with Sma1 and Hpa1 and by insertion of the fragment corresponding to the coding region of *Renilla* luciferase, after the coding region of OB-RGRP, into the filled-in BspE1 site of the vector pcDNA3-OB-RGRP.

The vector pcDNA3-YFP was obtained by subcloning the coding region of YFP from the vector pGFPtpz-N1 (Packard, Meriden, CT) inserted into the EcoRV site of the vector pcDNA3/CMV. The vector pcDNA3-OB-RGRP-YFP was obtained by insertion of the BamH1/BspE1 fragment of the vector pCDNA3-OB-RGRP non-stop into the vector pcDNA3-YFP digested with the BamH1 and Age1 enzymes.

The construct pcDNA3-GFP-OB-RGRP-Luc was obtained by insertion of the OB-RGRP-Luc fragment of the vector pcDNA3-OB-RGRP-Rluc, cleaved with EcoR1, into the EcoR1 site of the vector pcDNA3-YFP. The stop codon of the YFP was removed by site-directed mutagenesis.

The vector 6Myc-OBR-GRP (4TM) was obtained by insertion of the 6myc fragment of the vector pCDNA3-RSV-6Myc into the BamH1 and EcoR1 sites of the vector pCDNA3-OB-RGRP. The various OB-RGRP deletions (2 and 3 TM) were obtained by PCR and the insertion into the vector pcDNA3, into the EcoR1 and Xba1 sites. The coding sequence of MY047 was obtained by RT-PCR on mRNAs of human origin. The PCR fragment was digested with the EcoR1/Xba1 restriction enzymes and inserted into the vector pcDNA3-Topaze cleaved with the same enzymes. The stop codon of the YFP was then removed by site-directed mutagenesis, so as to obtain the vector pcDNA3-YFP-MY047. The vector pcDNA3-MY047-GFP was obtained by insertion of the DNA fragment obtained by PCR on the vector pcDNA3-YFP-MY047 and cleaved with BamH1, then inserted into the vector pcDNA3-YFP cleaved with the same enzyme. Insertion of the same fragment into the vector pcDNA3-Rluc cleaved with BamH1 made it possible to obtain the vector pcDNA3-MY047-Rluc.

All the constructs were verified by sequencing.

Cell culture and transfection

The HEK 293, COS-7 and HeLa cells were cultured in DMEM supplemented with 10% (v/v) of SVF, 4.5 g/liter of glucose, 100 U/ml of penicillin, 0.1 mg/ml of streptomycin and 1 mmol.L⁻¹ of glutamine (all from Life Technologies, Gaithersburg, MD). The transient transfections were carried out with the FuGene 6 reagent (Roche, Basel, Switzerland) according to the supplier's instructions.

Preparation of membranes and solubilization

The membranes were prepared as previously described (19), and resuspended in 75 mmol.L⁻¹ Tris (pH 7.4), 12.5 mmol.L⁻¹ MgCl₂ and 5 mmol.L⁻¹ EDTA, and immediately used in BRET experiments.

SDS PAGE and Western blotting

The total lysates were prepared by washing the cells once with cold PBS (pH 7.4) and denatured by adding loading buffer (30 mmol.L⁻¹ Tris HCl, pH 6.8, 1% glycerol, 5% SDS, 50 mmol.L⁻¹ DTT and 0.05% bromophenol blue). The total lysates or the immunoprecipitates were incubated for 10 minutes at 90°C and then loaded onto 10% acrylamide gel for separation by electrophoresis (SDS-PAGE). The proteins were then transferred onto a nitrocellulose membrane and revealed with specific primary antibodies: anti-YFP (8367-1 Living Colors) diluted to 1/200, anti-myc A14 (sc-789 TEBU Peprotech Santa Cruz Biotechnology) diluted to 1/500, then a secondary antibody coupled to peroxidase (anti-rabbit goat IgG; Jackson ImmunoResearch Laboratories, Inc., West Baltimore Pike) diluted to

1/10,000. The immunoreactive bands were revealed with an ECL kit (Pharmacia Biotech).

Immunoprecipitation

5 Two days after transfection, the cells were washed once with cold PBS, and the proteins were extracted by incubation for 15 minutes in lysis buffer (1X PBS, 1% Nonidet P40, 0.5% sodium deoxycholate, 0.1% SDS, 0.02% NaN₃, 10 mg.L⁻¹ benzamidine and 5 mg/L⁻¹ trypsin inhibitors). The lysate was centrifuged at 18000 g for 15 min and the supernatant was then incubated for 3 hours at 4°C
10 with an anti-myc antibody coupled to agarose beads (sc 40AC TEBU preprotech, Santa CRUZ Biotechnology). The precipitates were washed three times with cold lysis buffer and denatured with loading buffer for SDS-PAGE.

Radiobinding experiments

15 The radiobinding experiments were carried out as previously described (Barr et al.), with slight modifications. To determine the surface leptin binding, the cells cultured in the 6-well plates were washed twice with cold PBS and incubated in the binding buffer (DMEM, 25 mmol.L⁻¹ Hepes, pH 7.4, 1% BSA) containing 100,000 cpm/well of ¹²⁵I-leptin (PerkinElmer life sciences, Paris, France) in the
20 presence or absence of 200 nmol.L⁻¹ of leptin (PreproTech Inc, USA) for 4 h at 4°C. The cells were washed twice with cold PBS, then lyzed in 1N NaOH, and the radioactivity was determined in a γ counter. To determine the total binding of leptin, the cells cultured in dishes 10 cm in diameter were solubilized in 1.5 ml of binding buffer containing 0.15% of digitonin, for 2 h at 4°C. The extracts were
25 centrifuged for 30 min at maximum speed and at 4°C. The supernatants (0.2 ml) were incubated with 100,000 cpm of ¹²⁵I-leptin in the presence or absence of 200 nmol.L⁻¹ of leptin, in a total volume of 0.25 ml, with constant rotation at 4°C overnight. 0.5 ml of γ -globulin (1.25 mg/ml) and 0.5 ml of polyethylene glycol 6000 (25% w/v) were added in order to precipitate the receptor-ligand complexes, which
30 were centrifuged at 17,000 g for 3 min. The pellet was washed once with 1 ml of polyethylene glycol 6000 (12% w/v) and the radioactivity was determined in a γ -counter.

Reporter gene activation assay

35 The HeLa cells cultured in wells of 6-well plates were cotransfected with 500 ng of a reporter plasmid expressing *firefly* luciferase under the control of STAT3 or STAT5 factor response elements (a gift from Dr. Levy, University of New York, New York, USA), 250 pg of the expression vector pcDNA3-*Renilla* luciferase (used as internal standard between the samples) and with 500 ng of the
40 various OB-R expression vectors or the vector alone. 48 h after transfection, the cells were starved overnight in Optimem medium (Invitrogen, Groningen, The Netherlands) containing 1% of BSA, before stimulation with 10 nmol.L⁻¹ of leptin, or no stimulation, for 48 h. The cells were then washed once with PBS, then lyzed in passive lysis buffer (Promega Corporation, Madison, WI) for 15 min at ambient
45 temperature. The total lysates were centrifuged for 2 min at 15,000 g and the supernatants were used in an assay to measure luciferase (Dual Luciferase Assay System from Promega Corporation, Madison, WI) using a Berthold luminometer (Lumat LB 9507). The results are expressed as ratio of *firefly* luciferase activity to *Renilla* luciferase activity.

BRET measurements in microplates

48 h after transfection, the COS-7, HeLa or HEK 293 cells expressing the OB-R fusion proteins were detached and washed in PBS. $1-2 \times 10^5$ cells were distributed into wells of optiplate plates (96-well, Packard Instrument Company, Meriden, CT) in the presence or absence of the ligands, and incubated at 25°C. Alternatively, the same procedure was carried out with membranes prepared from the cells expressing the various constructs. The substrate, coelenterazine h (Molecular Probes, Eugene, OR), was added at a final concentration of $5 \mu\text{mol.L}^{-1}$ and the readings were carried out with a FusionTM luminometer/fluorimeter (Packard Instrument Company, Meriden, CT), which makes it possible to measure luminescence through two filters (luciferase filter: $485 \pm 10 \text{ nm}$; YFP filter: $530 \pm 12.5 \text{ nm}$). The BRET ratio was defined as the difference in emission at 530 nm/485 nm of the cells cotransfected with the Luc and YFP fusion proteins and the emission at 530 nm/485 nm of the Luc fusion protein transfected alone into the cells. The results are expressed as milliBRET units (mBU), 1 mBRET corresponding to the values of the differences in the ratios multiplied by 1000.

RT-PCR

The total RNAs were extracted by the method of Chomczynski and Sacchi (Chomczynski P., and Sacchi N. (1987) Anal. Biochem. 162, 156-159). 1 μg of RNA is denatured for 5 minutes at 68°C and then abruptly cooled for 5 min at 4°C. The denatured sample is reversed transcribed for 1 h at 37°C in 20 μl of RT reaction medium ($5 \mu\text{mol.L}^{-1}$ PdN6, $10 \mu\text{mol.L}^{-1}$ DTT, 50 mmol.L^{-1} Tris-HCl, pH=8.3, 75 mmol.L^{-1} KCl, 5 mmol.L^{-1} MgCl_2 , $500 \mu\text{mol.L}^{-1}$ dNTP, 200U RT-MMLV). A 2.5 μl aliquot of this reaction is used for a PCR reaction in a final volume of 25 μl (40 mmol.L^{-1} Tris-HCl, pH 8.4; 100 mmol.L^{-1} KCl; 1.5 mmol.L^{-1} MgCl_2 ; 0.2 mmol.L^{-1} of each dNTP; $0.141 \text{ mmol.L}^{-1}$ of primers specific for OB-RGRP (sense: CCGTGGCAGGAAGC, antisense: CAGCCACACGAGCAAG) and $0.035 \text{ mmol.L}^{-1}$ of primers specific for glyceraldehyde phosphate dehydrogenase (GAPDH) (sense: GGAGAAGGCTGGGCG, antisense: GATGGCATGGACTGTGG) and 2.5U of TAQ DNA polymerase). The following protocol was used for the PCR reaction: Initial denaturation for 3 min at 94°C, then 22 to 30 cycles of denaturation (20 sec at 94°C), hybridization (20 sec at 59°C), elongation (20 sec at 72°C) followed by a final elongation of 7 min at 72°C.

An aliquot of the PCR reaction was loaded onto a 2% agarose gel in order to separate the reaction products by electrophoresis. The expected sizes of fragments of GAPDH and of OBR-GRP are, respectively, 229 bp and 334 bp.

Oligonucleotide synthesis

The oligonucleotides were synthesized on an automatic DNA synthesizer ("Expedite MOSS" 8909 model from Applied Biosystems) by standard phosphoramidite chemistry and iodine oxidation. The demethylation was carried out with a 0.2 mol.L^{-1} solution of 3H-1,2-benzodithiol-3-one 1,1-dioxide in acetonitrile for 120 s. The detachment from the support and the deprotection were carried out in concentrated ammonia (18 h at 55°C), and the oligonucleotides were then purified by precipitation. The deprotection product was precipitated with 10 volumes of 1-butanol; the pellet taken up in one volume of 0.3 mol.L^{-1} NaCl was reprecipitated by adding 4 volumes of ethanol.

The analysis on a 20% polyacrylamide gel (in a buffer of 8 mol.L⁻¹ urea and 454 mmol.L⁻¹ Tris-borate, at pH 7.0) showed a greater than 80% proportion of product of expected length.

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Transfection of the antisense oligodeoxynucleotides

For the transfection of 300,000 cells cultured in a well of a 6-well plate, 10 µl of antisense ODN at 20 µmol.L⁻¹ were diluted in 175 µl of DMEM. 3 µl of oligofectamine (Invitrogen, Groningen, The Netherlands) and 12 µl of DMEM were incubated in a second tube for 10 min at ambient temperature. The oligofectamine/DMEM mixture was then added to the diluted antisense ODN, vortexed and incubated for 20 min at ambient temperature. During this time, the cells were washed once with PBS and once with DMEM, and then covered with 800 µl of DMEM. The ODN/oligofectamine mixture was then added dropwise to the cells and incubated for 4 h at 37°C, before adding 500 µl of DMEM supplemented with 30% serum.

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Example 1 : Topology and cellular location of OB-RGRP

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To study the topology and the subcellular location of OB-RGRP, the protein was tagged with the yellow variant of green fluorescent protein (YFP) at the end of its C-terminal tail. The fusion protein was expressed in HeLa cells and its location was determined by fluorescence microscopy.

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The results show that the fusion protein is preferentially targeted to the perinuclear membranes and into intracellular vesicles. Similar results were observed in HEK cells. No colocalization with cytoplasmic and nuclear proteins was observed, confirming the location of OB-RGRP in membranes (not shown). The exact nature of the membrane compartment was determined by colocalization studies with markers specific for subcellular compartments. A strong colocalization was observed with the invariant chain of MHC II molecules, a marker for the endocytic compartment.

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Initial analysis of the topology of OB-RGRP suggested an organization in 3 transmembrane (TM) domains (14). A similar organization has been proposed for MY047 (16). However, a new analysis of the hydrophobicity profile of the various protein sequences available for OB-RGRP and MY047 is also compatible with a 4-TM model (Fig. 2). The topology differs profoundly between these two models. In the 3-TM model, the N- and C-terminal ends are located on each side of the membrane, whereas in the 4-TM model, the two tails are oriented on the same side of the membrane (Fig. 3a). To determine the correct model, we used the resonance energy transfer (BRET) method which has recently been developed to follow protein-protein interactions in living cells (Xu et al. (1999) Proc Natl Acad Sci USA 96, 151-156). In the event of physical proximity (< 100 Å between the two interacting proteins, an energy transfer can take place between the energy-donor (Luc) and the energy-acceptor (YFP), fused to the two proteins of interest. We tagged the N-terminal tail of OB-RGRP with YFP, and the C-terminal tail with luciferase, and we observed the energy transfer by measuring BRET with this double fusion protein. The 3-TM model does not allow transfer since the two BRET partners are separated by the lipid bilayer. On the other hand, the 4-TM model predicts strong energy transfers since the two partners are located on the

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same side of the membrane. As shown in Figure 3b, a very strong energy transfer was detected for the double fusion protein in the intact cells, indicating that OB-RGRP has 4-TMs.

5 This set of results suggests that OB-RGRP is a membrane-bound protein with 4 transmembrane domains, having 3 short loops and short N- and C-terminal ends oriented on the same side of the membrane. OB-RGRP is mainly located in intracellular compartments.

10 Example 2: Oligomerization of OB-RGRP

Oligomerization is a property common to various proteins, including membrane-bound proteins such as tyrosine kinase receptors, cytokine receptors and phosphotyrosine phosphatases. It has been shown that this oligomerization plays an important role in the function of these proteins. To obtain elements in the function of OB-RGRP, we wanted to know whether this protein oligomerizes.

15 OB-RGRP was tagged with YFP at its C-terminal tail and expressed in HeLa cells. The proteins were separated by polyacrylamide gel electrophoresis under denaturing conditions (SDS-PAGE) and immunoblotting experiments were carried out with an anti-YFP antibody. Figure 4a reveals several bands specific for 20 OB-RGRP-YFP, corresponding to monomeric and dimeric forms and oligomeric complexes. Similar results were obtained with OB-RGRP tagged at the N-terminal, either with YFP or a myc epitope (Fig. 4 a,b). Formation of the OB-RGRP oligomers was observed on total cell extracts after immunoprecipitation. The use of a crosslinker on whole cells stabilizes the dimeric complexes, 25 indicating that the dimeric form is the predominant form of OB-RGRP in intact cells (Fig. 4a).

Surprisingly, OB-RGRP has unexpected properties since the oligomers are stable in the presence of various denaturing and/or dissociating agents such as 5% SDS, 1% Triton X-100, 1% Nonidet P40, 1% digitonin, 50 mmol.L⁻¹ DTT and 30 2% β -mercaptoethanol. However, similar observations were obtained for other membrane-bound proteins such as glycoporphin A and G protein-coupled β 2-adrenergic receptors. Studies on these proteins show, respectively, that **LIXXGVXXG** and **LXXXGXXXGXXXL** motifs in the transmembrane domains are essential for oligomer formation. Similar motifs were identified in the membrane 35 regions of OB-RGRP.

To identify the molecular determinants involved in the dimerization, we prepared OB-RGRP constructs exhibiting progressive deletions of the C-terminal tail (Fig. 5). A construct containing the first two potential TMs loses the ability to form oligomers. Addition of the 3rd TM restores the possibility of forming dimers. 40 However, the complete oligomerization profile was only observed in the presence of the 4 potential TMs.

Oligomers of membrane-bound proteins can be artifacts induced during the preparation of samples (solubilization, denaturation, etc.). For this reason, it is important to verify the oligomerization of proteins in living cells. Recently 45 developed energy transfer techniques such as BRET make it possible to follow such protein-protein interactions in living cells. Fusion proteins of OB-RGRP with luciferase and YFP were used to follow OB-RGRP oligomerization in living cells. Coexpression of the OB-RGRP-YFP or YFP-OB-RGRP constructs with the OB-RGRP-Luc construct induces an energy transfer (Fig. 6a). The specificity of this

interaction was shown by the lack of energy transfer during the coexpression with two different fusion proteins: β -arrestine2-YFP (Angers et al. (2000) Proc Natl Acad Sci USA 97, 3684-3689), or melatonin-Luc MT2 receptor (Ayoub et al. (2002) J Biol Chem 277, 21522-21528). We then expressed various ratios of the BRET partners (Fig. 6b). The BRET signal is increased in a hyperbolic manner as a function of the OB-RGRP-YFB/OB-RGRP-Luc ratio, reaching an asymptote which corresponds to saturation of the energy-donor molecules (OB-RGRP-Luc) by the acceptor molecules (OB-RGRP-YFP), which is expected in the case of a specific interaction.

Collectively, these results show that OB-RGRP is a dimeric membrane-bound protein which can also be involved in high molecular weight oligomeric complexes. The 3rd and 4th potential transmembrane domains appear to be important for oligomer formation.

Example 3: Interaction between OB-R and OB-RGRP

We used BRET technology to study a possible interaction between OB-R and OB-RGRP in living cells. An energy transfer was constitutively observed in the cells coexpressing the OB-R_s-Luc construct and the OB-RGRP-YFP construct, indicating proximity of the interaction partners (Fig. 7). The same results were obtained in cells coexpressing OB-R_s-Luc and the MYO47-YFP construct, and also in the reverse orientation: in cells coexpressing OB-RGRP-Luc and OB-R_s-YFP, or in cells coexpressing MYO47-Luc and OB-R_s-YFP. The specificity of these interactions was confirmed by the lack of energy transfer between OB-R_s-Luc, OB-RGRP-Luc, MYO47-Luc and a construct of the insulin receptor tagged with YFP (Boute et al. (2001) Mol Pharmacol 60, 640-645), and also in the reverse orientation: by the lack of energy transfer between a construct of the insulin receptor tagged with Luc and the OB-R_s-YFP, OB-RGRP-YFP and MYO47-YFP construct. Coexpression of OB-R_s-Luc and an OB-RGRP or MYO47 construct exhibiting the YFP tag at the N-terminal produces no significant signal, confirming the specificity of interaction with OB-RGRP-YFP and MYO47, and indicates that the N-terminal end of OB-RGRP and MYO47 must be involved in the interaction with OB-R.

No significant energy transfer was observed in the cells coexpressing the OB-R_i-Luc and OB-RGRP-YFP or YFP-OB-RGRP constructs. This is not due to a lack of functional OB-R_i-Luc expression since a specific BRET signal was observed in cells coexpressing OB-R_i-YFP in order to follow OB-R dimerization. The lack of BRET between the OB-R_i-Luc and OB-RGRP-YFP fusion proteins does not exclude a direct interaction between these two proteins since this may be explained by the fact that the distance between the two BRET partners (Luc and YFP) is greater than 100 Å, the maximum distance for obtaining a transfer. This should be the case since the N- and C-terminal ends of OB-RGRP should be located close to the transmembrane region of OB-R, whereas the C-terminal end of OB-R_i should more probably point toward the cytoplasm due to its long intracellular tail of approximately 300 amino acids. Given that the short and long isoforms of OB-R share the same trans- and juxtamembrane regions and that the interaction of OB-RGRP with OB-R_s is located at this level, it is probable that OB-RGRP interacts with OB-R_i in the same way as with OB-R_s.

Example 4: Effect of the overexpression of OB-RGRP on OB-R signaling

Constructs containing STAT3- or STAT5-response elements upstream of a luciferase reporter gene were coexpressed with OB-R_i in the presence or absence of various OB-RGRP constructs (Fig. 8). The two constructs were activated by leptin in a dose-dependent manner, with an EC₅₀ of approximately 50 pM. Similar results were obtained in HEK 293 cells stably expressing a reporter gene for STAT3. The overexpression of various OB-RGRP constructs had no reproducible effect on this activation, indicating that OB-RGRP is not a limiting factor.

Example 5: Effect of the overexpression of OB-RGRP on the expression of OB-R at the surface

In yeast knockout for OB-RGRP (Vps55), protein transport is disturbed between the golgi and the vacuoles (Belgareh-Touze et al. (2002) Molecular Biology Of The Cell 13, 1694-1708). Although OB-R are activated only when they are expressed at the plasma membrane, a considerable amount of receptors is accumulated in intracellular compartments (Barr, et al. (1999) J Biol Chem, 274, 21416-21424) (Lundin et al. (2000) Biochimica and Biophysica Acta 1499, 130-138). For this reason, we tested the effects of the overexpression of OB-RGRP on the expression of OB-R at the cell surface.

The receptor distribution was studied by ¹²⁵I-leptin-binding experiments. In agreement with other authors (Barr et al., 1999), we showed that only 10 – 20% of the OB-R_i and OB-R_s receptors are expressed at the surface of transfected COS cells (Fig. 9) and of HeLa cells. This is not an artifact due to the expression of exogenous receptors since similar values are obtained in HEK 293 cells expressing endogenous OB-R receptors (Fig. 9). The overexpression of OB-RGRP showed no modification of the total amount in the cells, nor of the % of receptors expressed at the surface (Fig. 9).

Example 6: Characterization of OB-RGRP specific antisense deoxynucleotides

OB-RGRP appears to have ubiquitous expression; for this reason, the decrease in expression of this protein was chosen as an alternative approach for studying its role in OB-R function. Fourteen antisenses specific for OB-RGRP (AS 1 to 14) and two random antisenses (AS 15 and 16) were chosen (see Figure 1), synthesized, and then tested for their ability to inhibit OB-RGRP expression using semiquantitative RT-PCR experiments in HeLa cells expressing OB-RGRP endogenously (Fig. 10). Only one of these antisenses (AS-14), derived from the untranslated 3' region of the OB-RGRP mRNA, interferes with OB-RGRP expression. Labeling of this antisense with the Cy3 fluorophore made it possible to show that all of the cells were transfected under our experimental conditions, in our various experiments.

Example 7: Effect of the OB-RGRP-specific antisense on the signaling and surface expression of the OB-R

HeLa cells were first cotransfected with the expression vectors for OB-R_i and the reporter gene for STAT3, and then with the antisenses. Leptin causes an approximately 1.5-fold increase in the basal activation of the reporter gene for STAT3 in the control cells without antisense, or with a control antisense (AS16) (Fig. 11). In the cells transfected with the antisense specific for OB-RGRP (AS-14), the basal and leptin-stimulated signaling is relatively increased compared to the control conditions. This shows that activation of the JAK/STAT pathway is increased in the cells exhibiting a decrease in OB-RGRP expression. These observations may be explained by an inhibitory effect of OB-RGRP on the basal and OB-R-stimulated activity and, in this case, OB-RGRP can be considered to be a regulator of OB-R signaling. Another alternative is that OB-RGRP might regulate the expression of the surface receptors by limiting the number of OB-R reaching the cell surface. This is in agreement with the fact that only 10 to 20% of the receptors expressed reach the cell surface. In this hypothesis, the decrease in OB-RGRP expression should increase the number of receptors at the cell surface, which should increase the signaling by these receptors. To test this hypothesis, we quantified the number of OB-R_i and OB-R_s receptors expressed at the cell surface in the presence (control) and absence (AS-14) of OB-RGRP (Fig. 12). Transfection of the random antisense showed no effect on the number of receptors expressed at the cell surface, whereas that of the specific antisense (AS-14) caused a 3-fold increase in the number of OB-R expressed at the plasma membrane. Similar results were obtained in nontransfected HeLa cells expressing endogenous receptors. Under these experimental conditions, the total number of receptors, measured by ¹²⁵I-leptin-binding experiments, showed no significant variations.

All our results are consistent with the role of OB-RGRP in yeast, in protein transport. The increase in surface expression of OB-R appears to be involved in the increase in signaling observed. However, we cannot entirely exclude the hypothesis that OB-RGRP directly regulates OB-R activity. The application of specific antisenses directed against OB-RGRP should be useful for increasing OB-R signaling in leptin-related disorders, such as human obesity, in which resistance to leptin is observed, characterized by an unadapted response to this hormone. The increase in expression of the receptors at the cell surface and in their signaling should be important for increasing the response to leptin in the case of human obesity, firstly by increasing leptin transport to the brain across the blood-brain barrier and, secondly, by increasing OB-R signaling in the hypothalamus.

The interaction between OB-RGRP and OB-R_s implies that the action of OB-RGRP takes place via this direct interaction with the receptors and that preventing this interaction may lead to the effects of the specific antisense ODN being reproduced. We propose using the BRET test of the interactions between OB-RGRP and OB-R_s, and MYO47 and OB-R_s, described above, as a test for screening molecules which may modulate this interaction. This test may be carried out either on whole or permeabilized cells coexpressing the proteins from

fusions of the OB-RGRP and OB-R_s, or MYO47 and OB-R_s BRET partners, or on membrane fractions derived from these cells.

CLAIMS

- 5 1. An optionally modified oligonucleotide comprising from 8 to 50 nucleotides which hybridizes specifically to the sequence SEQ ID No. 1 and which inhibits OB-RGRP expression.
2. The oligonucleotide as claimed in claim 1, which promotes the expression of leptin receptors at the cell surface.
- 10 3. The oligonucleotide as claimed in either of claims 1 and 2, which is an antisense oligonucleotide.
4. The oligonucleotide as claimed in one of claims 1 to 3, which comprises a sequence exhibiting at least 60% identity with the
15 sequence SEQ ID No. 2.
5. The oligonucleotide as claimed in one of claims 1 to 3, wherein nucleotides are thioesterified.
- 20 6. The oligonucleotide as claimed in one of claims 1 to 3, wherein nucleotides are 2'-O-methylated.
7. The oligonucleotide as claimed in one of claims 1 to 3, which has a triethylene glycol residue at its 3' end.
- 25 8. The oligonucleotide as claimed in one of claims 1 to 3, which is single-stranded.
9. The oligonucleotide as claimed in one of claims 1 to 8, which
30 comprises a sequence exhibiting at least 60% identity with the sequence SEQ ID No. 2, in which the nucleotides at positions 2, 4, 6, 7, 9, 11, 13, 15, 17, 19 and 20, in the 5' to 3' direction, are thioesterified.
- 35 10. The oligonucleotide as claimed in one of claims 1 to 8, which comprises a sequence exhibiting at least 60% identity with the
 sequence SEQ ID No. 2, in which the nucleotides at positions 1, 2, 3, 4, 5, 16, 17, 18, 19 and 20, in the 5' to 3' direction, are 2'-O-
40 methylated.
11. The oligonucleotide as claimed in one of claims 1 to 10, which is a DNA.
- 45 12. An oligonucleotide of the iRNA type comprising from 15 to 25 nucleotides, which hybridizes specifically to the sequence SEQ ID No. 21 and which inhibits the expression of OB-RGRP.
13. The oligonucleotide as claimed in claim 12, which is a double-stranded RNA.

14. A vector expressing an oligonucleotide as claimed in one of claims 1 to 4 and 12.
- 5 15. A cell containing a vector as claimed in either of claims 13 and 14.
16. A medicinal product containing an oligonucleotide, a vector or a cell as claimed in one of claims 1 to 15.
- 10 17. A pharmaceutical composition containing a pharmacologically active amount of an oligonucleotide, of a vector or of a cell as claimed in one of claims 1 to 15 and pharmaceutically acceptable excipients.
- 15 18. The use of an oligonucleotide, of a vector or a cell as claimed in one of claims 1 to 15, for producing a medicinal product for preventing and/or treating leptin-related pathological conditions.
- 20 19. A fusion protein, which is composed of a sequence exhibiting at least 65% identity with the sequence SEQ ID No. 4, or the sequence SEQ ID No. 16, or of a substantial part of the sequence SEQ ID No. 4 or of the sequence SEQ ID No. 16, and of an energy-donor or energy-acceptor protein, or of a substantial and active part of an energy-donor or energy-acceptor protein.
- 25 20. The fusion protein as claimed in claim 19, wherein the protein is a luciferase.
- 30 21. The fusion protein as claimed in claim 19, wherein the protein is GFP or a mutant of this protein or DsRed.
- 35 22. The fusion protein as claimed in claim 19, wherein the mutant of GFP is YFP, EYFP, wild-type GFP, GFPS65T or Topaz.
23. The fusion protein as claimed in claim 19, which has the sequence SEQ ID No. 6, SEQ ID No. 8, SEQ ID No. 18 or SEQ ID No. 20.
24. A nucleic acid encoding one of the proteins as claimed in one of claims 19 to 23.
- 40 25. The nucleic acid as claimed in claim 24, which has the sequence SEQ ID No. 5, SEQ ID No. 7, SEQ ID No. 17 or SEQ ID No. 19.
26. A nucleic acid which exhibits at least 65% identity with the sequence as claimed in claim 25.
- 45 27. A nucleic acid which hybridizes, under high stringency conditions, with the sequence as claimed in claim 25.
28. A cell comprising a nucleic acid as claimed in one of claims 24 to 27.

29. A cell expressing a protein as claimed in one of claims 19 to 23.

30. A fragment of a cell as claimed in either of claims 28 and 29.

31. A lysate of a cell as claimed in either of claims 28 and 29.

32. A membrane of a cell as claimed in either of claims 28 and 29.

33. A method for determining the modification, by a compound, of the interaction between a protein exhibiting at least 65% identity with the sequence SEQ ID No. 4 or the sequence SEQ ID No. 16, and the leptin receptor, comprising the steps consisting in:

- bringing said compound into contact with a protein exhibiting at least 65% identity with the sequence SEQ ID No. 4 or the sequence SEQ ID No. 16, and the leptin receptor, or cells, or fragments or lysates or membranes of cells, comprising such proteins, and optionally a suitable enzyme substrate, and

- measuring the interaction between the protein exhibiting at least 65% identity with the sequence SEQ ID No. 4 or the sequence SEQ ID No. 16, and the leptin receptor.

34. A method for determining the modification, by a compound, of the interaction between a protein exhibiting at least 65% identity with the sequence SEQ ID No. 4 or the sequence SEQ ID No. 16, and the leptin receptor, comprising the steps consisting in:

- bringing said compound into contact with an energy-donor fusion protein and an energy-acceptor fusion protein, or cells, or fragments or lysates or membranes of cells, comprising such a protein, and optionally a suitable enzyme substrate, and

- measuring the energy transfer.

35. The method as claimed in claim 34, wherein the energy-donor fusion protein is a protein from fusion between the leptin receptor, or a substantial part of the leptin receptor, and luciferase, or a substantial part of luciferase, and the energy-acceptor fusion protein is a fusion protein as claimed in claim 22.

36. The method as claimed in claim 34, wherein the energy-donor fusion protein is a fusion protein as claimed in claim 20, and the energy-acceptor fusion protein is a protein from fusion between the leptin receptor, or a substantial part of the leptin receptor, and YFP, or a substantial part of YFP.

37. The method as claimed in claim 34, wherein the energy transfer measured in the presence of the test compound is compared to that measured in the absence of the test compound.

38. The method as claimed in claim 34, wherein the energy transfer measured in the presence of the test compound and the leptin (or a

ligand of the receptor) is compared to that measured in the presence of the compound in the absence of leptin (or a ligand of the receptor).

5 39. A method for screening or detecting compounds intended for the prevention and/or treatment of leptin-related pathological conditions, comprising the steps consisting in:

10 - bringing said compound into contact with a protein exhibiting at least 65% identity with the sequence SEQ ID No. 4 or the sequence SEQ ID No. 16, and the leptin receptor, or cells, or fragments or lysates or membranes of cells, comprising such proteins, and optionally a suitable enzyme substrate, and
15 - measuring the interaction between a protein exhibiting at least 65% identity with the sequence SEQ ID No. 4 or the sequence SEQ ID No. 16, and the leptin receptor.

15 40. The method as claimed in either of claims 34 and 35, wherein the fusion protein is a protein of sequence SEQ ID No. 6, SEQ ID No. 8, SEQ ID No. 12, SEQ ID No. 14, SEQ ID No. 18 or SEQ ID No. 20.

20 41. The method as claimed in one of claims 33 to 40, wherein the cells are treated with a permeabilizing agent.

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Lys	Val	Tyr	Asp	Pro	Glu	Gln	Arg	Lys	Arg	Met	Ile	Thr	Gly	Pro	Gln	
145					150				155					160		
tgg tgg gcc agg tgc aag cag atg aac gtg ctg gac agc ttc atc aac																528
Trp	Trp	Ala	Arg	Cys	Lys	Gln	Met	Asn	Val	Leu	Asp	Ser	Phe	Ile	Asn	
				165					170					175		
tac tac gac agc gag aag cac gcc gag aac gcc gtg atc ttc ctg cac																576
Tyr	Tyr	Asp	Ser	Glu	Lys	His	Ala	Glu	Asn	Ala	Val	Ile	Phe	Leu	His	
			180					185					190			
ggc aac gcc gct agc agc tac ctg tgg agg cac gtg gtg ccc cac atc																624

Gly	Asn	Ala	Ala	Ser	Ser	Tyr	Leu	Trp	Arg	His	Val	Val	Pro	His	Ile	
		195					200					205				
gag	ccc	gtg	gcc	agg	tgc	atc	atc	ccc	gat	ctg	atc	ggc	atg	ggc	aag	672
Glu	Pro	Val	Ala	Arg	Cys	Ile	Ile	Pro	Asp	Leu	Ile	Gly	Met	Gly	Lys	
	210					215					220					
agc	ggc	aag	agc	ggc	aac	ggc	agc	tac	agg	ctg	ctg	gac	cac	tac	aag	720
Ser	Gly	Lys	Ser	Gly	Asn	Gly	Ser	Tyr	Arg	Leu	Leu	Asp	His	Tyr	Lys	
225					230				235						240	
tac	ctg	acc	gcc	tgg	ttc	gag	ctc	ctg	aac	ctg	ccc	aag	aag	atc	atc	768
Tyr	Leu	Thr	Ala	Trp	Phe	Glu	Leu	Leu	Asn	Leu	Pro	Lys	Lys	Ile	Ile	
				245					250					255		
ttc	gtg	ggc	cac	gac	tgg	ggc	gcc	tgc	ctg	gcc	ttc	cac	tac	agc	tac	816
Phe	Val	Gly	His	Asp	Trp	Gly	Ala	Cys	Leu	Ala	Phe	His	Tyr	Ser	Tyr	
			260					265					270			
gag	cac	cag	gac	aag	atc	aag	gcc	atc	gtg	cac	gcc	gag	agc	gtg	gtg	864
Glu	His	Gln	Asp	Lys	Ile	Lys	Ala	Ile	Val	His	Ala	Glu	Ser	Val	Val	
	275						280					285				
gac	gtg	atc	gag	agc	tgg	gac	gag	tgg	cca	gac	atc	gag	gag	gac	atc	912
Asp	Val	Ile	Glu	Ser	Trp	Asp	Glu	Trp	Pro	Asp	Ile	Glu	Glu	Asp	Ile	
	290					295					300					
gcc	ctg	atc	aag	agc	gag	gag	ggc	gag	aag	atg	gtg	ctg	gag	aac	aac	960
Ala	Leu	Ile	Lys	Ser	Glu	Glu	Gly	Glu	Lys	Met	Val	Leu	Glu	Asn	Asn	
305					310				315					320		
ttc	ttc	gtg	gag	acc	atg	ctg	ccc	agc	aag	atc	atg	aga	aag	ctg	gag	1008
Phe	Phe	Val	Glu	Thr	Met	Leu	Pro	Ser	Lys	Ile	Met	Arg	Lys	Leu	Glu	
				325					330					335		
ccc	gag	gag	ttc	gcc	gcc	tac	ctg	gag	ccc	ttc	aag	gag	aag	ggc	gag	1056
Pro	Glu	Glu	Phe	Ala	Ala	Tyr	Leu	Glu	Pro	Phe	Lys	Glu	Lys	Gly	Glu	
			340					345					350			
gtg	aga	aga	ccc	acc	ctg	agc	tgg	ccc	aga	gag	atc	ccc	ctg	gtg	aag	1104
Val	Arg	Arg	Pro	Thr	Leu	Ser	Trp	Pro	Arg	Glu	Ile	Pro	Leu	Val	Lys	
	355						360					365				
ggc	ggc	aag	ccc	gac	gtg	gtg	cag	atc	gtg	aga	aac	tac	aac	gcc	tac	1152
Gly	Gly	Lys	Pro	Asp	Val	Val	Gln	Ile	Val	Arg	Asn	Tyr	Asn	Ala	Tyr	
	370					375				380						
ctg	aga	gcc	agc	gac	gac	ctg	ccc	aag	atg	ttc	atc	gag	agc	gac	ccc	1200

Leu Arg Ala Ser Asp Asp Leu Pro Lys Met Phe Ile Glu Ser Asp Pro
 385 390 395 400

ggc ttc ttc agc aac gcc atc gtg gag ggc gcc aag aag ttc ccc aac 1248
 Gly Phe Phe Ser Asn Ala Ile Val Glu Gly Ala Lys Lys Phe Pro Asn
 405 410 415

acc gag ttc gtg aag gtg aag ggc ctg cac ttc agc cag gag gac gcc 1296
 Thr Glu Phe Val Lys Val Lys Gly Leu His Phe Ser Gln Glu Asp Ala
 420 425 430

ccc gac gag atg ggc aag tac atc aag agc ttc gtg gag aga gtg ctg 1344
 Pro Asp Glu Met Gly Lys Tyr Ile Lys Ser Phe Val Glu Arg Val Leu
 435 440 445

aag aac gag cag taa 1359
 Lys Asn Glu Gln
 450

<210> 6
 <211> 452
 <212> PRT
 <213> Artificial sequence
 <223> Artificial sequence description: OB. RGRP LUC

<400> 6
 Met Ala Gly Val Lys Ala Leu Val Ala Leu Ser Phe Ser Gly Ala Ile
 1 5 10 15
 Gly Leu Thr Phe Leu Met Leu Gly Cys Ala Leu Glu Asp Tyr Gly Val
 20 25 30
 Tyr Trp Pro Leu Phe Val Leu Ile Phe His Ala Ile Ser Pro Ile Pro
 35 40 45
 His Phe Ile Ala Lys Arg Val Thr Tyr Asp Ser Asp Ala Thr Ser Ser
 50 55 60
 Ala Cys Arg Glu Leu Ala Tyr Phe Phe Thr Thr Gly Ile Val Val Ser
 65 70 75 80
 Ala Phe Gly Phe Pro Val Ile Leu Ala Arg Val Ala Val Ile Lys Trp
 85 90 95
 Gly Ala Cys Gly Leu Val Leu Ala Gly Asn Ala Val Ile Phe Leu Thr
 100 105 110

Ile	Gln	Gly	Phe	Phe	Leu	Ile	Phe	Gly	Arg	Gly	Asp	Asp	Phe	Ser	Trp	115	120	125	
Glu	Gln	Trp	Ile	Pro	Gly	Asp	Pro	Pro	Ala	Arg	Ala	Thr	Met	Thr	Ser	130	135	140	
Lys	Val	Tyr	Asp	Pro	Glu	Gln	Arg	Lys	Arg	Met	Ile	Thr	Gly	Pro	Gln	145	150	155	160
Trp	Trp	Ala	Arg	Cys	Lys	Gln	Met	Asn	Val	Leu	Asp	Ser	Phe	Ile	Asn	165	170	175	
Tyr	Tyr	Asp	Ser	Glu	Lys	His	Ala	Glu	Asn	Ala	Val	Ile	Phe	Leu	His	180	185	190	
Gly	Asn	Ala	Ala	Ser	Ser	Tyr	Leu	Trp	Arg	His	Val	Val	Pro	His	Ile	195	200	205	
Glu	Pro	Val	Ala	Arg	Cys	Ile	Ile	Pro	Asp	Leu	Ile	Gly	Met	Gly	Lys	210	215	220	
Ser	Gly	Lys	Ser	Gly	Asn	Gly	Ser	Tyr	Arg	Leu	Leu	Asp	His	Tyr	Lys	225	230	235	240
Tyr	Leu	Thr	Ala	Trp	Phe	Glu	Leu	Leu	Asn	Leu	Pro	Lys	Lys	Ile	Ile	245	250	255	
Phe	Val	Gly	His	Asp	Trp	Gly	Ala	Cys	Leu	Ala	Phe	His	Tyr	Ser	Tyr	260	265	270	
Glu	His	Gln	Asp	Lys	Ile	Lys	Ala	Ile	Val	His	Ala	Glu	Ser	Val	Val	275	280	285	
Asp	Val	Ile	Glu	Ser	Trp	Asp	Glu	Trp	Pro	Asp	Ile	Glu	Glu	Asp	Ile	290	295	300	
Ala	Leu	Ile	Lys	Ser	Glu	Glu	Gly	Glu	Lys	Met	Val	Leu	Glu	Asn	Asn	305	310	315	320
Phe	Phe	Val	Glu	Thr	Met	Leu	Pro	Ser	Lys	Ile	Met	Arg	Lys	Leu	Glu	325	330	335	
Pro	Glu	Glu	Phe	Ala	Ala	Tyr	Leu	Glu	Pro	Phe	Lys	Glu	Lys	Gly	Glu	340	345	350	
Val	Arg	Arg	Pro	Thr	Leu	Ser	Trp	Pro	Arg	Glu	Ile	Pro	Leu	Val	Lys	355	360	365	

Gly Gly Lys Pro Asp Val Val Gln Ile Val Arg Asn Tyr Asn Ala Tyr
 370 375 380
 Leu Arg Ala Ser Asp Asp Leu Pro Lys Met Phe Ile Glu Ser Asp Pro
 385 390 395 400
 Gly Phe Phe Ser Asn Ala Ile Val Glu Gly Ala Lys Lys Phe Pro Asn
 405 410 415
 Thr Glu Phe Val Lys Val Lys Gly Leu His Phe Ser Gln Glu Asp Ala
 420 425 430
 Pro Asp Glu Met Gly Lys Tyr Ile Lys Ser Phe Val Glu Arg Val Leu
 435 440 445
 Lys Asn Glu Gln
 450

<210> 7
 <211> 1128
 <212> DNA
 <213> Artificial sequence

<220>
 <221> CDS
 <222> (1)..(1128)
 <220>
 <223> Artificial sequence description :OB RGRP YFP

<400> 7
 atg gcg ggc gtt aaa gct ctc gtg gca tta tcc ttc agt ggg gct att 48
 Met Ala Gly Val Lys Ala Leu Val Ala Leu Ser Phe Ser Gly Ala Ile
 1 5 10 15
 gga ctg act ttt ctt atg ctg gga tgt gcc tta gag gat tat ggc gtt 96
 Gly Leu Thr Phe Leu Met Leu Gly Cys Ala Leu Glu Asp Tyr Gly Val
 20 25 30
 tac tgg ccc tta ttc gtc ctg att ttc cac gcc atc tcc ccc atc ccc 144
 Tyr Trp Pro Leu Phe Val Leu Ile Phe His Ala Ile Ser Pro Ile Pro
 35 40 45
 cat ttc att gcc aaa aga gtc acc tat gac tca gat gca acc agt agt 192

His	Phe	Ile	Ala	Lys	Arg	Val	Thr	Tyr	Asp	Ser	Asp	Ala	Thr	Ser	Ser		
50						55					60						
gcc	tgt	cgg	gaa	ctg	gca	tat	ttc	ttc	act	act	gga	att	gtt	gtt	tct	240	
Ala	Cys	Arg	Glu	Leu	Ala	Tyr	Phe	Phe	Thr	Thr	Gly	Ile	Val	Val	Ser		
65					70				75						80		
gcc	ttt	gga	ttt	cct	gtt	att	ctt	gct	cgt	gtg	gct	gtg	atc	aaa	tgg	288	
Ala	Phe	Gly	Phe	Pro	Val	Ile	Leu	Ala	Arg	Val	Ala	Val	Ile	Lys	Trp		
				85					90					95			
gga	gcc	tgc	ggc	ctt	gtg	ttg	gca	ggc	aat	gca	gtc	att	ttc	ctt	aca	336	
Gly	Ala	Cys	Gly	Leu	Val	Leu	Ala	Gly	Asn	Ala	Val	Ile	Phe	Leu	Thr		
			100					105					110				
att	caa	ggg	ttt	ttc	ctt	ata	ttt	gga	aga	gga	gat	gat	ttt	agc	tgg	384	
Ile	Gln	Gly	Phe	Phe	Leu	Ile	Phe	Gly	Arg	Gly	Asp	Asp	Phe	Ser	Trp		
	115					120						125					
gag	cag	tgg	att	ccg	gtc	gcc	acc	atg	gtg	agc	aag	ggc	gag	gag	ctg	432	
Glu	Gln	Trp	Ile	Pro	Val	Ala	Thr	Met	Val	Ser	Lys	Gly	Glu	Glu	Leu		
	130					135					140						
ttc	acc	ggg	gtg	gtg	ccc	atc	ctg	gtc	gag	ctg	gac	ggc	gac	gta	aac	480	
Phe	Thr	Gly	Val	Val	Pro	Ile	Leu	Val	Glu	Leu	Asp	Gly	Asp	Val	Asn		
145					150					155					160		
ggc	cac	aag	ttc	agc	gtg	tcc	ggc	gag	ggc	gag	ggc	gat	gcc	acc	tac	528	
Gly	His	Lys	Phe	Ser	Val	Ser	Gly	Glu	Gly	Glu	Gly	Asp	Ala	Thr	Tyr		
				165				170						175			
ggc	aag	ctg	acc	ctg	aag	ttc	atc	tgc	acc	acc	ggc	aag	ctg	ccc	gtg	576	
Gly	Lys	Leu	Thr	Leu	Lys	Phe	Ile	Cys	Thr	Thr	Gly	Lys	Leu	Pro	Val		
			180					185					190				
ccc	tgg	ccc	acc	ctc	gtg	acc	acc	ttc	ggc	tac	ggc	gtg	cag	tgc	ttc	624	
Pro	Trp	Pro	Thr	Leu	Val	Thr	Thr	Phe	Gly	Tyr	Gly	Val	Gln	Cys	Phe		
	195					200						205					
gcc	cgc	tac	ccc	gac	cac	atg	cgc	cag	cac	gac	ttc	ttc	aag	tcc	gcc	672	
Ala	Arg	Tyr	Pro	Asp	His	Met	Arg	Gln	His	Asp	Phe	Phe	Lys	Ser	Ala		
	210					215				220							
atg	ccc	gaa	ggc	tac	gtc	cag	gag	cgc	acc	atc	ttc	ttc	aag	gac	gac	720	
Met	Pro	Glu	Gly	Tyr	Val	Gln	Glu	Arg	Thr	Ile	Phe	Phe	Lys	Asp	Asp		
225					230					235					240		
ggc	aac	tac	aag	acc	cgc	gcc	gag	gtg	aag	ttc	gag	ggc	gac	acc	ctg	768	

Gly	Asn	Tyr	Lys	Thr	Arg	Ala	Glu	Val	Lys	Phe	Glu	Gly	Asp	Thr	Leu		
				245					250					255			
gtg	aac	cgc	atc	gag	ctg	aag	ggc	atc	gac	ttc	aag	gag	gac	ggc	aac	816	
Val	Asn	Arg	Ile	Glu	Leu	Lys	Gly	Ile	Asp	Phe	Lys	Glu	Asp	Gly	Asn		
			260					265						270			
atc	ctg	ggg	cac	aag	ctg	gag	tac	aac	tac	aac	agc	cac	aac	gtc	tat	864	
Ile	Leu	Gly	His	Lys	Leu	Glu	Tyr	Asn	Tyr	Asn	Ser	His	Asn	Val	Tyr		
			275					280						285			
atc	atg	gcc	gac	aag	cag	aag	aac	ggc	atc	aag	gtg	aac	ttc	aag	atc	912	
Ile	Met	Ala	Asp	Lys	Gln	Lys	Asn	Gly	Ile	Lys	Val	Asn	Phe	Lys	Ile		
			290					295					300				
cgc	cac	aac	atc	gag	gac	ggc	agc	gtg	cag	ctc	gcc	gac	cac	tac	cag	960	
Arg	His	Asn	Ile	Glu	Asp	Gly	Ser	Val	Gln	Leu	Ala	Asp	His	Tyr	Gln		
305						310					315				320		
cag	aac	acc	ccc	atc	ggc	gac	ggc	ccc	gtg	ctg	ctg	ccc	gac	aac	cac	1008	
Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	Leu	Leu	Pro	Asp	Asn	His		
				325					330					335			
tac	ctg	agc	tac	cag	tcc	gcc	ctg	agc	aaa	gac	ccc	aac	gag	aag	cgc	1056	
Tyr	Leu	Ser	Tyr	Gln	Ser	Ala	Leu	Ser	Lys	Asp	Pro	Asn	Glu	Lys	Arg		
				340				345						350			
gat	cac	atg	gtc	ctg	ctg	gag	ttc	gtg	acc	gcc	gcc	ggg	atc	act	ctc	1104	
Asp	His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	Ala	Ala	Gly	Ile	Thr	Leu		
				355				360					365				
ggc	atg	gac	gag	ctg	tac	aag	taa									1128	
Gly	Met	Asp	Glu	Leu	Tyr	Lys											
			370			375											

<210> 8
 <211> 375
 <212> PRT
 <213> Artificial sequence
 <223> Artificial sequence description :OB RGRP YFP

<400> 8
 Met Ala Gly Val Lys Ala Leu Val Ala Leu Ser Phe Ser Gly Ala Ile
 1 5 10 15
 Gly Leu Thr Phe Leu Met Leu Gly Cys Ala Leu Glu Asp Tyr Gly Val

	20		25		30
Tyr Trp Pro Leu Phe Val Leu Ile Phe His Ala Ile Ser Pro Ile Pro					
	35		40		45
His Phe Ile Ala Lys Arg Val Thr Tyr Asp Ser Asp Ala Thr Ser Ser					
	50		55		60
Ala Cys Arg Glu Leu Ala Tyr Phe Phe Thr Thr Gly Ile Val Val Ser					
	65		70		75
Ala Phe Gly Phe Pro Val Ile Leu Ala Arg Val Ala Val Ile Lys Trp					
		85		90	95
Gly Ala Cys Gly Leu Val Leu Ala Gly Asn Ala Val Ile Phe Leu Thr					
		100		105	110
Ile Gln Gly Phe Phe Leu Ile Phe Gly Arg Gly Asp Asp Phe Ser Trp					
		115		120	125
Glu Gln Trp Ile Pro Val Ala Thr Met Val Ser Lys Gly Glu Glu Leu					
	130		135		140
Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn					
	145		150		155
Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr					
		165		170	175
Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val					
		180		185	190
Pro Trp Pro Thr Leu Val Thr Thr Phe Gly Tyr Gly Val Gln Cys Phe					
	195		200		205
Ala Arg Tyr Pro Asp His Met Arg Gln His Asp Phe Phe Lys Ser Ala					
	210		215		220
Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp					
	225		230		235
Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu					
		245		250	255
Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn					
		260		265	270
Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr					

gag aca gct gtt gaa cct aag ttt aat tca agt ggt act cac ttt tct	240
Glu Thr Ala Val Glu Pro Lys Phe Asn Ser Ser Gly Thr His Phe Ser	
65 70 75 80	
aac tta tcc aaa aca act ttc cac tgt tgc ttt cgg agt gag caa gat	288
Asn Leu Ser Lys Thr Thr Phe His Cys Cys Phe Arg Ser Glu Gln Asp	
85 90 95	
aga aac tgc tcc tta tgt gca gac aac att gaa gga aag aca ttt gtt	336
Arg Asn Cys Ser Leu Cys Ala Asp Asn Ile Glu Gly Lys Thr Phe Val	
100 105 110	
tca aca gta aat tct tta gtt ttt caa caa ata gat gca aac tgg aac	384
Ser Thr Val Asn Ser Leu Val Phe Gln Gln Ile Asp Ala Asn Trp Asn	
115 120 125	
ata cag tgc tgg cta aaa gga gac tta aaa tta ttc atc tgt tat gtg	432
Ile Gln Cys Trp Leu Lys Gly Asp Leu Lys Leu Phe Ile Cys Tyr Val	
130 135 140	
gag tca tta ttt aag aat cta ttc agg aat tat aac tat aag gtc cat	480
Glu Ser Leu Phe Lys Asn Leu Phe Arg Asn Tyr Asn Tyr Lys Val His	
145 150 155 160	
ctt tta tat gtt ctg cct gaa gtg tta gaa gat tca cct ctg gtt ccc	528
Leu Leu Tyr Val Leu Pro Glu Val Leu Glu Asp Ser Pro Leu Val Pro	
165 170 175	
caa aaa ggc agt ttt cag atg gtt cac tgc aat tgc agt gtt cat gaa	576
Gln Lys Gly Ser Phe Gln Met Val His Cys Asn Cys Ser Val His Glu	
180 185 190	
tgt tgt gaa tgt ctt gtg cct gtg cca aca gcc aaa ctc aac gac act	624
Cys Cys Glu Cys Leu Val Pro Val Pro Thr Ala Lys Leu Asn Asp Thr	
195 200 205	
ctc ctt atg tgt ttg aaa atc aca tct ggt gga gta att ttc cag tca	672
Leu Leu Met Cys Leu Lys Ile Thr Ser Gly Gly Val Ile Phe Gln Ser	
210 215 220	
cct cta atg tca gtt cag ccc ata aat atg gtg aag cct gat cca cca	720
Pro Leu Met Ser Val Gln Pro Ile Asn Met Val Lys Pro Asp Pro Pro	
225 230 235 240	
tta ggt ttg cat atg gaa atc aca gat gat ggt aat tta aag att tct	768
Leu Gly Leu His Met Glu Ile Thr Asp Asp Gly Asn Leu Lys Ile Ser	
245 250 255	

tgg tcc agc cca cca ttg gta cca ttt cca ctt caa tat caa gtg aaa	816
Trp Ser Ser Pro Pro Leu Val Pro Phe Pro Leu Gln Tyr Gln Val Lys	
260 265 270	
tat tca gag aat tct aca aca gtt atc aga gaa gct gac aag att gtc	864
Tyr Ser Glu Asn Ser Thr Thr Val Ile Arg Glu Ala Asp Lys Ile Val	
275 280 285	
tca gct aca tcc ctg cta gta gac agt ata ctt cct ggg tct tcg tat	912
Ser Ala Thr Ser Leu Leu Val Asp Ser Ile Leu Pro Gly Ser Ser Tyr	
290 295 300	
gag gtt cag gtg agg ggc aag aga ctg gat ggc cca gga atc tgg agt	960
Glu Val Gln Val Arg Gly Lys Arg Leu Asp Gly Pro Gly Ile Trp Ser	
305 310 315 320	
gac tgg agt act cct cgt gtc ttt acc aca caa gat gtc ata tac ttt	1008
Asp Trp Ser Thr Pro Arg Val Phe Thr Thr Gln Asp Val Ile Tyr Phe	
325 330 335	
cca cct aaa att ctg aca agt gtt ggg tct aat gtt tct ttt cac tgc	1056
Pro Pro Lys Ile Leu Thr Ser Val Gly Ser Asn Val Ser Phe His Cys	
340 345 350	
atc tat aag aag gaa aac aag att gtt ccc tca aaa gag att gtt tgg	1104
Ile Tyr Lys Lys Glu Asn Lys Ile Val Pro Ser Lys Glu Ile Val Trp	
355 360 365	
tgg atg aat tta gct gag aaa att cct caa agc cag tat gat gtt gtg	1152
Trp Met Asn Leu Ala Glu Lys Ile Pro Gln Ser Gln Tyr Asp Val Val	
370 375 380	
agt gat cat gtt agc aaa gtt act ttt ttc aat ctg aat gaa acc aaa	1200
Ser Asp His Val Ser Lys Val Thr Phe Phe Asn Leu Asn Glu Thr Lys	
385 390 395 400	
cct cga gga aag ttt acc tat gat gca gtg tac tgc tgc aat gaa cat	1248
Pro Arg Gly Lys Phe Thr Tyr Asp Ala Val Tyr Cys Cys Asn Glu His	
405 410 415	
gaa tgc cat cat cgc tat gct gaa tta tat gtg att gat gtc aat atc	1296
Glu Cys His His Arg Tyr Ala Glu Leu Tyr Val Ile Asp Val Asn Ile	
420 425 430	
aat atc tca tgt gaa act gat ggg tac tta act aaa atg act tgc aga	1344
Asn Ile Ser Cys Glu Thr Asp Gly Tyr Leu Thr Lys Met Thr Cys Arg	
435 440 445	

tgg tca acc agt aca atc cag tca ctt gcg gaa agc act ttg caa ttg Trp Ser Thr Ser Thr Ile Gln Ser Leu Ala Glu Ser Thr Leu Gln Leu 450 455 460	1392
agg tat cat agg agc agc ctt tac tgt tct gat att cca tct att cat Arg Tyr His Arg Ser Ser Leu Tyr Cys Ser Asp Ile Pro Ser Ile His 465 470 475 480	1440
ccc ata tct gag ccc aaa gat tgc tat ttg cag agt gat ggt ttt tat Pro Ile Ser Glu Pro Lys Asp Cys Tyr Leu Gln Ser Asp Gly Phe Tyr 485 490 495	1488
gaa tgc att ttc cag cca atc ttc cta tta tct ggc tac aca atg tgg Glu Cys Ile Phe Gln Pro Ile Phe Leu Leu Ser Gly Tyr Thr Met Trp 500 505 510	1536
att agg atc aat cac tct cta ggt tca ctt gac tct cca cca aca tgt Ile Arg Ile Asn His Ser Leu Gly Ser Leu Asp Ser Pro Pro Thr Cys 515 520 525	1584
gtc ctt cct gat tct gtg gtg aag cca ctg cct cca tcc agt gtg aaa Val Leu Pro Asp Ser Val Val Lys Pro Leu Pro Pro Ser Ser Val Lys 530 535 540	1632
gca gaa att act ata aac att gga tta ttg aaa ata tct tgg gaa aag Ala Glu Ile Thr Ile Asn Ile Gly Leu Leu Lys Ile Ser Trp Glu Lys 545 550 555 560	1680
cca gtc ttt cca gag aat aac ctt caa ttc cag att cgc tat ggt tta Pro Val Phe Pro Glu Asn Asn Leu Gln Phe Gln Ile Arg Tyr Gly Leu 565 570 575	1728
agt gga aaa gaa gta caa tgg aag atg tat gag gtt tat gat gca aaa Ser Gly Lys Glu Val Gln Trp Lys Met Tyr Glu Val Tyr Asp Ala Lys 580 585 590	1776
tca aaa tct gtc agt ctc cca gtt cca gac ttg tgt gca gtc tat gct Ser Lys Ser Val Ser Leu Pro Val Pro Asp Leu Cys Ala Val Tyr Ala 595 600 605	1824
gtt cag gtg cgc tgt aag agg cta gat gga ctg gga tat tgg agt aat Val Gln Val Arg Cys Lys Arg Leu Asp Gly Leu Gly Tyr Trp Ser Asn 610 615 620	1872
tgg agc aat cca gcc tac aca gtt gtc atg gat ata aaa gtt cct atg Trp Ser Asn Pro Ala Tyr Thr Val Val Met Asp Ile Lys Val Pro Met 625 630 635 640	1920

aga gga cct gaa ttt tgg aga ata att aat gga gat act atg aaa aag	1968
Arg Gly Pro Glu Phe Trp Arg Ile Ile Asn Gly Asp Thr Met Lys Lys	
645 650 655	
gag aaa aat gtc act tta ctt tgg aag ccc ctg atg aaa aat gac tca	2016
Glu Lys Asn Val Thr Leu Leu Trp Lys Pro Leu Met Lys Asn Asp Ser	
660 665 670	
ttg tgc agt gtt cag aga tat gtg ata aac cat cat act tcc tgc aat	2064
Leu Cys Ser Val Gln Arg Tyr Val Ile Asn His His Thr Ser Cys Asn	
675 680 685	
gga aca tgg tca gaa gat gtg gga aat cac acg aaa ttc act ttc ctg	2112
Gly Thr Trp Ser Glu Asp Val Gly Asn His Thr Lys Phe Thr Phe Leu	
690 695 700	
tgg aca gag caa gca cat act gtt acg gtt ctg gcc atc aat tca att	2160
Trp Thr Glu Gln Ala His Thr Val Thr Val Leu Ala Ile Asn Ser Ile	
705 710 715 720	
ggt gct tct gtt gca aat ttt aat tta acc ttt tca tgg cct atg agc	2208
Gly Ala Ser Val Ala Asn Phe Asn Leu Thr Phe Ser Trp Pro Met Ser	
725 730 735	
aaa gta aat atc gtg cag tca ctc agt gct tat cct tta aac agc agt	2256
Lys Val Asn Ile Val Gln Ser Leu Ser Ala Tyr Pro Leu Asn Ser Ser	
740 745 750	
tgt gtg att gtt tcc tgg ata cta tca ccc agt gat tac aag cta atg	2304
Cys Val Ile Val Ser Trp Ile Leu Ser Pro Ser Asp Tyr Lys Leu Met	
755 760 765	
tat ttt att att gag tgg aaa aat ctt aat gaa gat ggt gaa ata aaa	2352
Tyr Phe Ile Ile Glu Trp Lys Asn Leu Asn Glu Asp Gly Glu Ile Lys	
770 775 780	
tgg ctt aga atc tct tca tct gtt aag aag tat tat atc cat gat cat	2400
Trp Leu Arg Ile Ser Ser Ser Val Lys Lys Tyr Tyr Ile His Asp His	
785 790 795 800	
ttt atc ccc att gag aag tac cag ttc agt ctt tac cca ata ttt atg	2448
Phe Ile Pro Ile Glu Lys Tyr Gln Phe Ser Leu Tyr Pro Ile Phe Met	
805 810 815	
gaa gga gtg gga aaa cca aag ata att aat agt ttc act caa gat gat	2496
Glu Gly Val Gly Lys Pro Lys Ile Ile Asn Ser Phe Thr Gln Asp Asp	
820 825 830	

att gaa aaa cac cag agt gat gca ggt tta tat gta att gtg cca gta	2544
Ile Glu Lys His Gln Ser Asp Ala Gly Leu Tyr Val Ile Val Pro Val	
835 840 845	
att att tcc tct tcc atc tta ttg ctt gga aca tta tta ata tca cac	2592
Ile Ile Ser Ser Ser Ile Leu Leu Leu Gly Thr Leu Leu Ile Ser His	
850 855 860	
caa aga atg aaa aag cta ttt tgg gaa gat gtt ccg aac ccc aag aat	2640
Gln Arg Met Lys Lys Leu Phe Trp Glu Asp Val Pro Asn Pro Lys Asn	
865 870 875 880	
tgt tcc tgg gca caa gga ctt aat ttt cag aag aga acg gac att ctt	2688
Cys Ser Trp Ala Gln Gly Leu Asn Phe Gln Lys Arg Thr Asp Ile Leu	
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tga	2691

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 <212> PRT
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<400> 10

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Phe Lys Leu Ser Cys Met Pro Pro Asn Ser Thr Tyr Asp Tyr Phe Leu	
35 40 45	
Leu Pro Ala Gly Leu Ser Lys Asn Thr Ser Asn Ser Asn Gly His Tyr	
50 55 60	
Glu Thr Ala Val Glu Pro Lys Phe Asn Ser Ser Gly Thr His Phe Ser	
65 70 75 80	
Asn Leu Ser Lys Thr Thr Phe His Cys Cys Phe Arg Ser Glu Gln Asp	
85 90 95	
Arg Asn Cys Ser Leu Cys Ala Asp Asn Ile Glu Gly Lys Thr Phe Val	
100 105 110	
Ser Thr Val Asn Ser Leu Val Phe Gln Gln Ile Asp Ala Asn Trp Asn	

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Ile Gln Cys Trp Leu Lys Gly Asp Leu Lys Leu Phe Ile Cys Tyr Val		
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Glu Ser Leu Phe Lys Asn Leu Phe Arg Asn Tyr Asn Tyr Lys Val His		
145	150	155
Leu Leu Tyr Val Leu Pro Glu Val Leu Glu Asp Ser Pro Leu Val Pro		
165	170	175
Gln Lys Gly Ser Phe Gln Met Val His Cys Asn Cys Ser Val His Glu		
180	185	190
Cys Cys Glu Cys Leu Val Pro Val Pro Thr Ala Lys Leu Asn Asp Thr		
195	200	205
Leu Leu Met Cys Leu Lys Ile Thr Ser Gly Gly Val Ile Phe Gln Ser		
210	215	220
Pro Leu Met Ser Val Gln Pro Ile Asn Met Val Lys Pro Asp Pro Pro		
225	230	235
Leu Gly Leu His Met Glu Ile Thr Asp Asp Gly Asn Leu Lys Ile Ser		
245	250	255
Trp Ser Ser Pro Pro Leu Val Pro Phe Pro Leu Gln Tyr Gln Val Lys		
260	265	270
Tyr Ser Glu Asn Ser Thr Thr Val Ile Arg Glu Ala Asp Lys Ile Val		
275	280	285
Ser Ala Thr Ser Leu Leu Val Asp Ser Ile Leu Pro Gly Ser Ser Tyr		
290	295	300
Glu Val Gln Val Arg Gly Lys Arg Leu Asp Gly Pro Gly Ile Trp Ser		
305	310	315
Asp Trp Ser Thr Pro Arg Val Phe Thr Thr Gln Asp Val Ile Tyr Phe		
325	330	335
Pro Pro Lys Ile Leu Thr Ser Val Gly Ser Asn Val Ser Phe His Cys		
340	345	350
Ile Tyr Lys Lys Glu Asn Lys Ile Val Pro Ser Lys Glu Ile Val Trp		
355	360	365
Trp Met Asn Leu Ala Glu Lys Ile Pro Gln Ser Gln Tyr Asp Val Val		

370		375		380
Ser Asp His Val Ser Lys Val Thr Phe Phe Asn Leu Asn Glu Thr Lys				
385		390		400
Pro Arg Gly Lys Phe Thr Tyr Asp Ala Val Tyr Cys Cys Asn Glu His				
	405		410	415
Glu Cys His His Arg Tyr Ala Glu Leu Tyr Val Ile Asp Val Asn Ile				
	420		425	430
Asn Ile Ser Cys Glu Thr Asp Gly Tyr Leu Thr Lys Met Thr Cys Arg				
	435		440	445
Trp Ser Thr Ser Thr Ile Gln Ser Leu Ala Glu Ser Thr Leu Gln Leu				
	450		455	460
Arg Tyr His Arg Ser Ser Leu Tyr Cys Ser Asp Ile Pro Ser Ile His				
	465		470	475
Pro Ile Ser Glu Pro Lys Asp Cys Tyr Leu Gln Ser Asp Gly Phe Tyr				
	485		490	495
Glu Cys Ile Phe Gln Pro Ile Phe Leu Leu Ser Gly Tyr Thr Met Trp				
	500		505	510
Ile Arg Ile Asn His Ser Leu Gly Ser Leu Asp Ser Pro Pro Thr Cys				
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Val Leu Pro Asp Ser Val Val Lys Pro Leu Pro Pro Ser Ser Val Lys				
	530		535	540
Ala Glu Ile Thr Ile Asn Ile Gly Leu Leu Lys Ile Ser Trp Glu Lys				
	545		550	555
Pro Val Phe Pro Glu Asn Asn Leu Gln Phe Gln Ile Arg Tyr Gly Leu				
	565		570	575
Ser Gly Lys Glu Val Gln Trp Lys Met Tyr Glu Val Tyr Asp Ala Lys				
	580		585	590
Ser Lys Ser Val Ser Leu Pro Val Pro Asp Leu Cys Ala Val Tyr Ala				
	595		600	605
Val Gln Val Arg Cys Lys Arg Leu Asp Gly Leu Gly Tyr Trp Ser Asn				
	610		615	620
Trp Ser Asn Pro Ala Tyr Thr Val Val Met Asp Ile Lys Val Pro Met				

625		630		635		640
Arg Gly Pro Glu Phe Trp Arg Ile Ile Asn Gly Asp Thr Met Lys Lys						
	645		650		655	
Glu Lys Asn Val Thr Leu Leu Trp Lys Pro Leu Met Lys Asn Asp Ser						
	660		665		670	
Leu Cys Ser Val Gln Arg Tyr Val Ile Asn His His Thr Ser Cys Asn						
	675		680		685	
Gly Thr Trp Ser Glu Asp Val Gly Asn His Thr Lys Phe Thr Phe Leu						
	690		695		700	
Trp Thr Glu Gln Ala His Thr Val Thr Val Leu Ala Ile Asn Ser Ile						
705		710		715		720
Gly Ala Ser Val Ala Asn Phe Asn Leu Thr Phe Ser Trp Pro Met Ser						
	725		730		735	
Lys Val Asn Ile Val Gln Ser Leu Ser Ala Tyr Pro Leu Asn Ser Ser						
	740		745		750	
Cys Val Ile Val Ser Trp Ile Leu Ser Pro Ser Asp Tyr Lys Leu Met						
	755		760		765	
Tyr Phe Ile Ile Glu Trp Lys Asn Leu Asn Glu Asp Gly Glu Ile Lys						
	770		775		780	
Trp Leu Arg Ile Ser Ser Ser Val Lys Lys Tyr Tyr Ile His Asp His						
785		790		795		800
Phe Ile Pro Ile Glu Lys Tyr Gln Phe Ser Leu Tyr Pro Ile Phe Met						
	805		810		815	
Glu Gly Val Gly Lys Pro Lys Ile Ile Asn Ser Phe Thr Gln Asp Asp						
	820		825		830	
Ile Glu Lys His Gln Ser Asp Ala Gly Leu Tyr Val Ile Val Pro Val						
	835		840		845	
Ile Ile Ser Ser Ser Ile Leu Leu Leu Gly Thr Leu Leu Ile Ser His						
	850		855		860	
Gln Arg Met Lys Lys Leu Phe Trp Glu Asp Val Pro Asn Pro Lys Asn						
865		870		875		880
Cys Ser Trp Ala Gln Gly Leu Asn Phe Gln Lys Arg Thr Asp Ile Leu						

acg aca ttt gtt tca aca gta aat tct tta gtt ttt caa caa ata gat	432
Thr Thr Phe Val Ser Thr Val Asn Ser Leu Val Phe Gln Gln Ile Asp	
130 135 140	
gca aac tgg aac ata cag tgc tgg cta aaa gga gac tta aaa tta ttc	480
Ala Asn Trp Asn Ile Gln Cys Trp Leu Lys Gly Asp Leu Lys Leu Phe	
145 150 155 160	
atc tgt tat gtg gag tca tta ttt aag aat cta ttc agg aat tat aac	528
Ile Cys Tyr Val Glu Ser Leu Phe Lys Asn Leu Phe Arg Asn Tyr Asn	
165 170 175	
tat aag gtc cat ctt tta tat gtt ctg cct gaa gtg tta gaa gat tca	576
Tyr Lys Val His Leu Leu Tyr Val Leu Pro Glu Val Leu Glu Asp Ser	
180 185 190	
cct ctg gtt ccc caa aaa ggc agt ttt cag atg gtt cac tgc aat tgc	624
Pro Leu Val Pro Gln Lys Gly Ser Phe Gln Met Val His Cys Asn Cys	
195 200 205	
agt gtt cat gaa tgt tgt gaa tgt ctt gtg cct gtg cca aca gcc aaa	672
Ser Val His Glu Cys Cys Glu Cys Leu Val Pro Val Pro Thr Ala Lys	
210 215 220	
ctc aac gac act ctc ctt atg tgt ttg aaa atc aca tct ggt gga gta	720
Leu Asn Asp Thr Leu Leu Met Cys Leu Lys Ile Thr Ser Gly Gly Val	
225 230 235 240	
att ttc cgg tca cct cta atg tca gtt cag ccc ata aat atg gtg aag	768
Ile Phe Arg Ser Pro Leu Met Ser Val Gln Pro Ile Asn Met Val Lys	
245 250 255	
cct gat cca cca tta ggt ttg cat atg gaa atc aca gat gat ggt aat	816
Pro Asp Pro Pro Leu Gly Leu His Met Glu Ile Thr Asp Asp Gly Asn	
260 265 270	
tta aag att tct tgg tcc agc cca cca ttg gta cca ttt cca ctt caa	864
Leu Lys Ile Ser Trp Ser Ser Pro Pro Leu Val Pro Phe Pro Leu Gln	
275 280 285	
tat caa gtg aaa tat tca gag aat tct aca aca gtt atc aga gaa gct	912
Tyr Gln Val Lys Tyr Ser Glu Asn Ser Thr Thr Val Ile Arg Glu Ala	
290 295 300	
gac aag att gtc tca gct aca tcc ctg cta gta gac agt ata ctt cct	960
Asp Lys Ile Val Ser Ala Thr Ser Leu Leu Val Asp Ser Ile Leu Pro	
305 310 315 320	

ggg tct tgg tat gag gtt cag gtg agg ggc aag aga ctg gat ggc cca	1008
Gly Ser Ser Tyr Glu Val Gln Val Arg Gly Lys Arg Leu Asp Gly Pro	
325 330 335	
gga atc tgg agt gac tgg agt act cct cgt gtc ttt acc aca caa gat	1056
Gly Ile Trp Ser Asp Trp Ser Thr Pro Arg Val Phe Thr Thr Gln Asp	
340 345 350	
gtc ata tac ttt cca cct aaa att ctg aca agt gtt ggg tct aat gtt	1104
Val Ile Tyr Phe Pro Pro Lys Ile Leu Thr Ser Val Gly Ser Asn Val	
355 360 365	
tct ttt cac tgc atc tat aag aag gaa aac aag att gtt ccc tca aaa	1152
Ser Phe His Cys Ile Tyr Lys Lys Glu Asn Lys Ile Val Pro Ser Lys	
370 375 380	
gag att gtt tgg tgg atg aat tta gct gag aaa att cct caa agc cag	1200
Glu Ile Val Trp Trp Met Asn Leu Ala Glu Lys Ile Pro Gln Ser Gln	
385 390 395 400	
tat gat gtt gtg agt gat cat gtt agc aaa gtt act ttt ttc aat ctg	1248
Tyr Asp Val Val Ser Asp His Val Ser Lys Val Thr Phe Phe Asn Leu	
405 410 415	
aat gaa acc aaa cct cga gga aag ttt acc tat gat gca gtg tac tgc	1296
Asn Glu Thr Lys Pro Arg Gly Lys Phe Thr Tyr Asp Ala Val Tyr Cys	
420 425 430	
tgc aat gaa cat gaa tgc cat cat cgc tat gct gaa tta tat gtg att	1344
Cys Asn Glu His Glu Cys His His Arg Tyr Ala Glu Leu Tyr Val Ile	
435 440 445	
gat gtc aat atc aat atc tca tgt gaa act gat ggg tac tta act aaa	1392
Asp Val Asn Ile Asn Ile Ser Cys Glu Thr Asp Gly Tyr Leu Thr Lys	
450 455 460	
atg act tgc aga tgg tca acc agt aca atc cag tca ctt gcg gaa agc	1440
Met Thr Cys Arg Trp Ser Thr Ser Thr Ile Gln Ser Leu Ala Glu Ser	
465 470 475 480	
act ttg caa ttg agg tat cat agg agc agc ctt tac tgt tct gat att	1488
Thr Leu Gln Leu Arg Tyr His Arg Ser Ser Leu Tyr Cys Ser Asp Ile	
485 490 495	
cca tct att cat ccc ata tct gag ccc aaa gat tgc tat ttg cag agt	1536
Pro Ser Ile His Pro Ile Ser Glu Pro Lys Asp Cys Tyr Leu Gln Ser	
500 505 510	

gat ggt ttt tat gaa tgc att ttc cag cca atc ttc cta tta tct ggc	1584
Asp Gly Phe Tyr Glu Cys Ile Phe Gln Pro Ile Phe Leu Leu Ser Gly	
515 520 525	
tac aca atg tgg att agg atc aat cac tct cta ggt tca ctt gac tct	1632
Tyr Thr Met Trp Ile Arg Ile Asn His Ser Leu Gly Ser Leu Asp Ser	
530 535 540	
cca cca aca tgt gtc ctt cct gat tct gtg gtg aag cca ctg cct cca	1680
Pro Pro Thr Cys Val Leu Pro Asp Ser Val Val Lys Pro Leu Pro Pro	
545 550 555 560	
tcc agt gtg aaa gca gaa att act ata aac att gga tta ttg aaa ata	1728
Ser Ser Val Lys Ala Glu Ile Thr Ile Asn Ile Gly Leu Leu Lys Ile	
565 570 575	
tct tgg gaa aag cca gtc ttt cca gag aat aac ctt caa ttc cag att	1776
Ser Trp Glu Lys Pro Val Phe Pro Glu Asn Asn Leu Gln Phe Gln Ile	
580 585 590	
cgc tat ggt tta agt gga aaa gaa gta caa tgg aag atg tat gag gtt	1824
Arg Tyr Gly Leu Ser Gly Lys Glu Val Gln Trp Lys Met Tyr Glu Val	
595 600 605	
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Tyr Asp Ala Lys Ser Lys Ser Val Ser Leu Pro Val Pro Asp Leu Cys	
610 615 620	
gca gtc tat gct gtt cag gtg cgc tgt aag agg cta gat gga ctg gga	1920
Ala Val Tyr Ala Val Gln Val Arg Cys Lys Arg Leu Asp Gly Leu Gly	
625 630 635 640	
tat tgg agt aat tgg agc aat cca gcc tac aca gtt gtc atg gat ata	1968
Tyr Trp Ser Asn Trp Ser Asn Pro Ala Tyr Thr Val Val Met Asp Ile	
645 650 655	
aaa gtt cct atg aga gga cct gaa ttt tgg aga ata att aat gga gat	2016
Lys Val Pro Met Arg Gly Pro Glu Phe Trp Arg Ile Ile Asn Gly Asp	
660 665 670	
act atg aaa aag gag aaa aat gtc act tta ctt tgg aag ccc ctg atg	2064
Thr Met Lys Lys Glu Lys Asn Val Thr Leu Leu Trp Lys Pro Leu Met	
675 680 685	
aaa aat gac tca ttg tgc agt gtt cag aga tat gtg ata aac cat cat	2112
Lys Asn Asp Ser Leu Cys Ser Val Gln Arg Tyr Val Ile Asn His His	
690 695 700	

act	tcc	tgc	aat	gga	aca	tgg	tca	gaa	gat	gtg	gga	aat	cac	acg	aaa	2160
Thr	Ser	Cys	Asn	Gly	Thr	Trp	Ser	Glu	Asp	Val	Gly	Asn	His	Thr	Lys	
705					710					715					720	
ttc	act	ttc	ctg	tgg	aca	gag	caa	gca	cat	act	gtt	acg	gtt	ctg	gcc	2208
Phe	Thr	Phe	Leu	Trp	Thr	Glu	Gln	Ala	His	Thr	Val	Thr	Val	Leu	Ala	
				725					730						735	
atc	aat	tca	att	ggc	gct	tct	gtt	gca	aat	ttt	aat	tta	acc	ttt	tca	2256
Ile	Asn	Ser	Ile	Gly	Ala	Ser	Val	Ala	Asn	Phe	Asn	Leu	Thr	Phe	Ser	
			740					745					750			
tgg	cct	atg	agc	aaa	gta	aat	atc	gtg	cag	tca	ctc	agt	gct	tat	cct	2304
Trp	Pro	Met	Ser	Lys	Val	Asn	Ile	Val	Gln	Ser	Leu	Ser	Ala	Tyr	Pro	
		755					760					765				
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Leu	Asn	Ser	Ser	Cys	Val	Ile	Val	Ser	Trp	Ile	Leu	Ser	Pro	Ser	Asp	
	770					775					780					
tac	aag	cta	atg	tat	ttt	att	att	gag	tgg	aaa	aat	ctt	aat	gaa	gat	2400
Tyr	Lys	Leu	Met	Tyr	Phe	Ile	Ile	Glu	Trp	Lys	Asn	Leu	Asn	Glu	Asp	
785					790					795					800	
ggc	gaa	ata	aaa	tgg	ctt	aga	atc	tct	tca	tct	gtt	aag	aag	tat	tat	2448
Gly	Glu	Ile	Lys	Trp	Leu	Arg	Ile	Ser	Ser	Ser	Val	Lys	Lys	Tyr	Tyr	
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atc	cat	gat	cat	ttt	atc	ccc	att	gag	aag	tac	cag	ttc	agt	ctt	tac	2496
Ile	His	Asp	His	Phe	Ile	Pro	Ile	Glu	Lys	Tyr	Gln	Phe	Ser	Leu	Tyr	
			820					825					830			
cca	ata	ttt	atg	gaa	gga	gtg	gga	aaa	cca	aag	ata	att	aat	agt	ttc	2544
Pro	Ile	Phe	Met	Glu	Gly	Val	Gly	Lys	Pro	Lys	Ile	Ile	Asn	Ser	Phe	
		835					840					845				
act	caa	gat	gat	att	gaa	aaa	cac	cag	agt	gat	gca	ggc	tta	tat	gta	2592
Thr	Gln	Asp	Asp	Ile	Glu	Lys	His	Gln	Ser	Asp	Ala	Gly	Leu	Tyr	Val	
	850					855					860					
att	gtg	cca	gta	att	att	tcc	tct	tcc	atc	tta	ttg	ctt	gga	aca	tta	2640
Ile	Val	Pro	Val	Ile	Ile	Ser	Ser	Ser	Ile	Leu	Leu	Leu	Gly	Thr	Leu	
865					870					875					880	
tta	ata	tca	cac	caa	aga	atg	aaa	aag	cta	ttt	tgg	gaa	gat	gtt	ccg	2688
Leu	Ile	Ser	His	Gln	Arg	Met	Lys	Lys	Leu	Phe	Trp	Glu	Asp	Val	Pro	
				885					890					895		

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Asn Pro Lys Asn Cys Ser Trp Ala Gln Gly Leu Asn Phe Gln Lys Arg	
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acg gac att ctg gat cca ccg gct aga gcc acc atg acc agc aag gtg	2784
Thr Asp Ile Leu Asp Pro Pro Ala Arg Ala Thr Met Thr Ser Lys Val	
915 920 925	
tac gac ccc gag cag agg aag agg atg atc acc ggc ccc cag tgg tgg	2832
Tyr Asp Pro Glu Gln Arg Lys Arg Met Ile Thr Gly Pro Gln Trp Trp	
930 935 940	
gcc agg tgc aag cag atg aac gtg ctg gac agc ttc atc aac tac tac	2880
Ala Arg Cys Lys Gln Met Asn Val Leu Asp Ser Phe Ile Asn Tyr Tyr	
945 950 955 960	
gac agc gag aag cac gcc gag aac gcc gtg atc ttc ctg cac ggc aac	2928
Asp Ser Glu Lys His Ala Glu Asn Ala Val Ile Phe Leu His Gly Asn	
965 970 975	
gcc gct agc agc tac ctg tgg agg cac gtg gtg ccc cac atc gag ccc	2976
Ala Ala Ser Ser Tyr Leu Trp Arg His Val Val Pro His Ile Glu Pro	
980 985 990	
gtg gcc agg tgc atc atc ccc gat ctg atc ggc atg ggc aag agc ggc	3024
Val Ala Arg Cys Ile Ile Pro Asp Leu Ile Gly Met Gly Lys Ser Gly	
995 1000 1005	
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Lys Ser Gly Asn Gly Ser Tyr Arg Leu Leu Asp His Tyr Lys Tyr Leu	
1010 1015 1020	
acc gcc tgg ttc gag ctc ctg aac ctg ccc aag aag atc atc ttc gtg	3120
Thr Ala Trp Phe Glu Leu Leu Asn Leu Pro Lys Lys Ile Ile Phe Val	
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Gly His Asp Trp Gly Ala Cys Leu Ala Phe His Tyr Ser Tyr Glu His	
1045 1050 1055	
cag gac aag atc aag gcc atc gtg cac gcc gag agc gtg gtg gac gtg	3216
Gln Asp Lys Ile Lys Ala Ile Val His Ala Glu Ser Val Val Asp Val	
1060 1065 1070	
atc gag agc tgg gac gag tgg cca gac atc gag gag gac atc gcc ctg	3264
Ile Glu Ser Trp Asp Glu Trp Pro Asp Ile Glu Glu Asp Ile Ala Leu	
1075 1080 1085	

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Ile Lys Ser Glu Glu Gly Glu Lys Met Val Leu Glu Asn Asn Phe Phe	
1090 1095 1100	
gtg gag acc atg ctg ccc agc aag atc atg aga aag ctg gag ccc gag	3360
Val Glu Thr Met Leu Pro Ser Lys Ile Met Arg Lys Leu Glu Pro Glu	
1105 1110 1115 1120	
gag ttc gcc gcc tac ctg gag ccc ttc aag gag aag ggc gag gtg aga	3408
Glu Phe Ala Ala Tyr Leu Glu Pro Phe Lys Glu Lys Gly Glu Val Arg	
1125 1130 1135	
aga ccc acc ctg agc tgg ccc aga gag atc ccc ctg gtg aag ggc ggc	3456
Arg Pro Thr Leu Ser Trp Pro Arg Glu Ile Pro Leu Val Lys Gly Gly	
1140 1145 1150	
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Lys Pro Asp Val Val Gln Ile Val Arg Asn Tyr Asn Ala Tyr Leu Arg	
1155 1160 1165	
gcc agc gac gac ctg ccc aag atg ttc atc gag agc gac ccc ggc ttc	3552
Ala Ser Asp Asp Leu Pro Lys Met Phe Ile Glu Ser Asp Pro Gly Phe	
1170 1175 1180	
ttc agc aac gcc atc gtg gag ggc gcc aag aag ttc ccc aac acc gag	3600
Phe Ser Asn Ala Ile Val Glu Gly Ala Lys Lys Phe Pro Asn Thr Glu	
1185 1190 1195 1200	
ttc gtg aag gtg aag ggc ctg cac ttc agc cag gag gac gcc ccc gac	3648
Phe Val Lys Val Lys Gly Leu His Phe Ser Gln Glu Asp Ala Pro Asp	
1205 1210 1215	
gag atg ggc aag tac atc aag agc ttc gtg gag aga gtg ctg aag aac	3696
Glu Met Gly Lys Tyr Ile Lys Ser Phe Val Glu Arg Val Leu Lys Asn	
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Glu Gln	
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<210> 12

<211> 1234

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<213> Artificial sequence

<223> Artificial sequence description : OBR LUC

<400> 12

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Leu	Leu	Met	Leu	Phe	His	Leu	Gly	Leu	Gln	Ala	Ser	Ile	Ser	Ala	Arg
			20					25					30		
Gln	Glu	Gln	Lys	Leu	Ile	Ser	Glu	Glu	Asp	Leu	Thr	Arg	Tyr	Pro	Ile
		35					40					45			
Thr	Pro	Trp	Arg	Phe	Lys	Leu	Ser	Cys	Met	Pro	Pro	Asn	Ser	Thr	Tyr
	50					55						60			
Asp	Tyr	Phe	Leu	Leu	Pro	Ala	Gly	Leu	Ser	Lys	Asn	Thr	Ser	Asn	Ser
65					70					75					80
Asn	Gly	His	Tyr	Glu	Thr	Ala	Val	Glu	Pro	Lys	Phe	Asn	Ser	Ser	Gly
				85					90					95	
Thr	His	Phe	Ser	Asn	Leu	Ser	Lys	Thr	Thr	Phe	His	Cys	Cys	Phe	Arg
			100					105					110		
Ser	Glu	Gln	Asp	Arg	Asn	Cys	Ser	Leu	Cys	Ala	Asp	Asn	Ile	Glu	Gly
		115					120					125			
Thr	Thr	Phe	Val	Ser	Thr	Val	Asn	Ser	Leu	Val	Phe	Gln	Gln	Ile	Asp
	130					135					140				
Ala	Asn	Trp	Asn	Ile	Gln	Cys	Trp	Leu	Lys	Gly	Asp	Leu	Lys	Leu	Phe
145					150					155					160
Ile	Cys	Tyr	Val	Glu	Ser	Leu	Phe	Lys	Asn	Leu	Phe	Arg	Asn	Tyr	Asn
				165					170					175	
Tyr	Lys	Val	His	Leu	Leu	Tyr	Val	Leu	Pro	Glu	Val	Leu	Glu	Asp	Ser
			180					185					190		
Pro	Leu	Val	Pro	Gln	Lys	Gly	Ser	Phe	Gln	Met	Val	His	Cys	Asn	Cys
		195					200					205			
Ser	Val	His	Glu	Cys	Cys	Glu	Cys	Leu	Val	Pro	Val	Pro	Thr	Ala	Lys
	210					215					220				
Leu	Asn	Asp	Thr	Leu	Leu	Met	Cys	Leu	Lys	Ile	Thr	Ser	Gly	Gly	Val
225					230					235					240
Ile	Phe	Arg	Ser	Pro	Leu	Met	Ser	Val	Gln	Pro	Ile	Asn	Met	Val	Lys
				245					250					255	

Pro	Asp	Pro	Pro	Leu	Gly	Leu	His	Met	Glu	Ile	Thr	Asp	Asp	Gly	Asn	260	265	270
Leu	Lys	Ile	Ser	Trp	Ser	Ser	Pro	Pro	Leu	Val	Pro	Phe	Pro	Leu	Gln	275	280	285
Tyr	Gln	Val	Lys	Tyr	Ser	Glu	Asn	Ser	Thr	Thr	Val	Ile	Arg	Glu	Ala	290	295	300
Asp	Lys	Ile	Val	Ser	Ala	Thr	Ser	Leu	Leu	Val	Asp	Ser	Ile	Leu	Pro	305	310	315
Gly	Ser	Ser	Tyr	Glu	Val	Gln	Val	Arg	Gly	Lys	Arg	Leu	Asp	Gly	Pro	325	330	335
Gly	Ile	Trp	Ser	Asp	Trp	Ser	Thr	Pro	Arg	Val	Phe	Thr	Thr	Gln	Asp	340	345	350
Val	Ile	Tyr	Phe	Pro	Pro	Lys	Ile	Leu	Thr	Ser	Val	Gly	Ser	Asn	Val	355	360	365
Ser	Phe	His	Cys	Ile	Tyr	Lys	Lys	Glu	Asn	Lys	Ile	Val	Pro	Ser	Lys	370	375	380
Glu	Ile	Val	Trp	Trp	Met	Asn	Leu	Ala	Glu	Lys	Ile	Pro	Gln	Ser	Gln	385	390	395
Tyr	Asp	Val	Val	Ser	Asp	His	Val	Ser	Lys	Val	Thr	Phe	Phe	Asn	Leu	405	410	415
Asn	Glu	Thr	Lys	Pro	Arg	Gly	Lys	Phe	Thr	Tyr	Asp	Ala	Val	Tyr	Cys	420	425	430
Cys	Asn	Glu	His	Glu	Cys	His	His	Arg	Tyr	Ala	Glu	Leu	Tyr	Val	Ile	435	440	445
Asp	Val	Asn	Ile	Asn	Ile	Ser	Cys	Glu	Thr	Asp	Gly	Tyr	Leu	Thr	Lys	450	455	460
Met	Thr	Cys	Arg	Trp	Ser	Thr	Ser	Thr	Ile	Gln	Ser	Leu	Ala	Glu	Ser	465	470	475
Thr	Leu	Gln	Leu	Arg	Tyr	His	Arg	Ser	Ser	Leu	Tyr	Cys	Ser	Asp	Ile	485	490	495
Pro	Ser	Ile	His	Pro	Ile	Ser	Glu	Pro	Lys	Asp	Cys	Tyr	Leu	Gln	Ser	500	505	510

Asp	Gly	Phe	Tyr	Glu	Cys	Ile	Phe	Gln	Pro	Ile	Phe	Leu	Leu	Ser	Gly	515	520	525	
Tyr	Thr	Met	Trp	Ile	Arg	Ile	Asn	His	Ser	Leu	Gly	Ser	Leu	Asp	Ser	530	535	540	
Pro	Pro	Thr	Cys	Val	Leu	Pro	Asp	Ser	Val	Val	Lys	Pro	Leu	Pro	Pro	545	550	555	560
Ser	Ser	Val	Lys	Ala	Glu	Ile	Thr	Ile	Asn	Ile	Gly	Leu	Leu	Lys	Ile	565	570	575	
Ser	Trp	Glu	Lys	Pro	Val	Phe	Pro	Glu	Asn	Asn	Leu	Gln	Phe	Gln	Ile	580	585	590	
Arg	Tyr	Gly	Leu	Ser	Gly	Lys	Glu	Val	Gln	Trp	Lys	Met	Tyr	Glu	Val	595	600	605	
Tyr	Asp	Ala	Lys	Ser	Lys	Ser	Val	Ser	Leu	Pro	Val	Pro	Asp	Leu	Cys	610	615	620	
Ala	Val	Tyr	Ala	Val	Gln	Val	Arg	Cys	Lys	Arg	Leu	Asp	Gly	Leu	Gly	625	630	635	640
Tyr	Trp	Ser	Asn	Trp	Ser	Asn	Pro	Ala	Tyr	Thr	Val	Val	Met	Asp	Ile	645	650	655	
Lys	Val	Pro	Met	Arg	Gly	Pro	Glu	Phe	Trp	Arg	Ile	Ile	Asn	Gly	Asp	660	665	670	
Thr	Met	Lys	Lys	Glu	Lys	Asn	Val	Thr	Leu	Leu	Trp	Lys	Pro	Leu	Met	675	680	685	
Lys	Asn	Asp	Ser	Leu	Cys	Ser	Val	Gln	Arg	Tyr	Val	Ile	Asn	His	His	690	695	700	
Thr	Ser	Cys	Asn	Gly	Thr	Trp	Ser	Glu	Asp	Val	Gly	Asn	His	Thr	Lys	705	710	715	720
Phe	Thr	Phe	Leu	Trp	Thr	Glu	Gln	Ala	His	Thr	Val	Thr	Val	Leu	Ala	725	730	735	
Ile	Asn	Ser	Ile	Gly	Ala	Ser	Val	Ala	Asn	Phe	Asn	Leu	Thr	Phe	Ser	740	745	750	
Trp	Pro	Met	Ser	Lys	Val	Asn	Ile	Val	Gln	Ser	Leu	Ser	Ala	Tyr	Pro	755	760	765	

Thr Ala Trp Phe Glu Leu Leu Asn Leu Pro Lys Lys Ile Ile Phe Val
025 1030 1035 1040

Gly His Asp Trp Gly Ala Cys Leu Ala Phe His Tyr Ser Tyr Glu His
1045 1050 1055

Gln Asp Lys Ile Lys Ala Ile Val His Ala Glu Ser Val Val Asp Val
1060 1065 1070

Ile Glu Ser Trp Asp Glu Trp Pro Asp Ile Glu Glu Asp Ile Ala Leu
1075 1080 1085

Ile Lys Ser Glu Glu Gly Glu Lys Met Val Leu Glu Asn Asn Phe Phe
1090 1095 1100

Val Glu Thr Met Leu Pro Ser Lys Ile Met Arg Lys Leu Glu Pro Glu
1105 1110 1115 1120

Glu Phe Ala Ala Tyr Leu Glu Pro Phe Lys Glu Lys Gly Glu Val Arg
1125 1130 1135

Arg Pro Thr Leu Ser Trp Pro Arg Glu Ile Pro Leu Val Lys Gly Gly
1140 1145 1150

Lys Pro Asp Val Val Gln Ile Val Arg Asn Tyr Asn Ala Tyr Leu Arg
1155 1160 1165

Ala Ser Asp Asp Leu Pro Lys Met Phe Ile Glu Ser Asp Pro Gly Phe
1170 1175 1180

Phe Ser Asn Ala Ile Val Glu Gly Ala Lys Lys Phe Pro Asn Thr Glu
1185 1190 1195 1200

Phe Val Lys Val Lys Gly Leu His Phe Ser Gln Glu Asp Ala Pro Asp
1205 1210 1215

Glu Met Gly Lys Tyr Ile Lys Ser Phe Val Glu Arg Val Leu Lys Asn
1220 1225 1230

Glu Gln

<210> 13

<211> 3486

<212> DNA

<213> Artificial sequence

atc tgt tat gtg gag tca tta ttt aag aat cta ttc agg aat tat aac	528
Ile Cys Tyr Val Glu Ser Leu Phe Lys Asn Leu Phe Arg Asn Tyr Asn	
165 170 175	
tat aag gtc cat ctt tta tat gtt ctg cct gaa gtg tta gaa gat tca	576
Tyr Lys Val His Leu Leu Tyr Val Leu Pro Glu Val Leu Glu Asp Ser	
180 185 190	
cct ctg gtt ccc caa aaa ggc agt ttt cag atg gtt cac tgc aat tgc	624
Pro Leu Val Pro Gln Lys Gly Ser Phe Gln Met Val His Cys Asn Cys	
195 200 205	
agt gtt cat gaa tgt tgt gaa tgt ctt gtg cct gtg cca aca gcc aaa	672
Ser Val His Glu Cys Cys Glu Cys Leu Val Pro Val Pro Thr Ala Lys	
210 215 220	
ctc aac gac act ctc ctt atg tgt ttg aaa atc aca tct ggt gga gta	720
Leu Asn Asp Thr Leu Leu Met Cys Leu Lys Ile Thr Ser Gly Gly Val	
225 230 235 240	
att ttc cgg tca cct cta atg tca gtt cag ccc ata aat atg gtg aag	768
Ile Phe Arg Ser Pro Leu Met Ser Val Gln Pro Ile Asn Met Val Lys	
245 250 255	
cct gat cca cca tta ggt ttg cat atg gaa atc aca gat gat ggt aat	816
Pro Asp Pro Pro Leu Gly Leu His Met Glu Ile Thr Asp Asp Gly Asn	
260 265 270	
tta aag att tct tgg tcc agc cca cca ttg gta cca ttt cca ctt caa	864
Leu Lys Ile Ser Trp Ser Ser Pro Pro Leu Val Pro Phe Pro Leu Gln	
275 280 285	
tat caa gtg aaa tat tca gag aat tct aca aca gtt atc aga gaa gct	912
Tyr Gln Val Lys Tyr Ser Glu Asn Ser Thr Thr Val Ile Arg Glu Ala	
290 295 300	
gac aag att gtc tca gct aca tcc ctg cta gta gac agt ata ctt cct	960
Asp Lys Ile Val Ser Ala Thr Ser Leu Leu Val Asp Ser Ile Leu Pro	
305 310 315 320	
ggg tct tgg tat gag gtt cag gtg agg ggc aag aga ctg gat ggc cca	1008
Gly Ser Ser Tyr Glu Val Gln Val Arg Gly Lys Arg Leu Asp Gly Pro	
325 330 335	
gga atc tgg agt gac tgg agt act cct cgt gtc ttt acc aca caa gat	1056
Gly Ile Trp Ser Asp Trp Ser Thr Pro Arg Val Phe Thr Thr Gln Asp	
340 345 350	

gtc ata tac ttt cca cct aaa att ctg aca agt gtt ggg tct aat gtt	1104
Val Ile Tyr Phe Pro Pro Lys Ile Leu Thr Ser Val Gly Ser Asn Val	
355 360 365	
tct ttt cac tgc atc tat aag aag gaa aac aag att gtt ccc tca aaa	1152
Ser Phe His Cys Ile Tyr Lys Lys Glu Asn Lys Ile Val Pro Ser Lys	
370 375 380	
gag att gtt tgg tgg atg aat tta gct gag aaa att cct caa agc cag	1200
Glu Ile Val Trp Trp Met Asn Leu Ala Glu Lys Ile Pro Gln Ser Gln	
385 390 395 400	
tat gat gtt gtg agt gat cat gtt agc aaa gtt act ttt ttc aat ctg	1248
Tyr Asp Val Val Ser Asp His Val Ser Lys Val Thr Phe Phe Asn Leu	
405 410 415	
aat gaa acc aaa cct cga gga aag ttt acc tat gat gca gtg tac tgc	1296
Asn Glu Thr Lys Pro Arg Gly Lys Phe Thr Tyr Asp Ala Val Tyr Cys	
420 425 430	
tgc aat gaa cat gaa tgc cat cat cgc tat gct gaa tta tat gtg att	1344
Cys Asn Glu His Glu Cys His His Arg Tyr Ala Glu Leu Tyr Val Ile	
435 440 445	
gat gtc aat atc aat atc tca tgt gaa act gat ggg tac tta act aaa	1392
Asp Val Asn Ile Asn Ile Ser Cys Glu Thr Asp Gly Tyr Leu Thr Lys	
450 455 460	
atg act tgc aga tgg tca acc agt aca atc cag tca ctt gcg gaa agc	1440
Met Thr Cys Arg Trp Ser Thr Ser Thr Ile Gln Ser Leu Ala Glu Ser	
465 470 475 480	
act ttg caa ttg agg tat cat agg agc agc ctt tac tgt tct gat att	1488
Thr Leu Gln Leu Arg Tyr His Arg Ser Ser Leu Tyr Cys Ser Asp Ile	
485 490 495	
cca tct att cat ccc ata tct gag ccc aaa gat tgc tat ttg cag agt	1536
Pro Ser Ile His Pro Ile Ser Glu Pro Lys Asp Cys Tyr Leu Gln Ser	
500 505 510	
gat ggt ttt tat gaa tgc att ttc cag cca atc ttc cta tta tct ggc	1584
Asp Gly Phe Tyr Glu Cys Ile Phe Gln Pro Ile Phe Leu Leu Ser Gly	
515 520 525	
tac aca atg tgg att agg atc aat cac tct cta ggt tca ctt gac tct	1632
Tyr Thr Met Trp Ile Arg Ile Asn His Ser Leu Gly Ser Leu Asp Ser	
530 535 540	

cca cca aca tgt gtc ctt cct gat tct gtg gtg aag cca ctg cct cca	1680
Pro Pro Thr Cys Val Leu Pro Asp Ser Val Val Lys Pro Leu Pro Pro	
545 550 555 560	
tcc agt gtg aaa gca gaa att act ata aac att gga tta ttg aaa ata	1728
Ser Ser Val Lys Ala Glu Ile Thr Ile Asn Ile Gly Leu Leu Lys Ile	
565 570 575	
tct tgg gaa aag cca gtc ttt cca gag aat aac ctt caa ttc cag att	1776
Ser Trp Glu Lys Pro Val Phe Pro Glu Asn Asn Leu Gln Phe Gln Ile	
580 585 590	
cgc tat ggt tta agt gga aaa gaa gta caa tgg aag atg tat gag gtt	1824
Arg Tyr Gly Leu Ser Gly Lys Glu Val Gln Trp Lys Met Tyr Glu Val	
595 600 605	
tat gat gca aaa tca aaa tct gtc agt ctc cca gtt cca gac ttg tgt	1872
Tyr Asp Ala Lys Ser Lys Ser Val Ser Leu Pro Val Pro Asp Leu Cys	
610 615 620	
gca gtc tat gct gtt cag gtg cgc tgt aag agg cta gat gga ctg gga	1920
Ala Val Tyr Ala Val Gln Val Arg Cys Lys Arg Leu Asp Gly Leu Gly	
625 630 635 640	
tat tgg agt aat tgg agc aat cca gcc tac aca gtt gtc atg gat ata	1968
Tyr Trp Ser Asn Trp Ser Asn Pro Ala Tyr Thr Val Val Met Asp Ile	
645 650 655	
aaa gtt cct atg aga gga cct gaa ttt tgg aga ata att aat gga gat	2016
Lys Val Pro Met Arg Gly Pro Glu Phe Trp Arg Ile Ile Asn Gly Asp	
660 665 670	
act atg aaa aag gag aaa aat gtc act tta ctt tgg aag ccc ctg atg	2064
Thr Met Lys Lys Glu Lys Asn Val Thr Leu Leu Trp Lys Pro Leu Met	
675 680 685	
aaa aat gac tca ttg tgc agt gtt cag aga tat gtg ata aac cat cat	2112
Lys Asn Asp Ser Leu Cys Ser Val Gln Arg Tyr Val Ile Asn His His	
690 695 700	
act tcc tgc aat gga aca tgg tca gaa gat gtg gga aat cac acg aaa	2160
Thr Ser Cys Asn Gly Thr Trp Ser Glu Asp Val Gly Asn His Thr Lys	
705 710 715 720	
ttc act ttc ctg tgg aca gag caa gca cat act gtt acg gtt ctg gcc	2208
Phe Thr Phe Leu Trp Thr Glu Gln Ala His Thr Val Thr Val Leu Ala	
725 730 735	

atc aat tca att ggt gct tct gtt gca aat ttt aat tta acc ttt tca	2256
Ile Asn Ser Ile Gly Ala Ser Val Ala Asn Phe Asn Leu Thr Phe Ser	
740 745 750	
tgg cct atg agc aaa gta aat atc gtg cag tca ctc agt gct tat cct	2304
Trp Pro Met Ser Lys Val Asn Ile Val Gln Ser Leu Ser Ala Tyr Pro	
755 760 765	
tta aac agc agt tgt gtg att gtt tcc tgg ata cta tca ccc agt gat	2352
Leu Asn Ser Ser Cys Val Ile Val Ser Trp Ile Leu Ser Pro Ser Asp	
770 775 780	
tac aag cta atg tat ttt att att gag tgg aaa aat ctt aat gaa gat	2400
Tyr Lys Leu Met Tyr Phe Ile Ile Glu Trp Lys Asn Leu Asn Glu Asp	
785 790 795 800	
ggg gaa ata aaa tgg ctt aga atc tct tca tct gtt aag aag tat tat	2448
Gly Glu Ile Lys Trp Leu Arg Ile Ser Ser Ser Val Lys Lys Tyr Tyr	
805 810 815	
atc cat gat cat ttt atc ccc att gag aag tac cag ttc agt ctt tac	2496
Ile His Asp His Phe Ile Pro Ile Glu Lys Tyr Gln Phe Ser Leu Tyr	
820 825 830	
cca ata ttt atg gaa gga gtg gga aaa cca aag ata att aat agt ttc	2544
Pro Ile Phe Met Glu Gly Val Gly Lys Pro Lys Ile Ile Asn Ser Phe	
835 840 845	
act caa gat gat att gaa aaa cac cag agt gat gca ggt tta tat gta	2592
Thr Gln Asp Asp Ile Glu Lys His Gln Ser Asp Ala Gly Leu Tyr Val	
850 855 860	
att gtg cca gta att att tcc tct tcc atc tta ttg ctt gga aca tta	2640
Ile Val Pro Val Ile Ile Ser Ser Ser Ile Leu Leu Leu Gly Thr Leu	
865 870 875 880	
tta ata tca cac caa aga atg aaa aag cta ttt tgg gaa gat gtt ccg	2688
Leu Ile Ser His Gln Arg Met Lys Lys Leu Phe Trp Glu Asp Val Pro	
885 890 895	
aac ccc aag aat tgt tcc tgg gca caa gga ctt aat ttt cag aag aga	2736
Asn Pro Lys Asn Cys Ser Trp Ala Gln Gly Leu Asn Phe Gln Lys Arg	
900 905 910	
acg gac att ctg gat cca ccg gtc gcc acc atg gtg agc aag ggc gag	2784
Thr Asp Ile Leu Asp Pro Pro Val Ala Thr Met Val Ser Lys Gly Glu	
915 920 925	

gag ctg ttc acc ggg gtg gtg ccc atc ctg gtc gag ctg gac ggc gac	2832
Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp	
930 935 940	
gta aac ggc cac aag ttc agc gtg tcc ggc gag ggc gag ggc gat gcc	2880
Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala	
945 950 955 960	
acc tac ggc aag ctg acc ctg aag ttc atc tgc acc acc ggc aag ctg	2928
Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu	
965 970 975	
ccc gtg ccc tgg ccc acc ctc gtg acc acc ttc ggc tac ggc gtg cag	2976
Pro Val Pro Trp Pro Thr Leu Val Thr Thr Phe Gly Tyr Gly Val Gln	
980 985 990	
tgc ttc gcc cgc tac ccc gac cac atg cgc cag cac gac ttc ttc aag	3024
Cys Phe Ala Arg Tyr Pro Asp His Met Arg Gln His Asp Phe Phe Lys	
995 1000 1005	
tcc gcc atg ccc gaa ggc tac gtc cag gag cgc acc atc ttc ttc aag	3072
Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys	
1010 1015 1020	
gac gac ggc aac tac aag acc cgc gcc gag gtg aag ttc gag ggc gac	3120
Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp	
1025 1030 1035 1040	
acc ctg gtg aac cgc atc gag ctg aag ggc atc gac ttc aag gag gac	3168
Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp	
1045 1050 1055	
ggc aac atc ctg ggg cac aag ctg gag tac aac tac aac agc cac aac	3216
Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn	
1060 1065 1070	
gtc tat atc atg gcc gac aag cag aag aac ggc atc aag gtg aac ttc	3264
Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Phe	
1075 1080 1085	
aag atc cgc cac aac atc gag gac ggc agc gtg cag ctc gcc gac cac	3312
Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His	
1090 1095 1100	
tac cag cag aac acc ccc atc ggc gac ggc ccc gtg ctg ctg ccc gac	3360
Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp	
1105 1110 1115 1120	

aac cac tac ctg agc tac cag tcc gcc ctg agc aaa gac ccc aac gag 3408
 Asn His Tyr Leu Ser Tyr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu
 1125 1130 1135

aag cgc gat cac atg gtc ctg ctg gag ttc gtg acc gcc gcc ggg atc 3456
 Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile
 1140 1145 1150

act ctc ggc atg gac gag ctg tac aag taa 3486
 Thr Leu Gly Met Asp Glu Leu Tyr Lys
 1155 1160

<210> 14

<211> 1161

<212> PRT

<213> Artificial sequence

<223> Artificial sequence description : OBR YFP

<400> 14

Met Val Leu Ala Ser Ser Thr Thr Ser Ile His Thr Met Leu Leu Leu
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Leu Leu Met Leu Phe His Leu Gly Leu Gln Ala Ser Ile Ser Ala Arg
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Gln Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu Thr Arg Tyr Pro Ile
 35 40 45

Thr Pro Trp Arg Phe Lys Leu Ser Cys Met Pro Pro Asn Ser Thr Tyr
 50 55 60

Asp Tyr Phe Leu Leu Pro Ala Gly Leu Ser Lys Asn Thr Ser Asn Ser
 65 70 75 80

Asn Gly His Tyr Glu Thr Ala Val Glu Pro Lys Phe Asn Ser Ser Gly
 85 90 95

Thr His Phe Ser Asn Leu Ser Lys Thr Thr Phe His Cys Cys Phe Arg
 100 105 110

Ser Glu Gln Asp Arg Asn Cys Ser Leu Cys Ala Asp Asn Ile Glu Gly
 115 120 125

Thr Thr Phe Val Ser Thr Val Asn Ser Leu Val Phe Gln Gln Ile Asp
 130 135 140

Ala Asn Trp Asn Ile Gln Cys Trp Leu Lys Gly Asp Leu Lys Leu Phe			
145	150	155	160
Ile Cys Tyr Val Glu Ser Leu Phe Lys Asn Leu Phe Arg Asn Tyr Asn			
	165	170	175
Tyr Lys Val His Leu Leu Tyr Val Leu Pro Glu Val Leu Glu Asp Ser			
	180	185	190
Pro Leu Val Pro Gln Lys Gly Ser Phe Gln Met Val His Cys Asn Cys			
	195	200	205
Ser Val His Glu Cys Cys Glu Cys Leu Val Pro Val Pro Thr Ala Lys			
	210	215	220
Leu Asn Asp Thr Leu Leu Met Cys Leu Lys Ile Thr Ser Gly Gly Val			
225	230	235	240
Ile Phe Arg Ser Pro Leu Met Ser Val Gln Pro Ile Asn Met Val Lys			
	245	250	255
Pro Asp Pro Pro Leu Gly Leu His Met Glu Ile Thr Asp Asp Gly Asn			
	260	265	270
Leu Lys Ile Ser Trp Ser Ser Pro Pro Leu Val Pro Phe Pro Leu Gln			
	275	280	285
Tyr Gln Val Lys Tyr Ser Glu Asn Ser Thr Thr Val Ile Arg Glu Ala			
	290	295	300
Asp Lys Ile Val Ser Ala Thr Ser Leu Leu Val Asp Ser Ile Leu Pro			
305	310	315	320
Gly Ser Ser Tyr Glu Val Gln Val Arg Gly Lys Arg Leu Asp Gly Pro			
	325	330	335
Gly Ile Trp Ser Asp Trp Ser Thr Pro Arg Val Phe Thr Thr Gln Asp			
	340	345	350
Val Ile Tyr Phe Pro Pro Lys Ile Leu Thr Ser Val Gly Ser Asn Val			
	355	360	365
Ser Phe His Cys Ile Tyr Lys Lys Glu Asn Lys Ile Val Pro Ser Lys			
	370	375	380
Glu Ile Val Trp Trp Met Asn Leu Ala Glu Lys Ile Pro Gln Ser Gln			
385	390	395	400

Tyr Asp Val Val Ser Asp His Val Ser Lys Val Thr Phe Phe Asn Leu
 405 410 415
 Asn Glu Thr Lys Pro Arg Gly Lys Phe Thr Tyr Asp Ala Val Tyr Cys
 420 425 430
 Cys Asn Glu His Glu Cys His His Arg Tyr Ala Glu Leu Tyr Val Ile
 435 440 445
 Asp Val Asn Ile Asn Ile Ser Cys Glu Thr Asp Gly Tyr Leu Thr Lys
 450 455 460
 Met Thr Cys Arg Trp Ser Thr Ser Thr Ile Gln Ser Leu Ala Glu Ser
 465 470 475 480
 Thr Leu Gln Leu Arg Tyr His Arg Ser Ser Leu Tyr Cys Ser Asp Ile
 485 490 495
 Pro Ser Ile His Pro Ile Ser Glu Pro Lys Asp Cys Tyr Leu Gln Ser
 500 505 510
 Asp Gly Phe Tyr Glu Cys Ile Phe Gln Pro Ile Phe Leu Leu Ser Gly
 515 520 525
 Tyr Thr Met Trp Ile Arg Ile Asn His Ser Leu Gly Ser Leu Asp Ser
 530 535 540
 Pro Pro Thr Cys Val Leu Pro Asp Ser Val Val Lys Pro Leu Pro Pro
 545 550 555 560
 Ser Ser Val Lys Ala Glu Ile Thr Ile Asn Ile Gly Leu Leu Lys Ile
 565 570 575
 Ser Trp Glu Lys Pro Val Phe Pro Glu Asn Asn Leu Gln Phe Gln Ile
 580 585 590
 Arg Tyr Gly Leu Ser Gly Lys Glu Val Gln Trp Lys Met Tyr Glu Val
 595 600 605
 Tyr Asp Ala Lys Ser Lys Ser Val Ser Leu Pro Val Pro Asp Leu Cys
 610 615 620
 Ala Val Tyr Ala Val Gln Val Arg Cys Lys Arg Leu Asp Gly Leu Gly
 625 630 635 640
 Tyr Trp Ser Asn Trp Ser Asn Pro Ala Tyr Thr Val Val Met Asp Ile
 645 650 655

Lys	Val	Pro	Met	Arg	Gly	Pro	Glu	Phe	Trp	Arg	Ile	Ile	Asn	Gly	Asp	
			660					665					670			
Thr	Met	Lys	Lys	Glu	Lys	Asn	Val	Thr	Leu	Leu	Trp	Lys	Pro	Leu	Met	
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Thr	Ser	Cys	Asn	Gly	Thr	Trp	Ser	Glu	Asp	Val	Gly	Asn	His	Thr	Lys	
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Phe	Thr	Phe	Leu	Trp	Thr	Glu	Gln	Ala	His	Thr	Val	Thr	Val	Leu	Ala	
				725					730					735		
Ile	Asn	Ser	Ile	Gly	Ala	Ser	Val	Ala	Asn	Phe	Asn	Leu	Thr	Phe	Ser	
			740					745					750			
Trp	Pro	Met	Ser	Lys	Val	Asn	Ile	Val	Gln	Ser	Leu	Ser	Ala	Tyr	Pro	
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Leu	Asn	Ser	Ser	Cys	Val	Ile	Val	Ser	Trp	Ile	Leu	Ser	Pro	Ser	Asp	
	770					775					780					
Tyr	Lys	Leu	Met	Tyr	Phe	Ile	Ile	Glu	Trp	Lys	Asn	Leu	Asn	Glu	Asp	
785					790					795					800	
Gly	Glu	Ile	Lys	Trp	Leu	Arg	Ile	Ser	Ser	Ser	Val	Lys	Lys	Tyr	Tyr	
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Ile	His	Asp	His	Phe	Ile	Pro	Ile	Glu	Lys	Tyr	Gln	Phe	Ser	Leu	Tyr	
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Pro	Ile	Phe	Met	Glu	Gly	Val	Gly	Lys	Pro	Lys	Ile	Ile	Asn	Ser	Phe	
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Ile	Val	Pro	Val	Ile	Ile	Ser	Ser	Ser	Ile	Leu	Leu	Leu	Gly	Thr	Leu	
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Leu	Ile	Ser	His	Gln	Arg	Met	Lys	Lys	Leu	Phe	Trp	Glu	Asp	Val	Pro	
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Asn	Pro	Lys	Asn	Cys	Ser	Trp	Ala	Gln	Gly	Leu	Asn	Phe	Gln	Lys	Arg	
			900					905					910			

Thr Asp Ile Leu Asp Pro Pro Val Ala Thr Met Val Ser Lys Gly Glu
 915 920 925

Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp
 930 935 940

Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala
 945 950 955 960

Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu
 965 970 975

Pro Val Pro Trp Pro Thr Leu Val Thr Thr Phe Gly Tyr Gly Val Gln
 980 985 990

Cys Phe Ala Arg Tyr Pro Asp His Met Arg Gln His Asp Phe Phe Lys
 995 1000 1005

Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys
 1010 1015 1020

Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp
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Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp
 1045 1050 1055

Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn
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Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Phe
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Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His
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Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp
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Asn His Tyr Leu Ser Tyr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu
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Tyr Trp Pro Leu Phe Val Leu Phe Phe Tyr Ile Leu Ser Pro Ile Pro
35 40 45
Tyr Cys Ile Ala Arg Arg Leu Val Asp Asp Thr Asp Ala Met Ser Asn
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Ala Cys Lys Glu Leu Ala Ile Phe Leu Thr Thr Gly Ile Val Val Ser
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Ala Phe Gly Leu Pro Ile Val Phe Ala Arg Ala His Leu Ile Glu Trp
85 90 95
Gly Ala Cys Ala Leu Val Leu Thr Gly Asn Thr Val Ile Phe Ala Thr
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Ile Leu Gly Phe Phe Leu Val Phe Gly Ser Asn Asp Asp Phe Ser Trp
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Gln Gln Trp
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gga ctg atg ttt ttg atg ctt gga tgt gcc ctt cca ata tac aac aaa	96
Gly Leu Met Phe Leu Met Leu Gly Cys Ala Leu Pro Ile Tyr Asn Lys	
20 25 30	
tac tgg ccc ctc ttt gtt cta ttt ttt tac atc ctt tca cct att cca	144
Tyr Trp Pro Leu Phe Val Leu Phe Phe Tyr Ile Leu Ser Pro Ile Pro	
35 40 45	
tac tgc ata gca aga aga tta gtg gat gat aca gat gct atg agt aac	192
Tyr Cys Ile Ala Arg Arg Leu Val Asp Asp Thr Asp Ala Met Ser Asn	
50 55 60	
gct tgt aag gaa ctt gcc atc ttt ctt aca acg ggc att gtc gtg tca	240
Ala Cys Lys Glu Leu Ala Ile Phe Leu Thr Thr Gly Ile Val Val Ser	
65 70 75 80	
gct ttt gga ctc cct att gta ttt gcc aga gca cat ctg att gag tgg	288
Ala Phe Gly Leu Pro Ile Val Phe Ala Arg Ala His Leu Ile Glu Trp	
85 90 95	
gga gct tgt gca ctt gtt ctc aca gga aac aca gtc atc ttt gca act	336
Gly Ala Cys Ala Leu Val Leu Thr Gly Asn Thr Val Ile Phe Ala Thr	
100 105 110	
ata cta ggc ttt ttc ttg gtc ttt gga agc aat gac gac ttc agc tgg	384
Ile Leu Gly Phe Phe Leu Val Phe Gly Ser Asn Asp Asp Phe Ser Trp	
115 120 125	
cag cag tgg cga ccg gtg gat cca ccg gct aga gcc acc atg acc agc	432
Gln Gln Trp Arg Pro Val Asp Pro Pro Ala Arg Ala Thr Met Thr Ser	
130 135 140	
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Lys Val Tyr Asp Pro Glu Gln Arg Lys Arg Met Ile Thr Gly Pro Gln	
145 150 155 160	
tgg tgg gcc agg tgc aag cag atg aac gtg ctg gac agc ttc atc aac	528
Trp Trp Ala Arg Cys Lys Gln Met Asn Val Leu Asp Ser Phe Ile Asn	
165 170 175	
tac tac gac agc gag aag cac gcc gag aac gcc gtg atc ttc ctg cac	576
Tyr Tyr Asp Ser Glu Lys His Ala Glu Asn Ala Val Ile Phe Leu His	
180 185 190	

ggc aac gcc gct agc agc tac ctg tgg agg cac gtg gtg ccc cac atc	624
Gly Asn Ala Ala Ser Ser Tyr Leu Trp Arg His Val Val Pro His Ile	
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Glu Pro Val Ala Arg Cys Ile Ile Pro Asp Leu Ile Gly Met Gly Lys	
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Ser Gly Lys Ser Gly Asn Gly Ser Tyr Arg Leu Leu Asp His Tyr Lys	
225 230 235 240	
tac ctg acc gcc tgg ttc gag ctc ctg aac ctg ccc aag aag atc atc	768
Tyr Leu Thr Ala Trp Phe Glu Leu Leu Asn Leu Pro Lys Lys Ile Ile	
245 250 255	
ttc gtg ggc cac gac tgg ggc gcc tgc ctg gcc ttc cac tac agc tac	816
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260 265 270	
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Glu His Gln Asp Lys Ile Lys Ala Ile Val His Ala Glu Ser Val Val	
275 280 285	
gac gtg atc gag agc tgg gac gag tgg cca gac atc gag gag gac atc	912
Asp Val Ile Glu Ser Trp Asp Glu Trp Pro Asp Ile Glu Glu Asp Ile	
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Ala Leu Ile Lys Ser Glu Glu Gly Glu Lys Met Val Leu Glu Asn Asn	
305 310 315 320	
ttc ttc gtg gag acc atg ctg ccc agc aag atc atg aga aag ctg gag	1008
Phe Phe Val Glu Thr Met Leu Pro Ser Lys Ile Met Arg Lys Leu Glu	
325 330 335	
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Pro Glu Glu Phe Ala Ala Tyr Leu Glu Pro Phe Lys Glu Lys Gly Glu	
340 345 350	
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Val Arg Arg Pro Thr Leu Ser Trp Pro Arg Glu Ile Pro Leu Val Lys	
355 360 365	
ggc ggc aag ccc gac gtg gtg cag atc gtg aga aac tac aac gcc tac	1152
Gly Gly Lys Pro Asp Val Val Gln Ile Val Arg Asn Tyr Asn Ala Tyr	
370 375 380	

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Gly	Phe	Phe	Ser	Asn	Ala	Ile	Val	Glu	Gly	Ala	Lys	Lys	Phe	Pro	Asn	
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acc	gag	ttc	gtg	aag	gtg	aag	ggc	ctg	cac	ttc	agc	cag	gag	gac	gcc	1296
Thr	Glu	Phe	Val	Lys	Val	Lys	Gly	Leu	His	Phe	Ser	Gln	Glu	Asp	Ala	
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ccc	gac	gag	atg	ggc	aag	tac	atc	aag	agc	ttc	gtg	gag	aga	gtg	ctg	1344
Pro	Asp	Glu	Met	Gly	Lys	Tyr	Ile	Lys	Ser	Phe	Val	Glu	Arg	Val	Leu	
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Tyr	Trp	Pro	Leu	Phe	Val	Leu	Phe	Phe	Tyr	Ile	Leu	Ser	Pro	Ile	Pro
		35					40					45			
Tyr	Cys	Ile	Ala	Arg	Arg	Leu	Val	Asp	Asp	Thr	Asp	Ala	Met	Ser	Asn
	50					55				60					
Ala	Cys	Lys	Glu	Leu	Ala	Ile	Phe	Leu	Thr	Thr	Gly	Ile	Val	Val	Ser
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Ala	Phe	Gly	Leu	Pro	Ile	Val	Phe	Ala	Arg	Ala	His	Leu	Ile	Glu	Trp
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Gly	Ala	Cys	Ala	Leu	Val	Leu	Thr	Gly	Asn	Thr	Val	Ile	Phe	Ala	Thr

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Ile Leu Gly Phe Phe Leu Val Phe Gly Ser Asn Asp Asp Phe Ser Trp					
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Gln Gln Trp Arg Pro Val Asp Pro Pro Ala Arg Ala Thr Met Thr Ser					
	130		135		140
Lys Val Tyr Asp Pro Glu Gln Arg Lys Arg Met Ile Thr Gly Pro Gln					
	145		150		155
Trp Trp Ala Arg Cys Lys Gln Met Asn Val Leu Asp Ser Phe Ile Asn					
			165		170
Tyr Tyr Asp Ser Glu Lys His Ala Glu Asn Ala Val Ile Phe Leu His					
			180		185
Gly Asn Ala Ala Ser Ser Tyr Leu Trp Arg His Val Val Pro His Ile					
			195		200
Glu Pro Val Ala Arg Cys Ile Ile Pro Asp Leu Ile Gly Met Gly Lys					
			210		215
Ser Gly Lys Ser Gly Asn Gly Ser Tyr Arg Leu Leu Asp His Tyr Lys					
			225		230
Tyr Leu Thr Ala Trp Phe Glu Leu Leu Asn Leu Pro Lys Lys Ile Ile					
			245		250
Phe Val Gly His Asp Trp Gly Ala Cys Leu Ala Phe His Tyr Ser Tyr					
			260		265
Glu His Gln Asp Lys Ile Lys Ala Ile Val His Ala Glu Ser Val Val					
			275		280
Asp Val Ile Glu Ser Trp Asp Glu Trp Pro Asp Ile Glu Glu Asp Ile					
			290		295
Ala Leu Ile Lys Ser Glu Glu Gly Glu Lys Met Val Leu Glu Asn Asn					
			305		310
Phe Phe Val Glu Thr Met Leu Pro Ser Lys Ile Met Arg Lys Leu Glu					
			325		330
Pro Glu Glu Phe Ala Ala Tyr Leu Glu Pro Phe Lys Glu Lys Gly Glu					
			340		345
Val Arg Arg Pro Thr Leu Ser Trp Pro Arg Glu Ile Pro Leu Val Lys					

Tyr Cys Ile Ala Arg Arg Leu Val Asp Asp Thr Asp Ala Met Ser Asn	
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Ala Cys Lys Glu Leu Ala Ile Phe Leu Thr Thr Gly Ile Val Val Ser	
65 70 75 80	
gct ttt gga ctc cct att gta ttt gcc aga gca cat ctg att gag tgg	288
Ala Phe Gly Leu Pro Ile Val Phe Ala Arg Ala His Leu Ile Glu Trp	
85 90 95	
gga gct tgt gca ctt gtt ctc aca gga aac aca gtc atc ttt gca act	336
Gly Ala Cys Ala Leu Val Leu Thr Gly Asn Thr Val Ile Phe Ala Thr	
100 105 110	
ata cta ggc ttt ttc ttg gtc ttt gga agc aat gac gac ttc agc tgg	384
Ile Leu Gly Phe Phe Leu Val Phe Gly Ser Asn Asp Asp Phe Ser Trp	
115 120 125	
cag cag tgg cga ccg gtg gat cca ccg gtc gcc acc atg gtg agc aag	432
Gln Gln Trp Arg Pro Val Asp Pro Pro Val Ala Thr Met Val Ser Lys	
130 135 140	
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Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp	
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Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly	
165 170 175	
gat gcc acc tac ggc aag ctg acc ctg aag ttc atc tgc acc acc ggc	576
Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly	
180 185 190	
aag ctg ccc gtg ccc tgg ccc acc ctc gtg acc acc ttc ggc tac ggc	624
Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Phe Gly Tyr Gly	
195 200 205	
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225 230 235 240	
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Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu	
245 250 255	
ggc gac acc ctg gtg aac cgc atc gag ctg aag ggc atc gac ttc aag	816
Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys	
260 265 270	
gag gac ggc aac atc ctg ggg cac aag ctg gag tac aac tac aac agc	864
Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser	
275 280 285	
cac aac gtc tat atc atg gcc gac aag cag aag aac ggc atc aag gtg	912
His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Val	
290 295 300	
aac ttc aag atc cgc cac aac atc gag gac ggc agc gtg cag ctc gcc	960
Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala	
305 310 315 320	
gac cac tac cag cag aac acc ccc atc ggc gac ggc ccc gtg ctg ctg	1008
Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu	
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Pro Asp Asn His Tyr Leu Ser Tyr Gln Ser Ala Leu Ser Lys Asp Pro	
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<211> 379

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<213> Artificial sequence

<223> Artificial sequence description : MY47 YFP

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Tyr	Cys	Ile	Ala	Arg	Arg	Leu	Val	Asp	Asp	Thr	Asp	Ala	Met	Ser	Asn	50	55	60	
Ala	Cys	Lys	Glu	Leu	Ala	Ile	Phe	Leu	Thr	Thr	Gly	Ile	Val	Val	Ser	65	70	75	80
Ala	Phe	Gly	Leu	Pro	Ile	Val	Phe	Ala	Arg	Ala	His	Leu	Ile	Glu	Trp	85	90	95	
Gly	Ala	Cys	Ala	Leu	Val	Leu	Thr	Gly	Asn	Thr	Val	Ile	Phe	Ala	Thr	100	105	110	
Ile	Leu	Gly	Phe	Phe	Leu	Val	Phe	Gly	Ser	Asn	Asp	Asp	Phe	Ser	Trp	115	120	125	
Gln	Gln	Trp	Arg	Pro	Val	Asp	Pro	Pro	Val	Ala	Thr	Met	Val	Ser	Lys	130	135	140	
Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile	Leu	Val	Glu	Leu	Asp	145	150	155	160
Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Ser	Gly	Glu	Gly	Glu	Gly	165	170	175	
Asp	Ala	Thr	Tyr	Gly	Lys	Leu	Thr	Leu	Lys	Phe	Ile	Cys	Thr	Thr	Gly	180	185	190	
Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val	Thr	Thr	Phe	Gly	Tyr	Gly	195	200	205	
Val	Gln	Cys	Phe	Ala	Arg	Tyr	Pro	Asp	His	Met	Arg	Gln	His	Asp	Phe	210	215	220	
Phe	Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Val	Gln	Glu	Arg	Thr	Ile	Phe	225	230	235	240
Phe	Lys	Asp	Asp	Gly	Asn	Tyr	Lys	Thr	Arg	Ala	Glu	Val	Lys	Phe	Glu	245	250	255	
Gly	Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys	Gly	Ile	Asp	Phe	Lys	260	265	270	
Glu	Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu	Tyr	Asn	Tyr	Asn	Ser	275	280	285	

Figure 1:

```

AS 01: 3'-teg-G*G*G C*C*C*G G*C A*C*C G*T*C*C T*T*C*G
AS 02: 3'-teg-G*G*G*T*C A A G*C*C*C T*C*T G*T A*C*C*G
AS 03: 3'-teg-G*C*C*C T*C*T G*T A*C*C G*C*C*C G*C*A*A
AS 04: 3'-teg-T*A*C*C G*C*C*C G*C A A*T*T*T*C G A*G*A
AS 05: 3'-teg-T*T*C*G A G A*G*C A*C*C G*T A A*T A*G*G
AS 06: 3'-teg-G*A*A*T A*C G A*C*C*C*T A*C A*C G G*A*A
AS 07: 3'-teg-A*C*A*C G G A A*T*C*T C*C*T*A A*T A*C*C
AS 08: 3'-teg-C*T*C*C*T A A*T A*C*C G*C A A*A*T G*A*C
AS 09: 3'-teg-C*C*G*C*A A A*T G A*C*C*G G G A*A T*A*A
AS 10: 3'-teg-A*C*G G A*C A G*C*C*C T*T*G A*C*C G*T*A
AS 11: 3'-teg-G*G*A*C A G*C*C*C T*T*G A*C*C G*T A*T A*A*A
AS 12: 3'-teg-G*C*C*C T*T*G A*C*C G*T A*T A A*A G*A*A
AS 13: 3'-teg-G*G*A A*C*A*C A A*C*C*G T*C*C*G T*T*A*C
AS 14: 3'-teg-T*G*T A*C A*C G*T G*T A*C G*C*C G*T A*A
AS 15: 3'-teg-G*C*C*T C*C*T G*T C*C*A G*C*C*G C*C*A*A
AS 16: 3'-teg-G*G*A*C*C G A*C*A T*T*G C*A*C G*T C*T A*A*A

```

*: Thioester

_ : 2'-O-Methylation

teg: Triethylene glycol spacer

Figure 2

	10	20	30	40	50	60
OB-RGRP_human	-----	MAG-VKALVALSFSGAIGLTFMLMLGCALEDYGVYWPLFVLI	FHAIS			
My47_human	-----	MAG-IKALISLSFGGAIGLMFLMLGCALPIYNKYWPLFVLI	FFYILS			
yt02_C.elegans	MCCHIH	IQCFDCCSMKNTILAVAALAFAGVVGLTFLVLGCALPRYGTWTPMFVITFYVLS				
YJ14_Yeast	-----	MMEFKVSPLTKIISLSGFLALGFLLVLSCAL--FHNYYP	LF	DILIFLLA		
Consensus	MCCHIH	IQCF2222MAG2IKALI2LSF4GAIGLTFMLMLGCALP3YG4YWPLFV24FY4LS				

	70	80	90	100	110	120
OB-RGRP_human	PIPHFIAKR----	VTYDS	DATSSACRE	LAYFFTTGIVVSAFGFPVILARVAVIKWGACG		
My47_human	PIPYCIARR----	LVDDT	DAMSNACKELAI	FLTTGIVVSAFGLPIVFARAH	LIEWGACA	
yt02_C.elegans	PVPLLIARR----	FQEDMTGTN-	ACIELALFIT	TGIVISAFALPIVLAHAGTIAMSACF		
YJ14_Yeast	PIPNTIFNAGNKYHTSD	FMSDSSNTGQDLA	HFLTGMLVTSGIALPVV	FYHCQLIGHLSCI		
Consensus	PIP44IARRGNKYH44DDMDATS	SNAC4ELA4FLTTGIVVSAF2LP2V2A2A4LI4WGAC4				

	130	140	150
OB-RGRP_human	LVLAGNAVIFLTIQGF	FLIFGRGDDFSWEQW-	
My47_human	LVLTGNTVIFATILGF	FLVFGSKDDFSWQQW-	
yt02_C.elegans	LIFIANSTINFSVIF	YFRIFNGEDMNGMSLW-	
YJ14_Yeast	MCMIGGLIIYSSIVIF	FKWFFKKDFNEDDSLFG	
Consensus	LVLIGN42IFSTI4GF	FLIFG44DDFSWS2WG	

Figure 3A

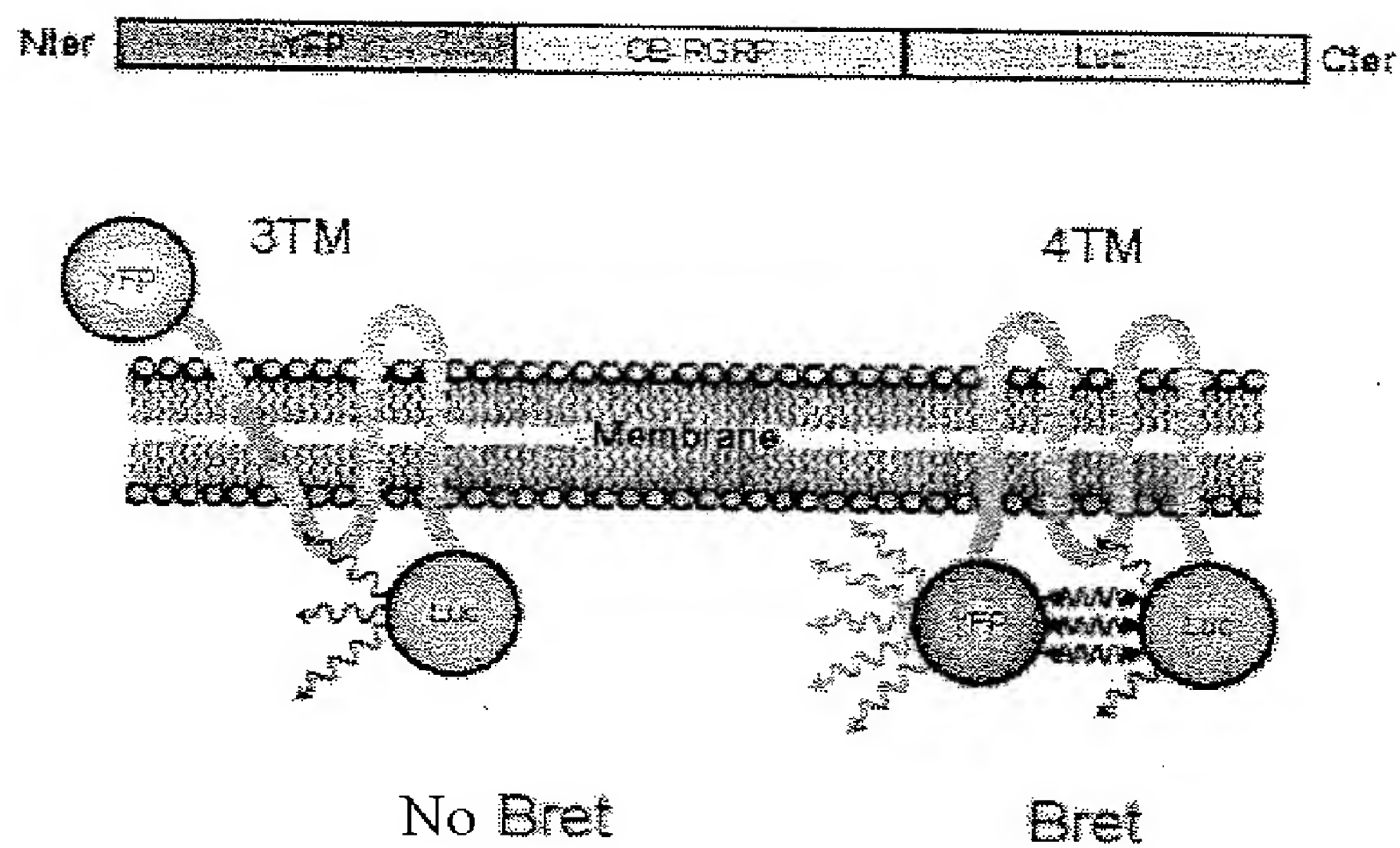


Figure 3B

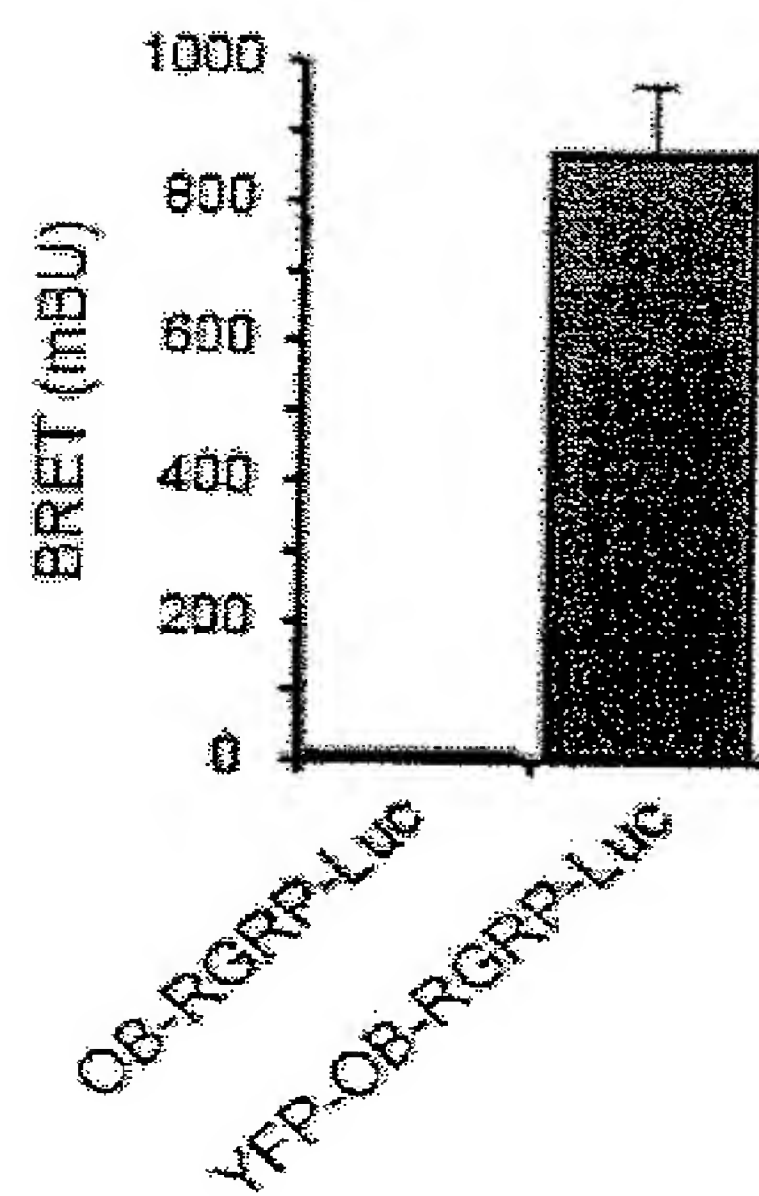


Figure 4 A

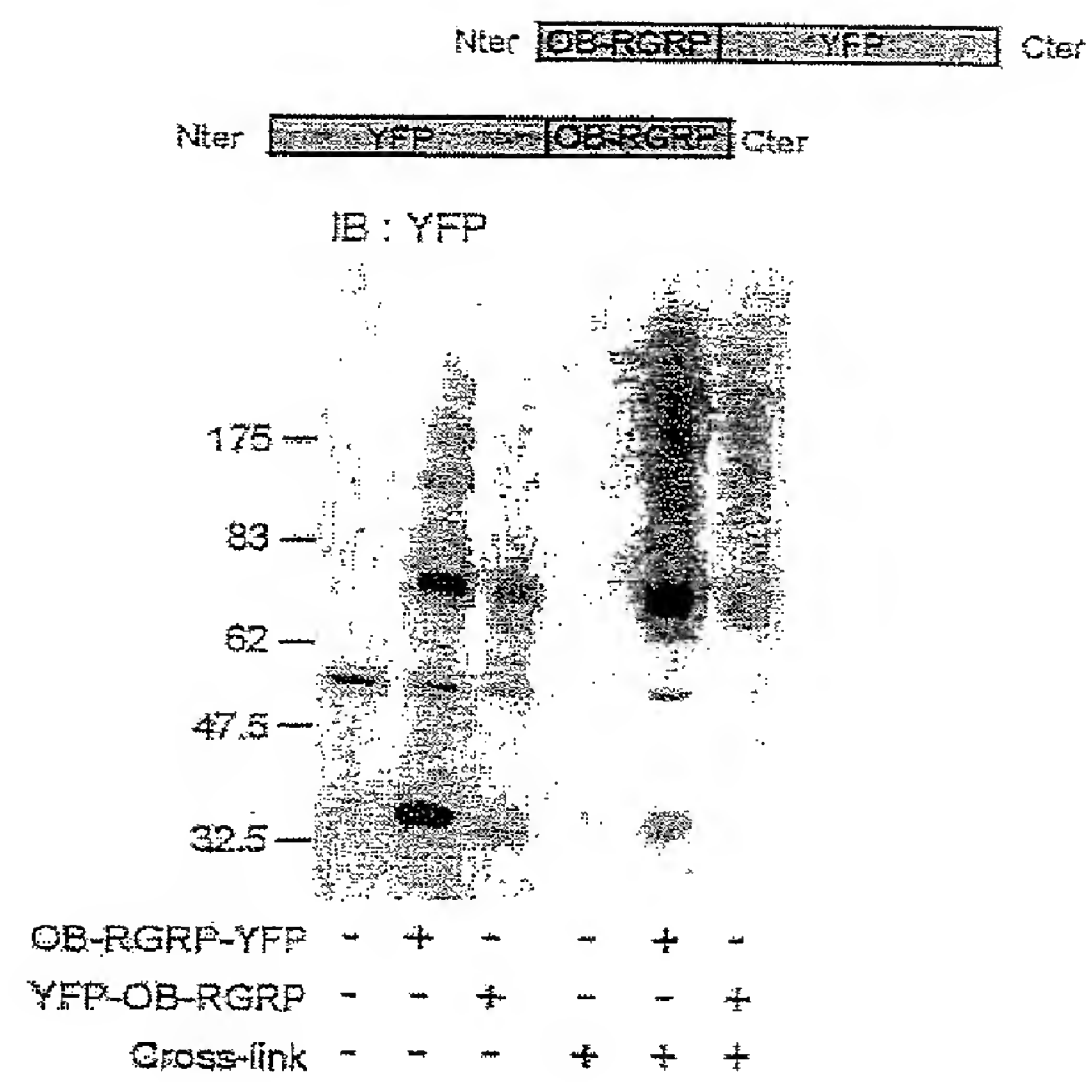


Figure 4B

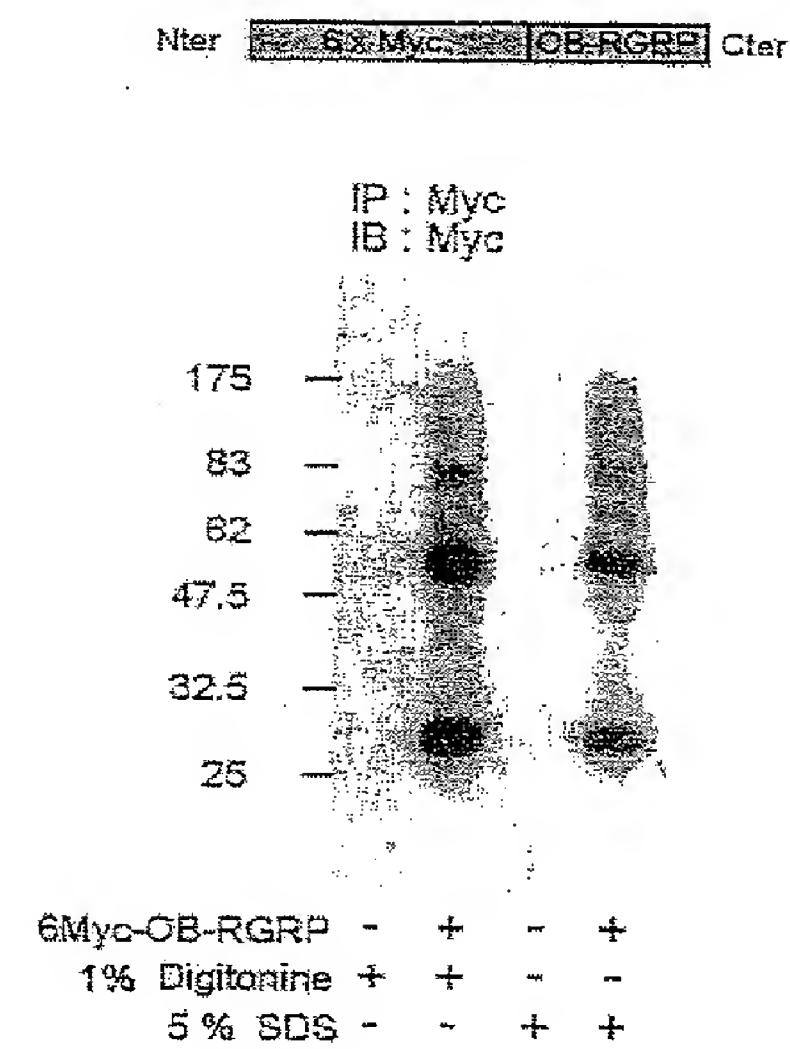


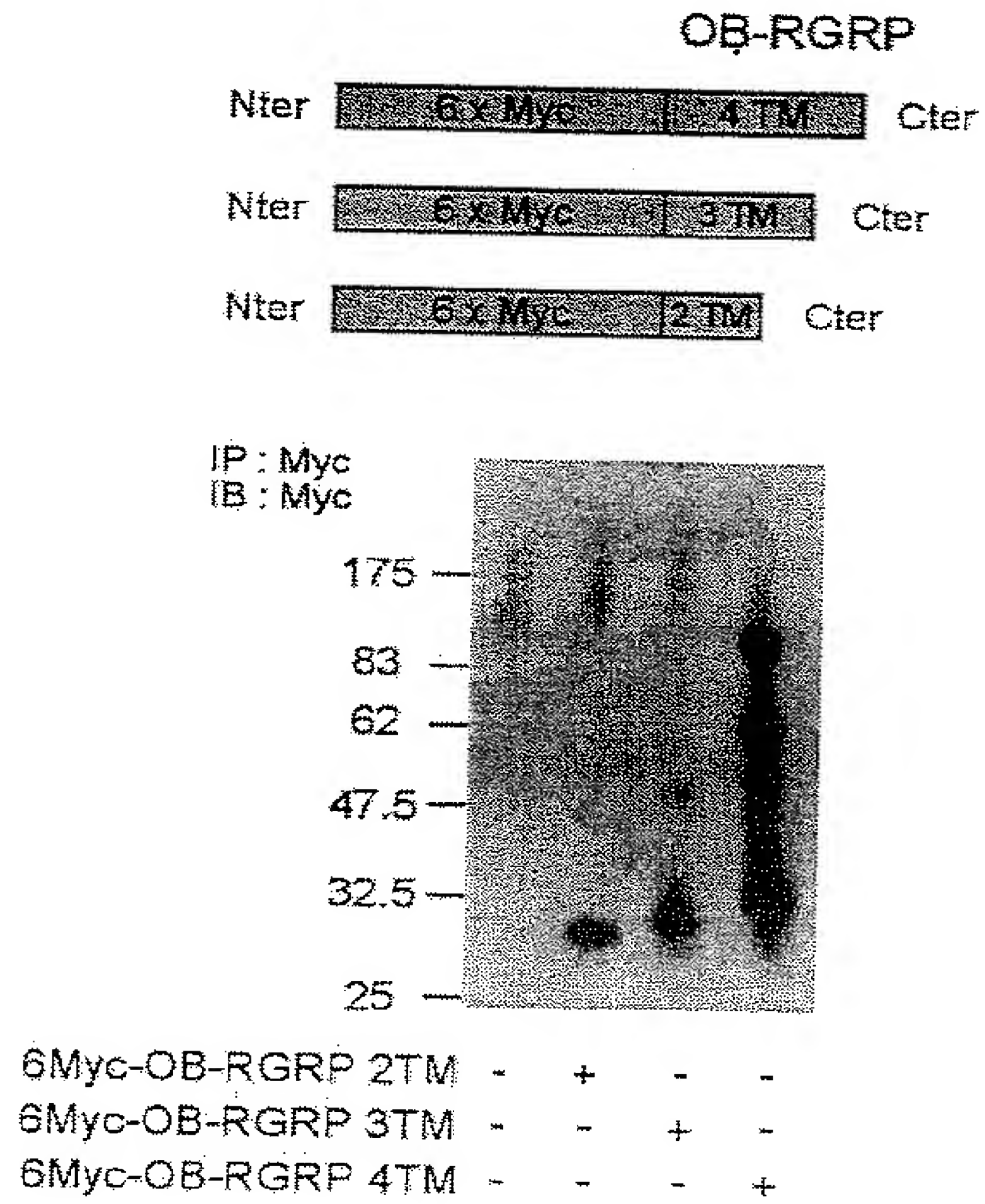
Figure 5

Figure 6 A

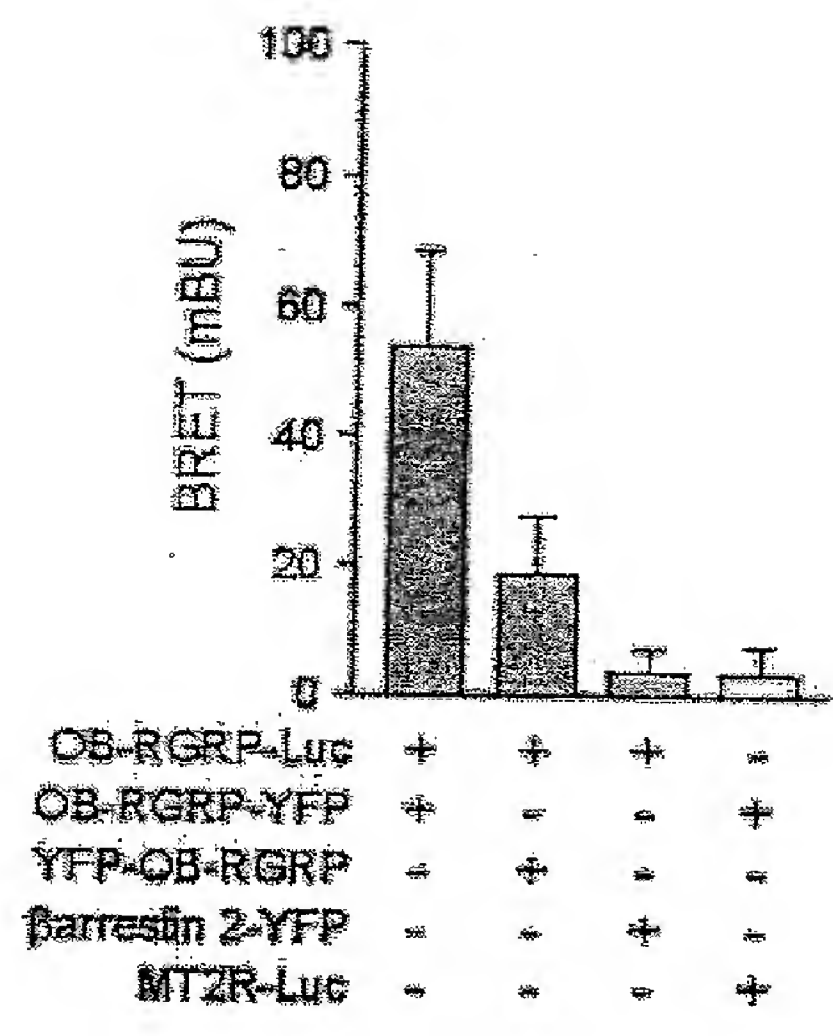


Figure 6B

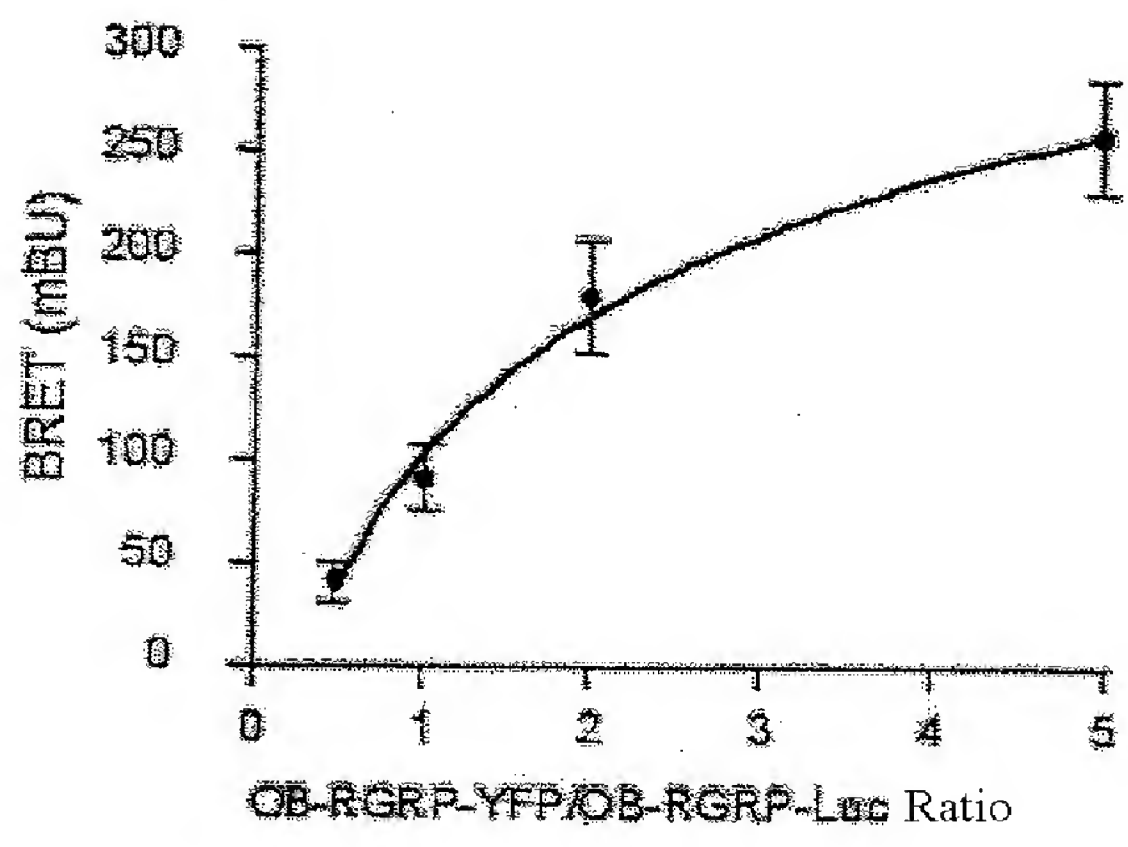


Figure 7

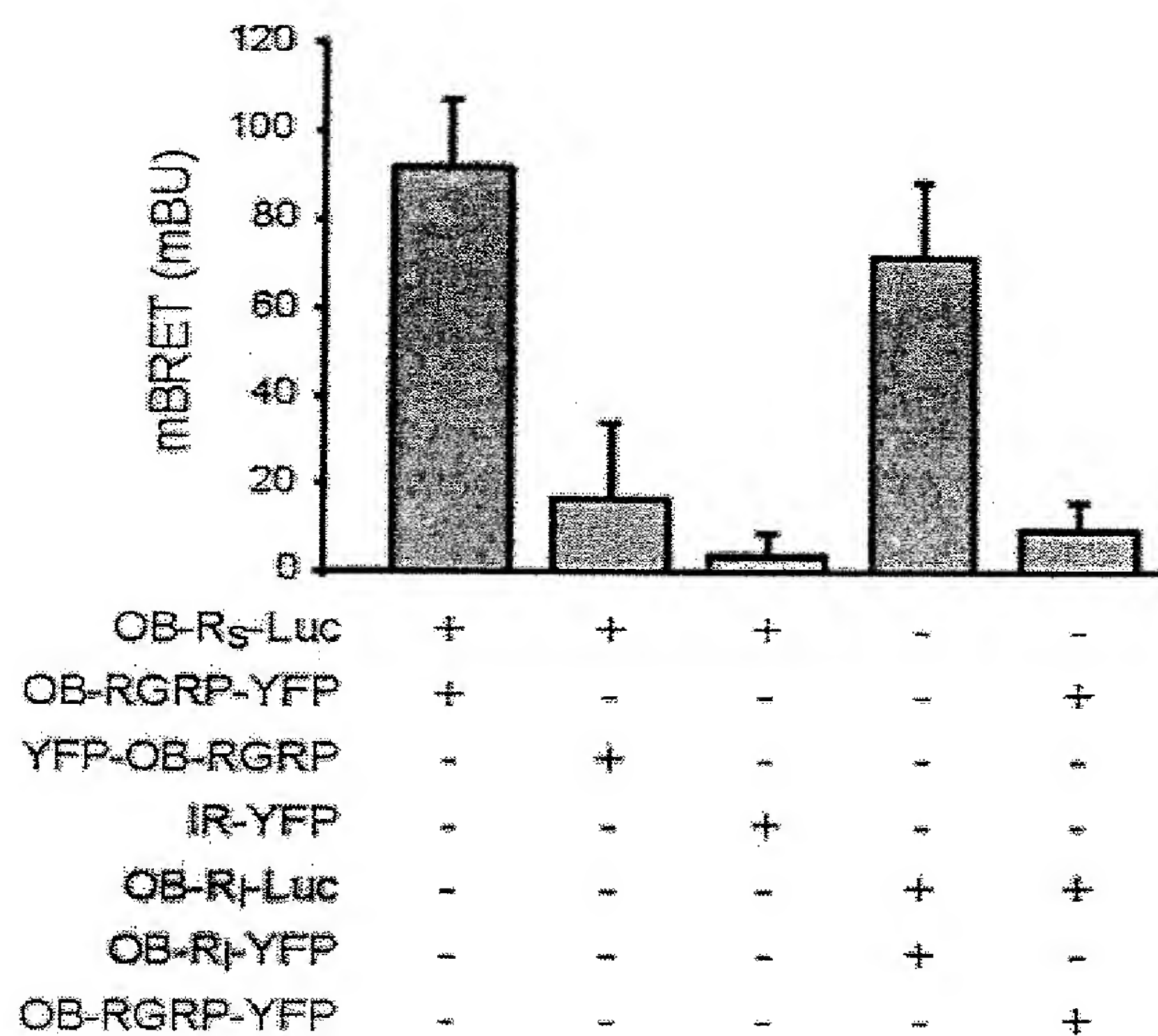


Figure 8 a

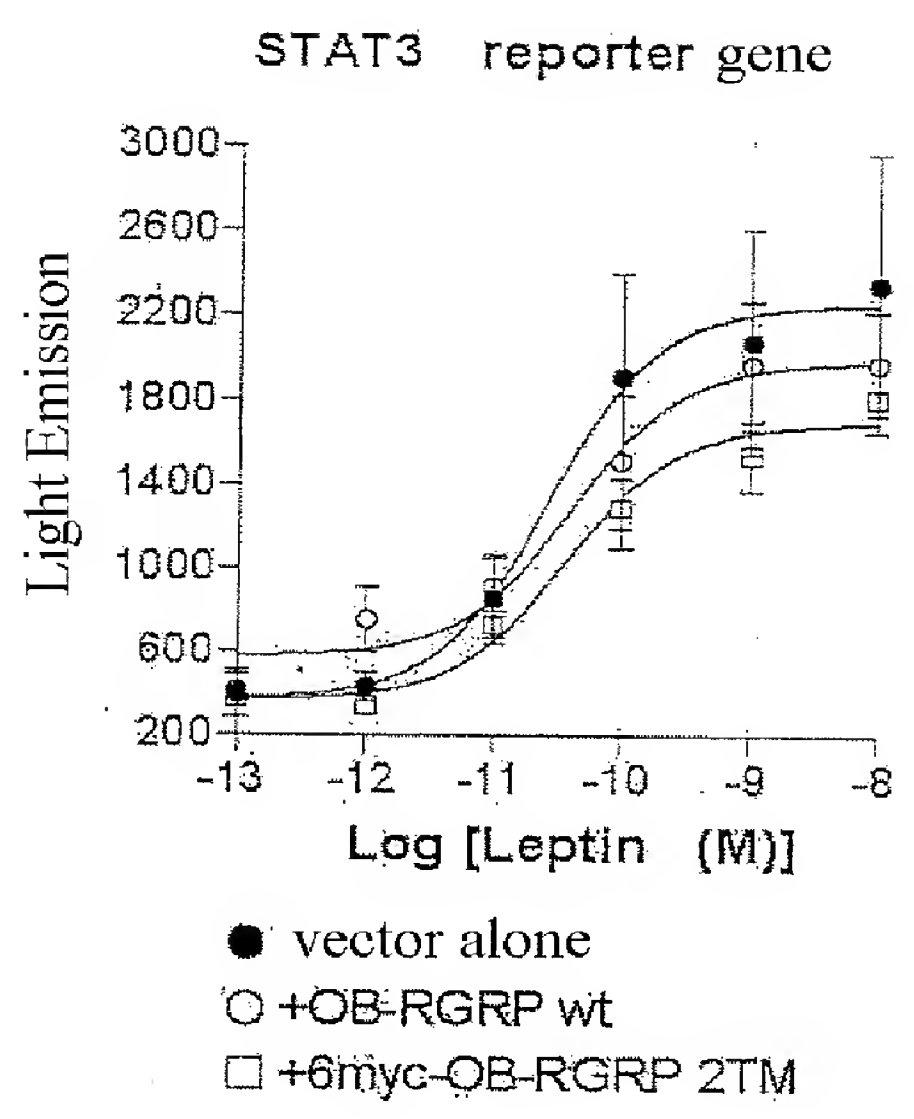


Figure 8b

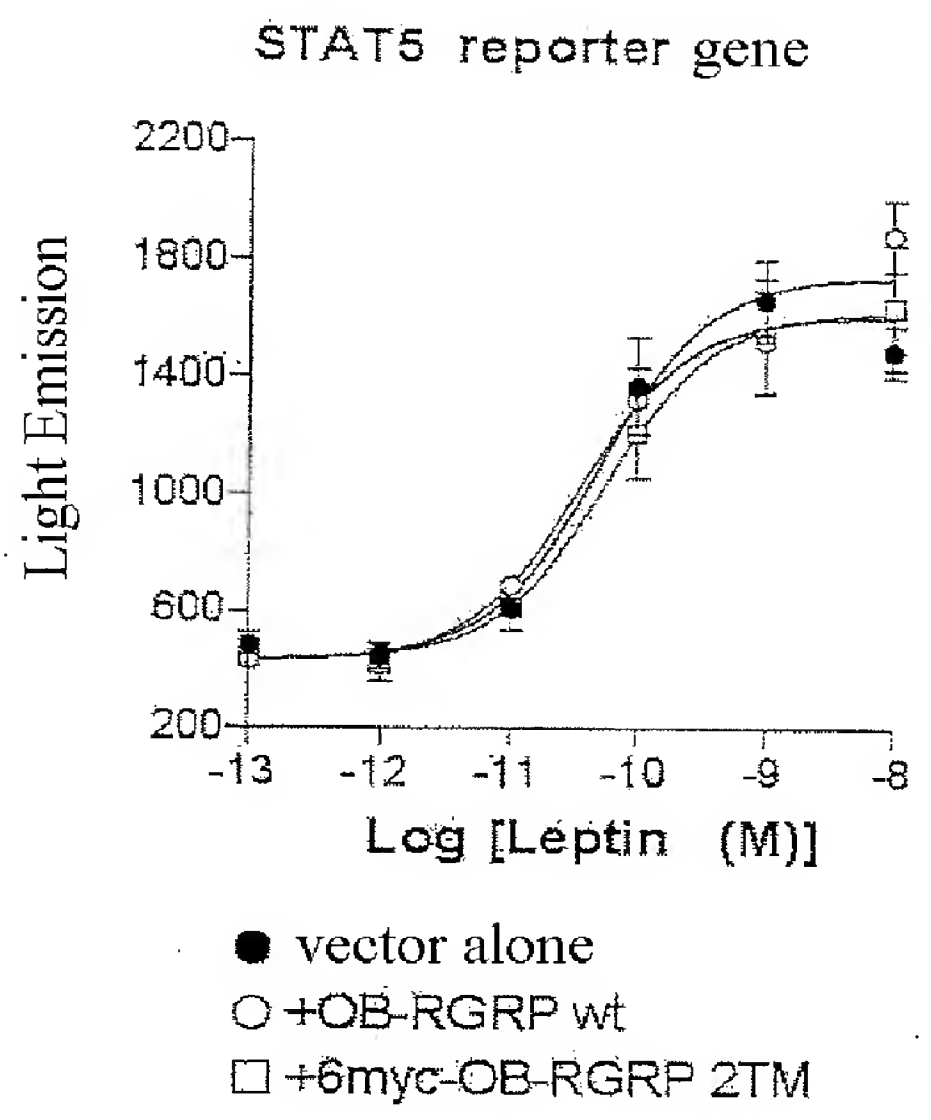


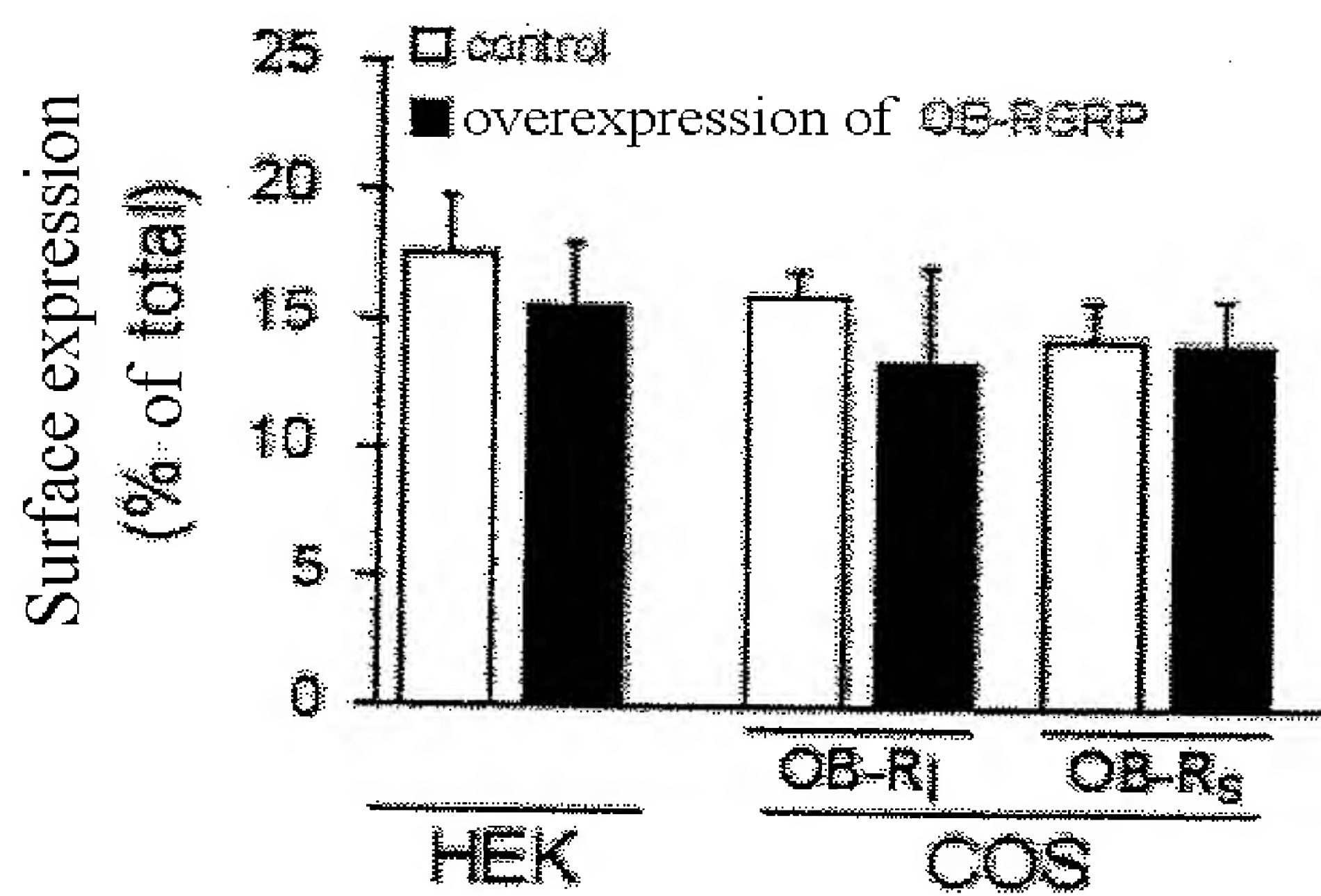
Figure 9

Figure 10a

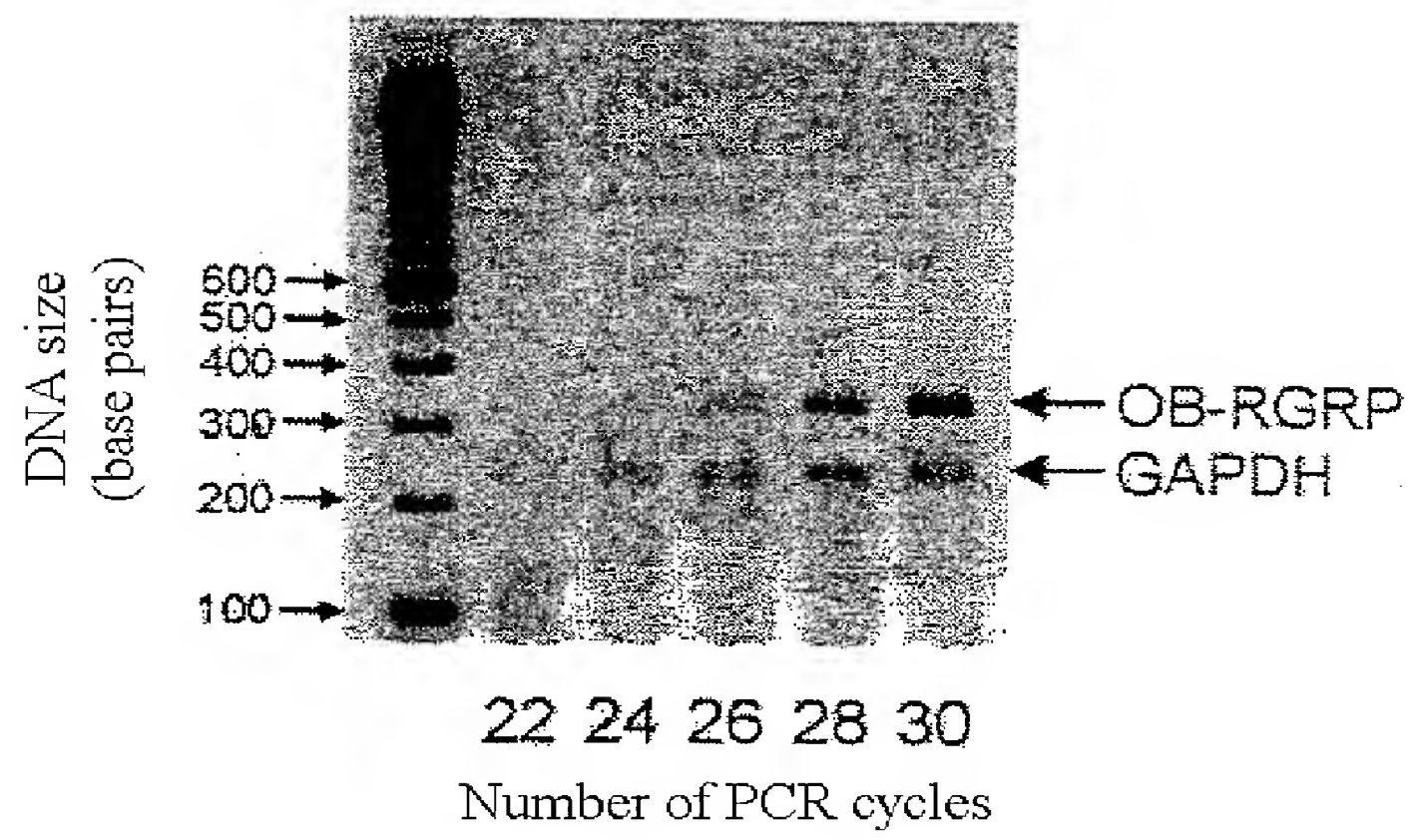


Figure 10b

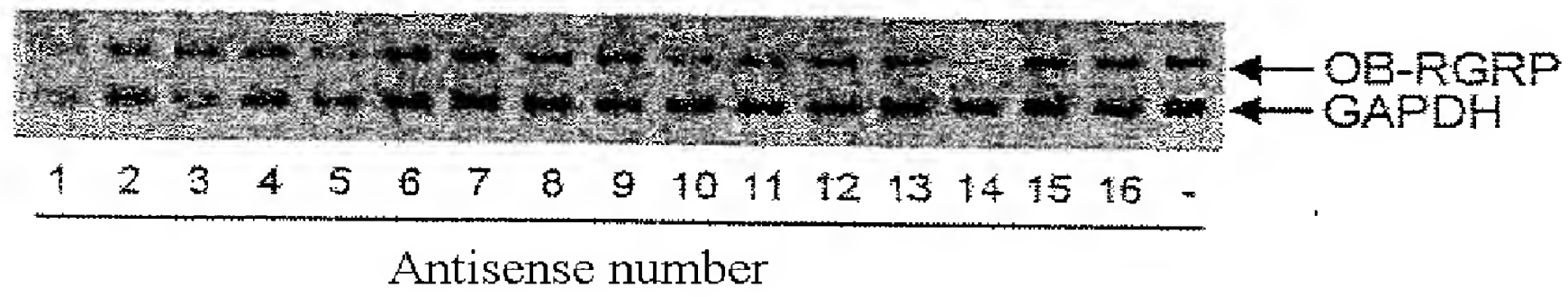
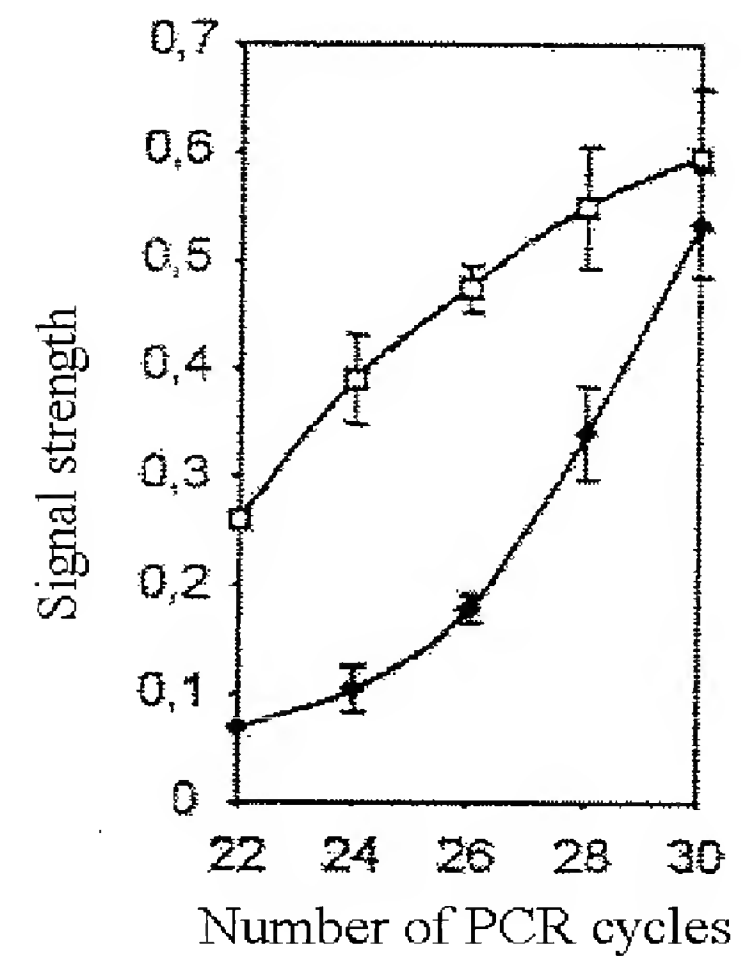


Figure 10c

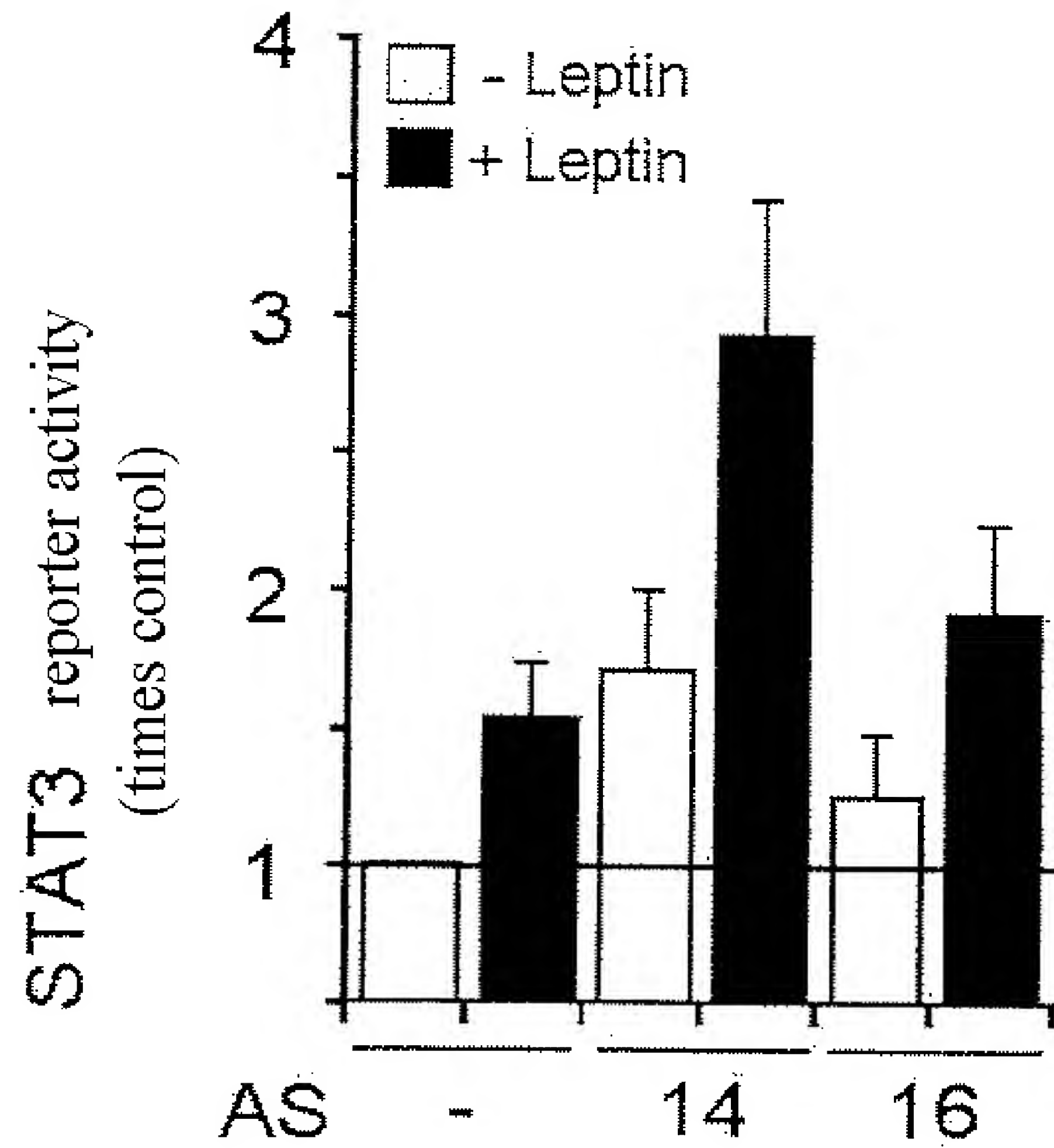
Figure 11

Figure 12

